

M2D1: Prepare cells for RNA purification

03/08/2018

1. Prelab discussion
2. $\frac{1}{2}$ class to TC to seed cells for RNA purification
3. $\frac{1}{2}$ group paper discussion of Dietlein et al.

Office hours

Noreen

- M 2-5pm
- in 16-317

Leslie

- W and F 4-5pm
- in 56-341c or lab

Josephine

- T 4-5pm
- R 10-11am
- in 56-341c or lab

+ Extra

Saturday, 03/10

- 10am-5pm
- in 56-302

Please email us if you can't make office hours and we will schedule a time to meet!

Sign up for journal club

- Pick 1 of 24 papers, or suggest your own
- Present M2D6 (April 3rd) or M2D7 (April 5th)
- Sign up by adding your name next to paper [LMM/TR/Color]

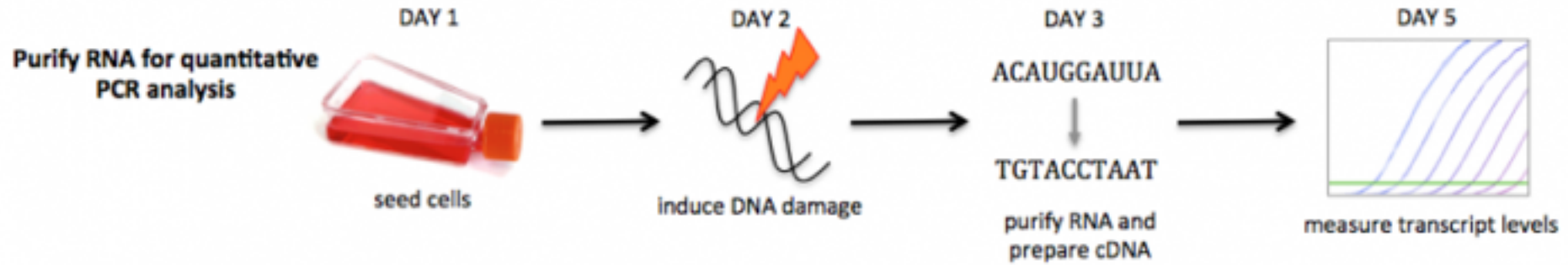
– first come first serve!

– you **cannot** switch paper after M2D⁴~~3~~ (March ~~15th~~) *March 20th*

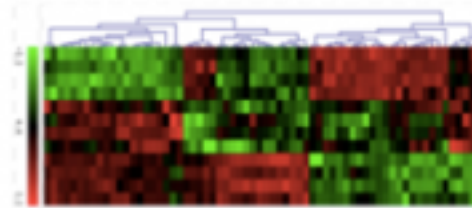
– only one T/R presenter and one W/F presenter per article

Slot	Day 6 (T/R)	Day 7 (T/R)	Day 6 (W/F)	Day 7 (W/F)
1	<input type="text"/>			
2				
3				
4				
5				
6				
7				

M2: Experimental overview



Analyze RNA-seq results

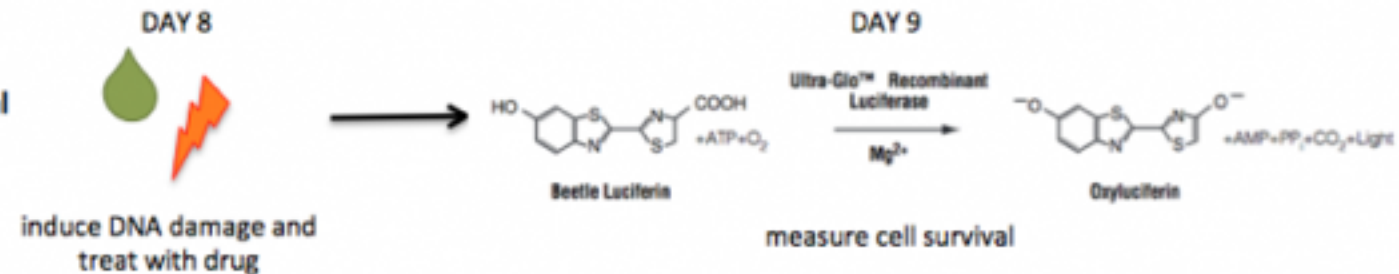


DAY 4: Evaluate altered gene expression

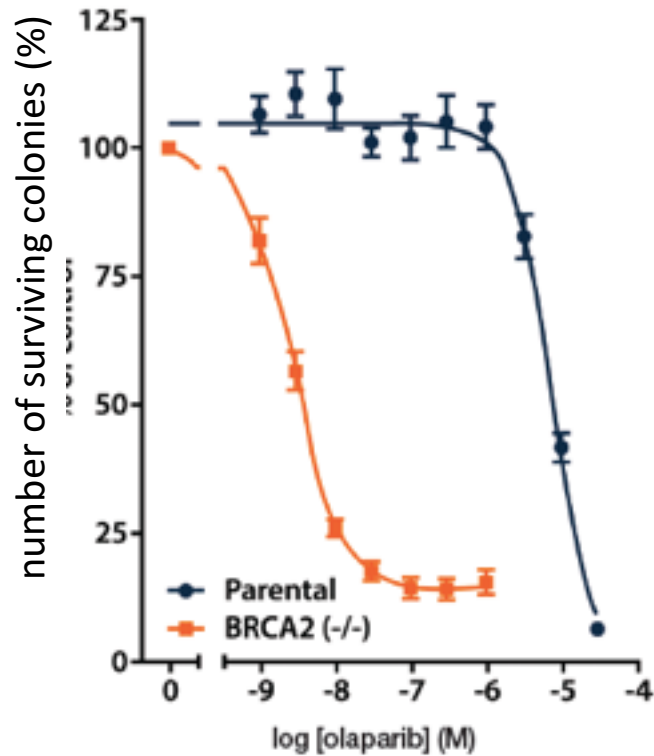
DAY 5: Investigate public databases

DAY 8: Identify regulatory motifs

Examine effect of drug treatment on cell survival



Our cell lines: DLD-1 and BRCA2-/-



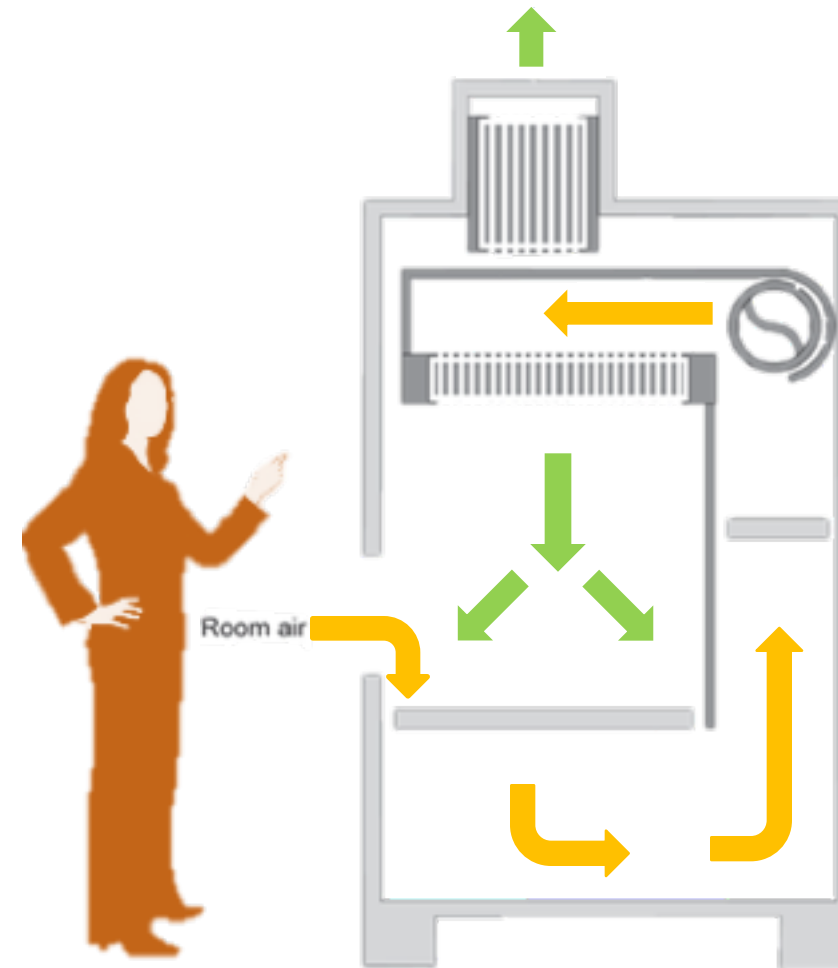
Note: olaparib is a PARP inhibitor (chemotherapy)

- DLD-1 = ~~wild-type~~ (or parental)
 - from the colon of a male with colorectal adenocarcinoma
- BRCA2-/- = mutant
 - disruption of exon 11 from BRCA2 gene
 - deficient in DNA repair (by homologous recombination)

“HR”

Tissue culture sterile technique

- **70% ethanol** is your BFF:
 - wipe cabinet before and after use
 - wipe everything that enters the cabinet
- Do not disturb air flow:
 - Do not block grille or slots
 - Minimize side-to-side arm movements
 - Work > 6" away from sash
 - Leave blower *on*
- Do not talk into incubator!
- Only open sterile items in hood



Mammalian cell culture medium

What do cells need to survive? *growth/division/viability*



- Defined
 - RPMI 1640 (Roswell Park Memorial Institute)
 - (a lot of phosphate)
 - often used to culture lymphoid cells

• Salts

• glucose

• amino acids

• vitamins

• phenol red: pH ~~acid~~ indicator



- FBS: fetal bovine serum

UN defined

• growth factors

• cytokines

• lipids

• cholesterol

defined

- antibiotics:

- penicillin

- streptomycin

] prevent bacterial growth



Mammalian cell culture terminology

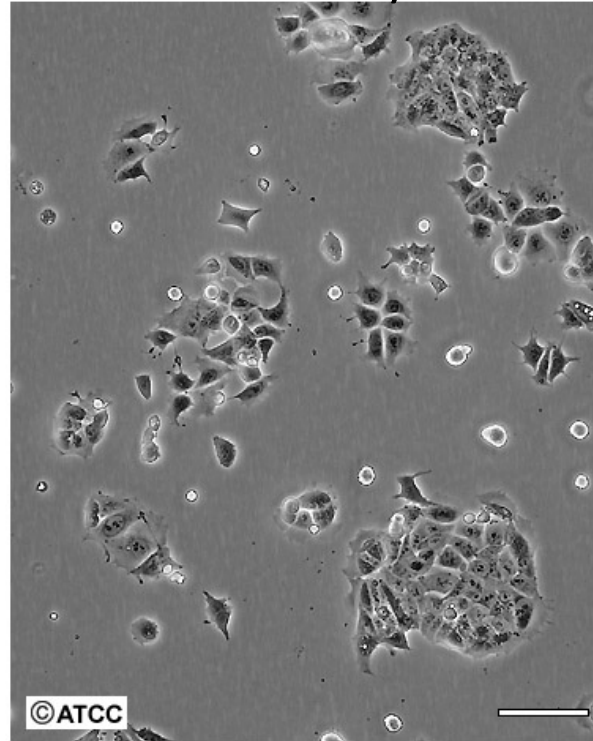
- confluence = density at ~80%. split cells

- splitting = subculturing

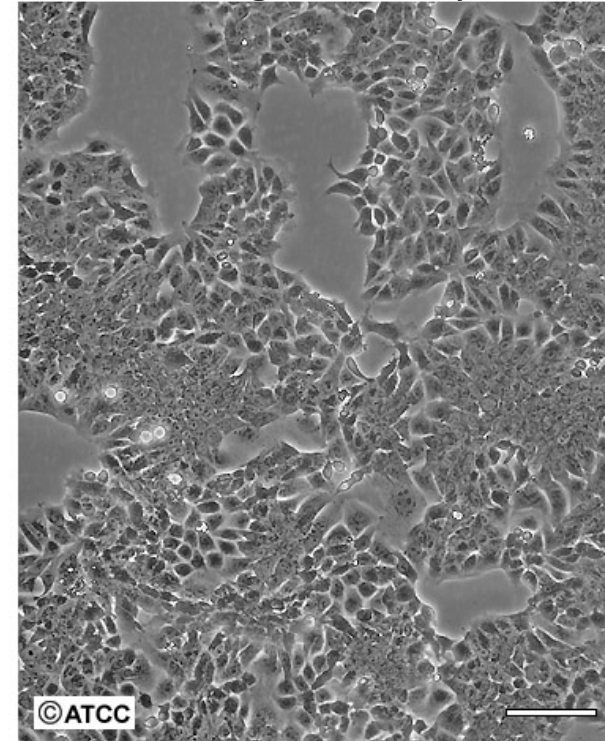
- seeding = ~20-40% of a confluent culture on new dish

DLD-1
↙ ↘

Low Density



High Density

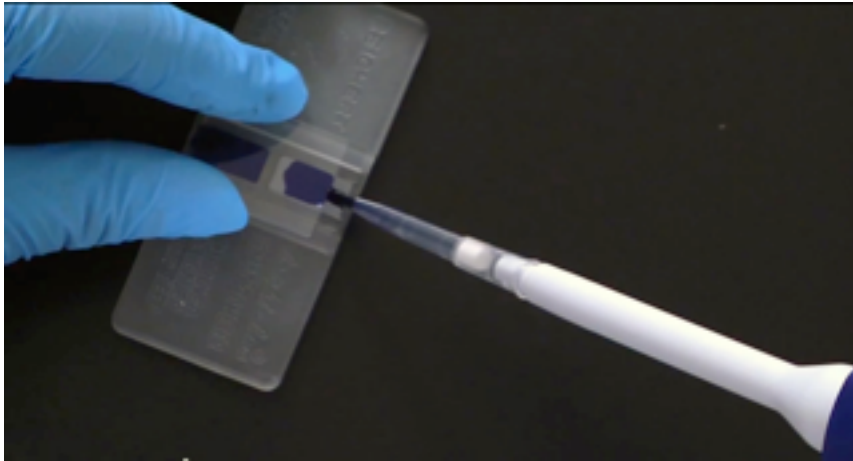


General steps for splitting cells +WHY?

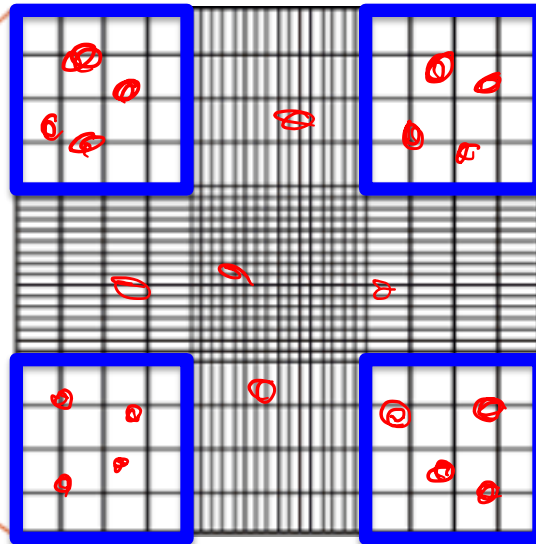
1. Look at cells, estimate confluence
estimate growth/viability
2. Rinse with PBS
wash debris/anti trypsin agents/serum (FBS)
3. Detach cells with trypsin
breaks substrate + cell adhesions
4. Count cells
seed specific # in new vessel
5. "Seed" new culture vessel
room to divide/grow



Calculating number of cells



- Hemacytometer 10ul cell suspension
- Trypan blue - stains dead cells
- # cells / mL = 10,000 x average of 4 corners



$$\frac{16}{4} = 4 \times 10,000 \text{ cells/mL} =$$

$$40,000 \text{ cells/mL}$$

Today in lab:

1. Tissue Culture (TC)

- 1st: Red, Orange, Yellow, & ~~Green~~ ^{Blue}
 - 2nd: ~~Blue~~ ^{Green}, Pink, Purple, White, & Grey
- Protocols printed for TC use, no need to move laptops etc.
- Do not wear PPE in or out of TC room

2. Paper discussion of Dietlein *et al.*

- Homework due ~~Wednesday~~ ^{Tuesday}, M2D2
 - Sign up for journal club day (and article → wait till next week)
 - Create a single slide from Dietlein *et al.*
- Don't forget about Mod1 assignments!