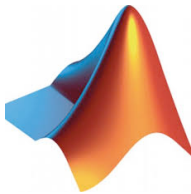


M2D7: Assess protein function

11/04/2015

note: no class nor lab on 11/10 - 11/11 😊

Today in lab



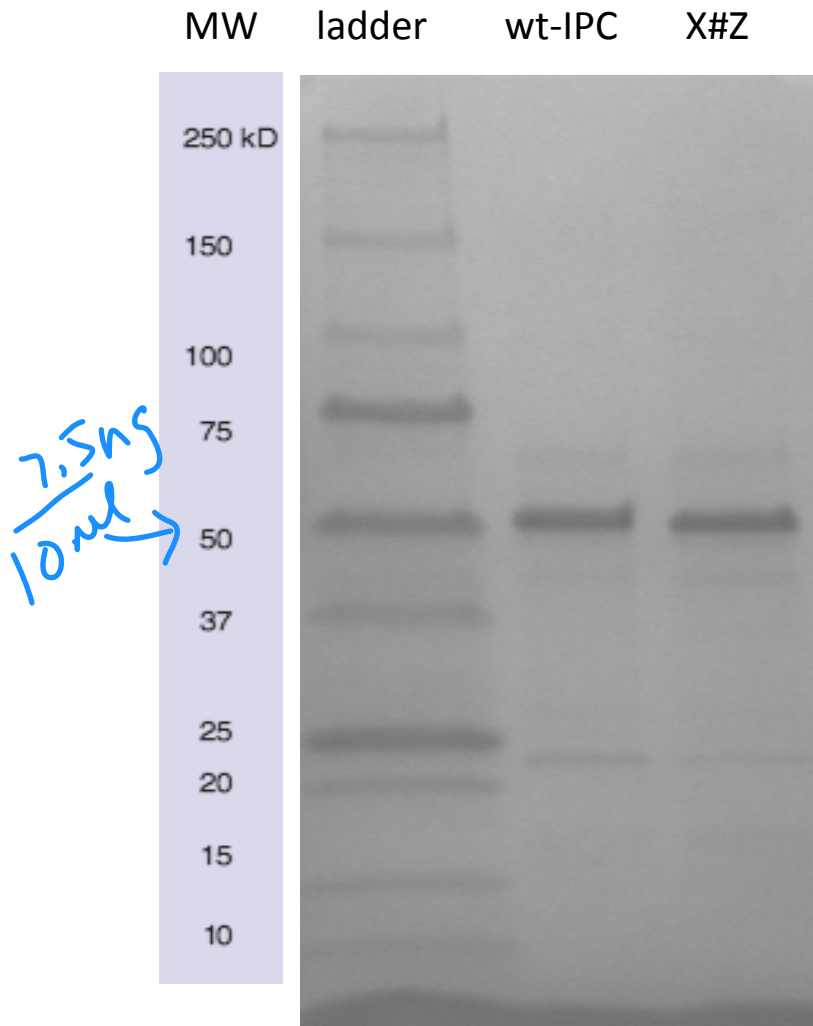
- Quiz 4 ✓
- Lab notebook graded: M2D3 *10pm*
andreakw@mit.edu
- Analyze data with Excel
- Analyze data with MATLAB
- Ask questions
 - about M2D7
 - about all of M2

A few more assignments for M2



- Methods (due M2D6)
 - you did really well!
 - handed back (with peer's feedback) on M2D8
- Protein engineering research report
 - due by 5pm on ~~Saturday~~ ^{Sunday}, November 14 ~~14~~ 15
 - extra office hours, come with specific questions
 - to 56-302 on Wed., Nov. 11th, 10am-4pm
 - to 16-239 on Thu. and Fri., Nov. 12th-13th, 10-11am
 - to 16-317 on Thu. and Fri., Nov. 12th-13th, 6-8pm
 - to 16-429b on Fri., Nov. 13th, 10am-12pm
- Blog post ^{8p M}
 - due by 5pm on Sunday, November 15
 - write about M2, journal club, etc...

Estimate protein concentration using Bradford assay or unstained ladder



- Bradford assay extrapolation from BSA calibration curve
 - don't forget units!
 - mg/mL
- Or comparison with unstained ladder
 - 50 kDa band has 7.5 ng / 10 μ L
 - your wt-IPC is 2 x as bright
 - hence [wt-IPC] = 15 ng / 18 μ L
 - and [IPC(X#Z)] = 15 ng / 18 μ L
 - Convert to M (mol/L) using 1 Da = 1 g/mol

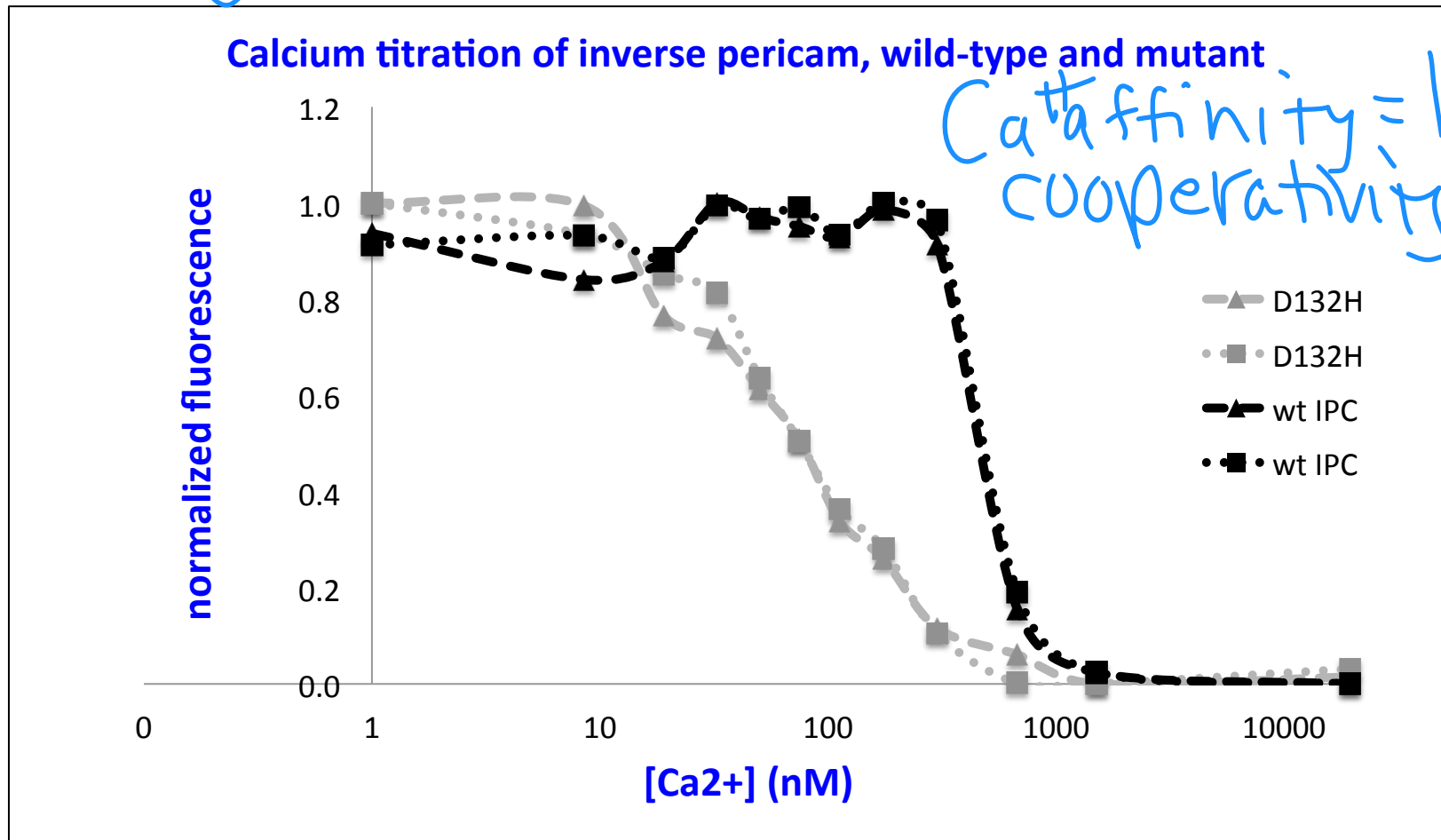
Plot your IPC-calcium titration data in Excel

- Normalize data (or average of 2 data sets):

0 → 1

$$S = \frac{F - F_{\min}}{F_{\max} - F_{\min}}$$

min, = 1 row
max



y = fraction of protein bound to ligand

P = protein

L = ligand

Would anyone like to go through the derivation of

$$K_D = \frac{[L][P]}{[LP]}$$

$$Y = \frac{L}{K_d + L} ?$$

$$Y = \frac{[PL]}{[PL] + [P]}$$

$$[LP] = \frac{[L][P]}{K_D}$$

$$\frac{[L][P]/K_D}{[P] + [L][P]/K_D} = \frac{[L]}{K_D + [L]}$$

$$\begin{aligned} &= \frac{[L][P]}{K_D + [L][P]} \\ &= \frac{[L]}{K_D + [L]} \end{aligned}$$

Analyze data further in MATLAB

vectors of 12 values: L = [calcium]
enter in uM
for 0, enter 0.001
for 1, enter 0.999

1. Enter your data:

- L = [ligand] = [Ca²⁺] in μM
- S_wt: signal wild-type IPC
- m1 is *your* mutant, m2 is another team's

2. logspace (a, b, N) [calcium] range from ~1 nM to 20 uM

- generates a row vector of N logarithmically equally spaced points between decades 10^a and 10^b .
- choose a = -3 , b = 2 , and N = 10,000

3. A ./ B

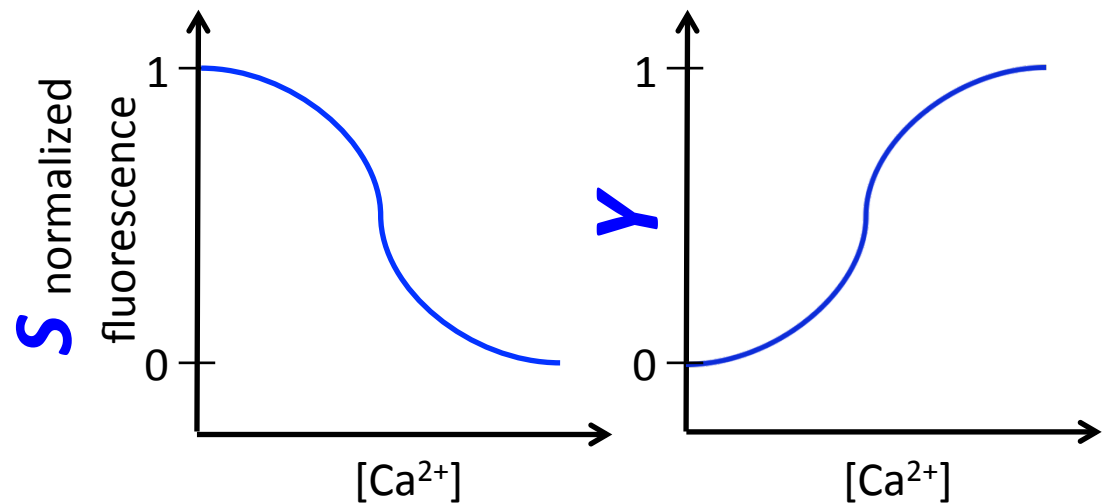
- divides element by element

$$\begin{bmatrix} 2 & 4 & 6 \\ 3 & 6 & 9 \\ 4 & 8 & 12 \end{bmatrix} ./ \begin{bmatrix} 2 & 2 & 2 \\ 3 & 3 & 3 \\ 4 & 4 & 4 \end{bmatrix} = \begin{bmatrix} 1 & 2 & 3 \\ 1 & 2 & 3 \\ 1 & 2 & 3 \end{bmatrix}$$

MATLAB code analyzes data along 3 models

- As in lecture, convenient to go back to *pericam* formalism in equations:

$$Y = 1 - S$$



Part 1: fit apparent K_d

$$Y = \frac{L}{K_d + L}$$

Part 2: fit K_d and n

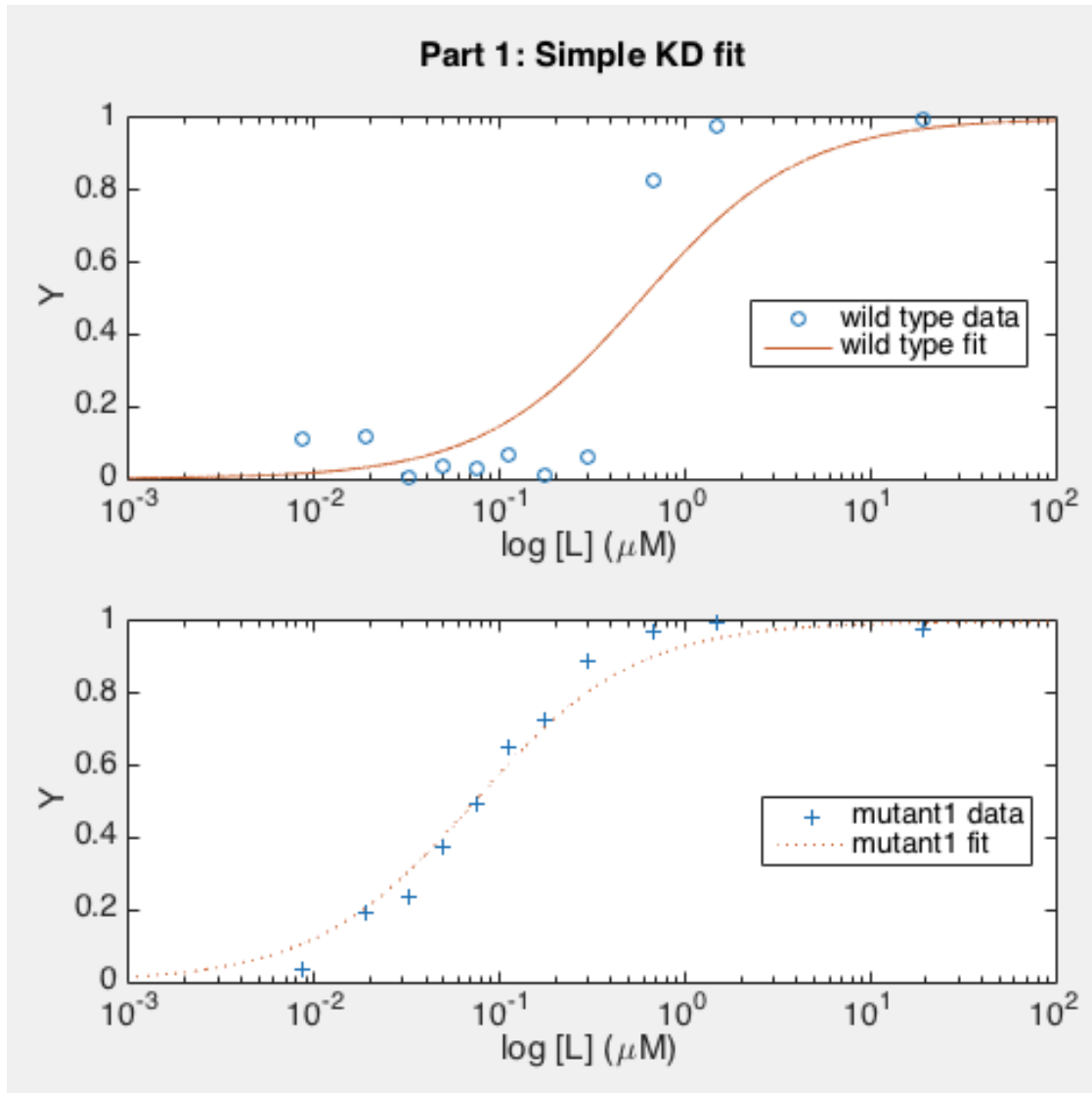
$$Y = \frac{L^n}{K_d^n + L^n}$$

Part 3: fit K_d and n
Hill analysis

$$\log\left(\frac{Y}{1-Y}\right) = n \log(L) - n \log(K_d)$$

Part 1: fit apparent K_d

$$Y = \frac{L}{K_d + L}$$



KD1_wt = 0.5858 μM

KD1_m1 = 0.0729 μM

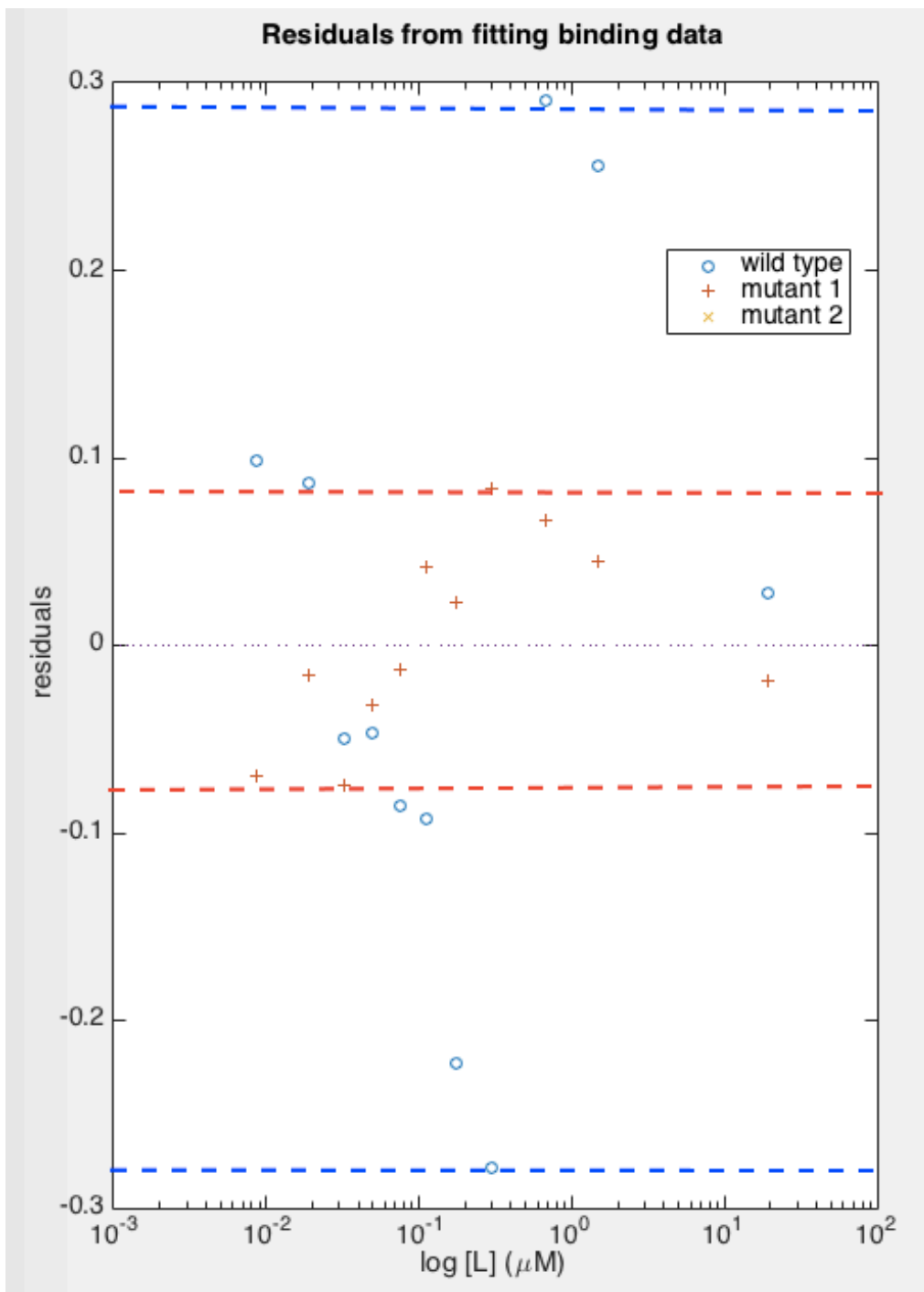
- How good is the fit?
 - for wt-IPC?
 - for mutant?

this model confirms that the mutant has a higher affinity for calcium than wt-IPC

poor fit, especially for wt-IPC

Part 1

$$Y = \frac{L}{K_d + L}$$



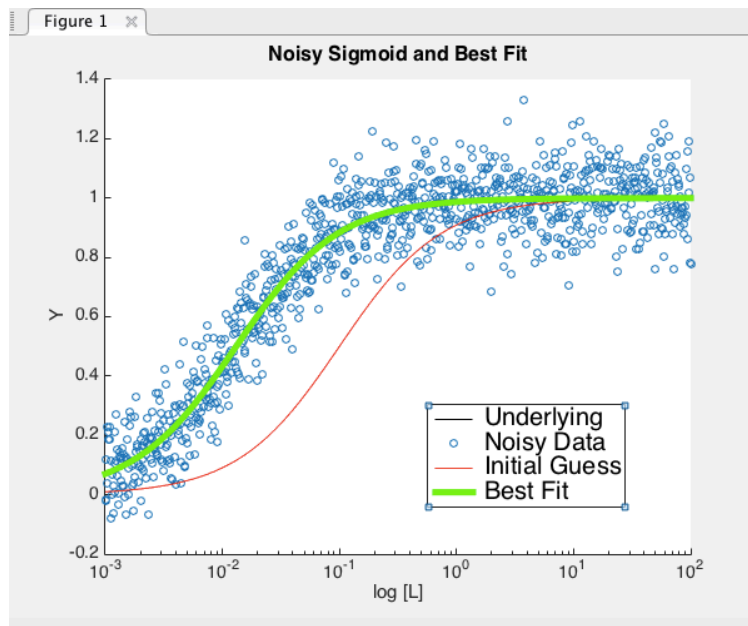
- How good is the fit?
 - for wt-IPC?
 - for mutant?

➤ Quantify *residuals*:
distribution and amplitude

$$\text{residuals} = Y_{\text{experimental}} - Y_{\text{model}}$$

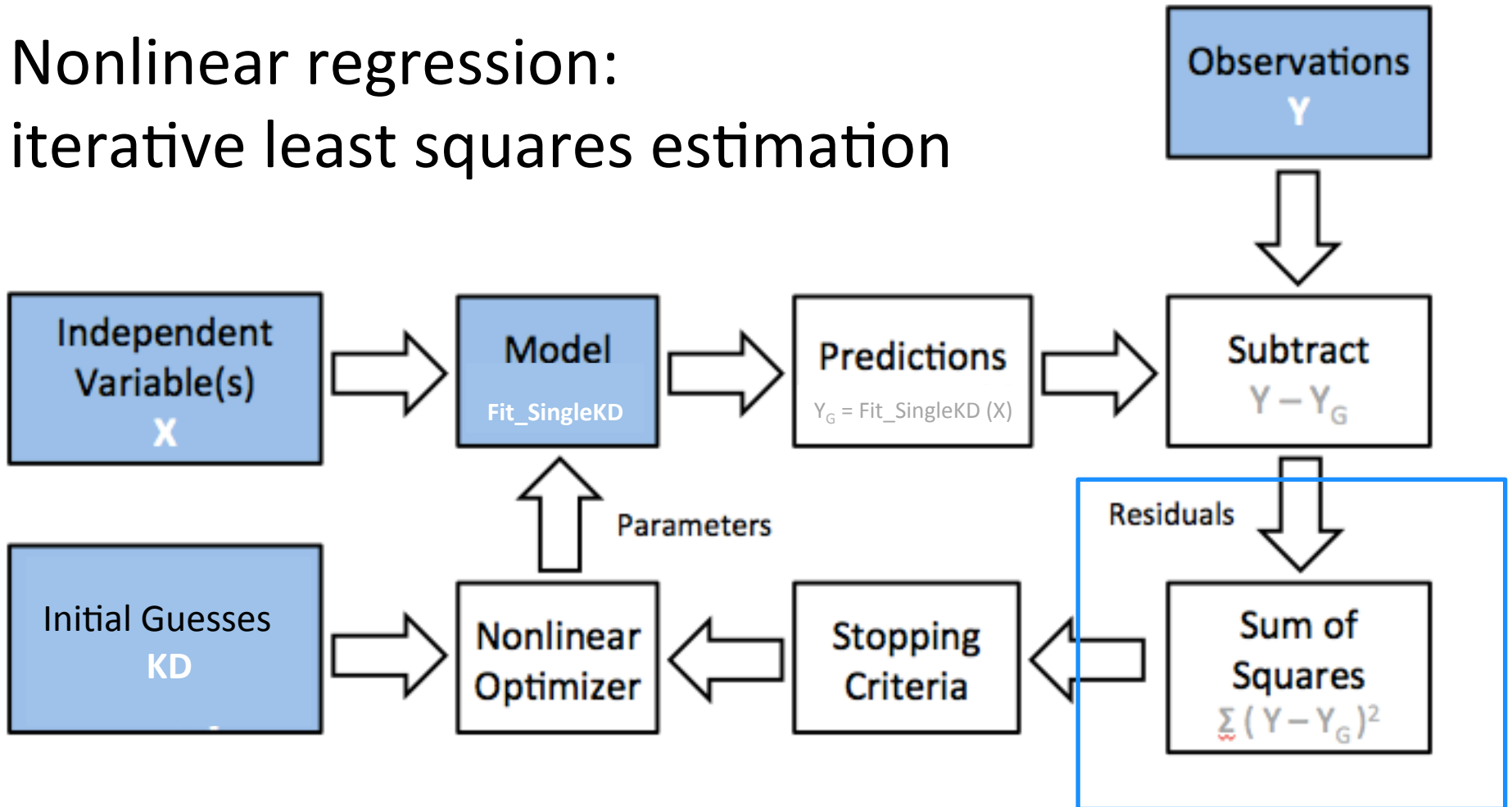
Nonlinear regression is at the core of the MATLAB code

- `nlinfit(X, Y, @model, initialGuess)`
 - X (predictors): calcium concentrations
 - Y (responses): fluorescence signal
 - model: `Fit_SingleKD`
 $x ./ (KD + x);$
 - initialGuess: starting value for `KD`



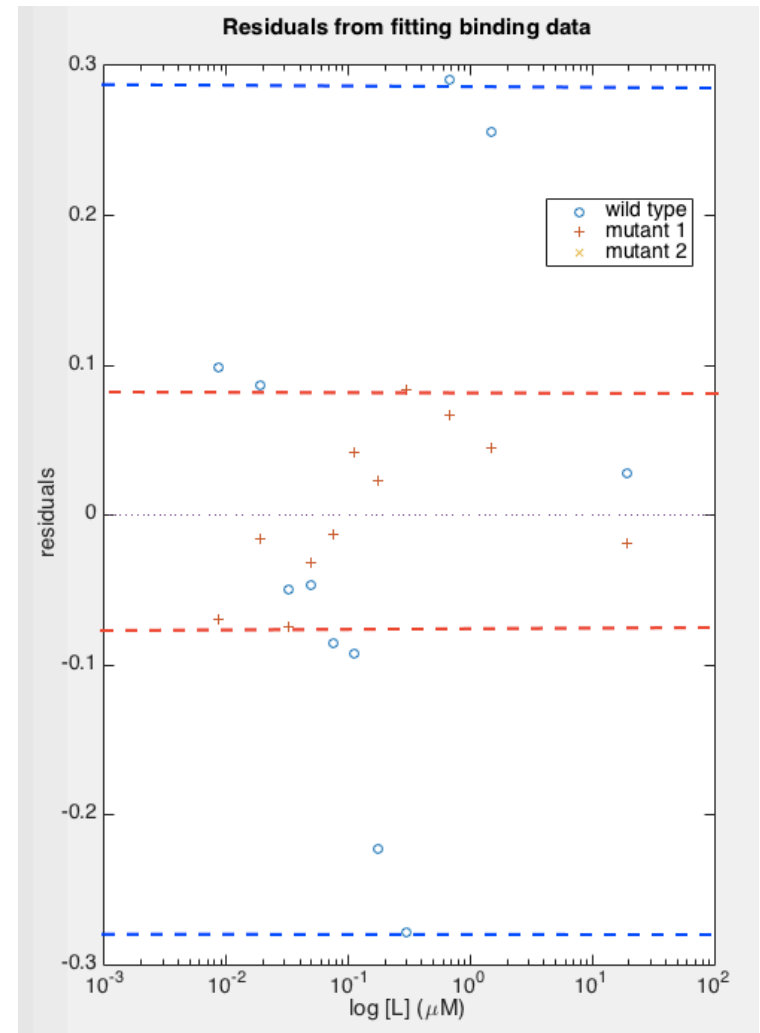
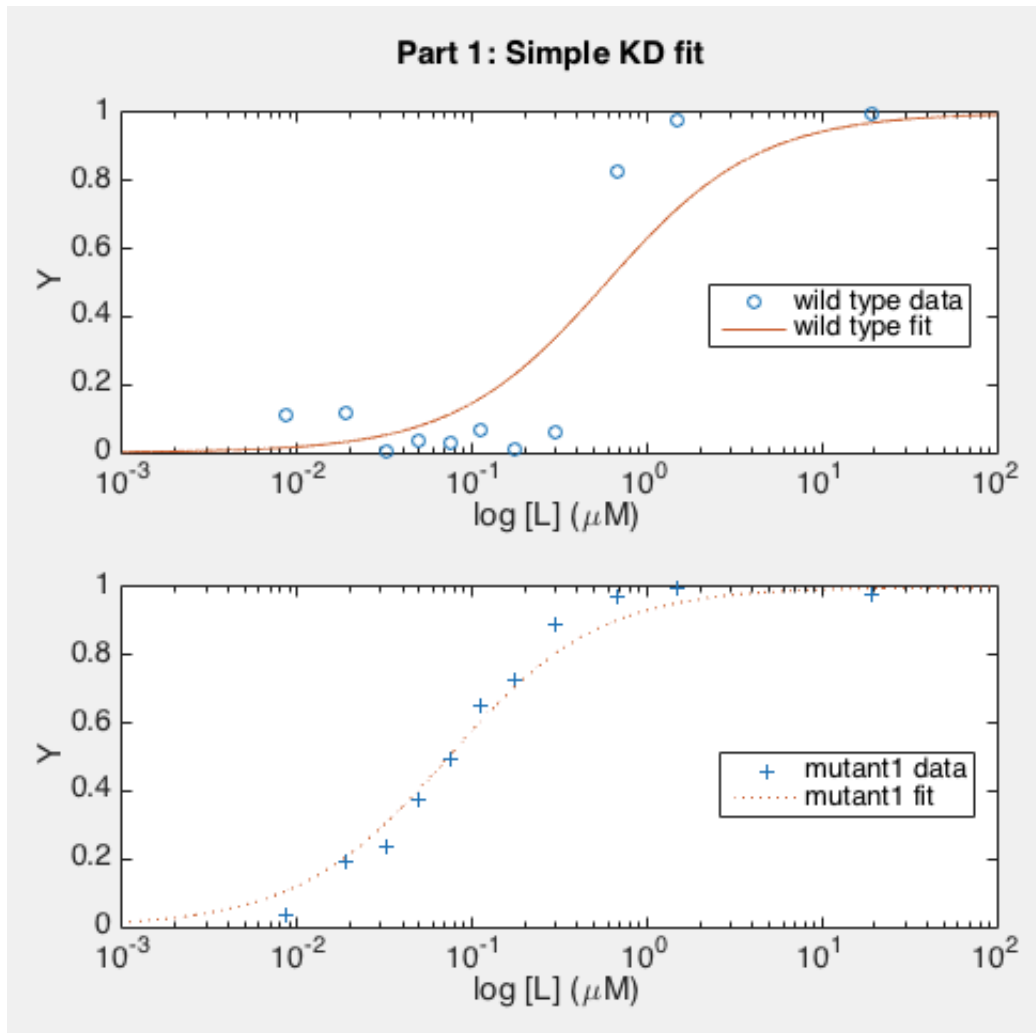
- Find parameters that can explain $Y = \text{model}(\text{Parameters}, X)$ and start your search with `parameters0 = initialGuess`

Nonlinear regression: iterative least squares estimation



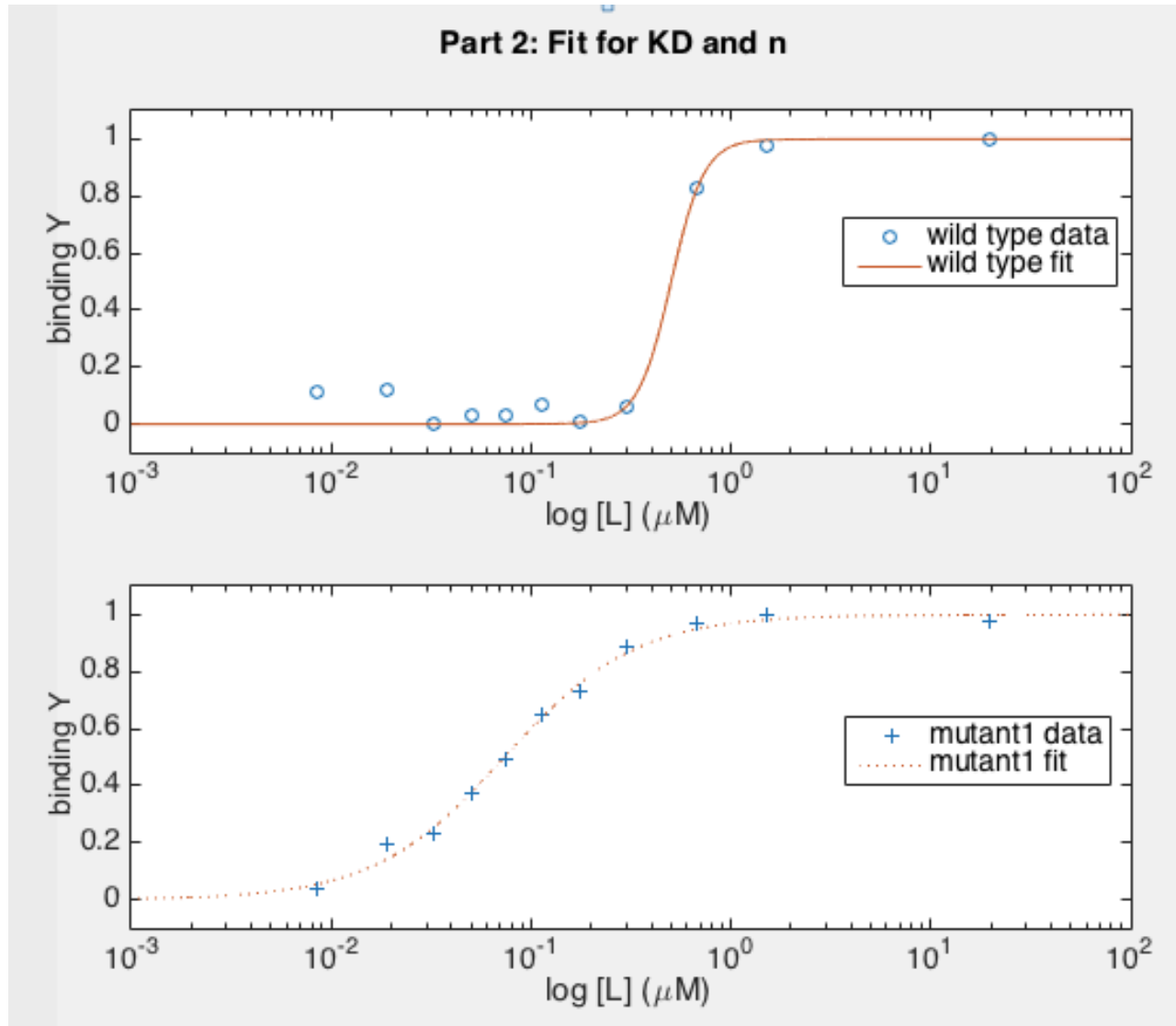
- Optimum reached = changing any of the parameters will result in a higher residual sum of squares.
- Optimizer stops when parameters or sum of squared residuals changes less than tolerance, or when maximum number of iterations reached.

... and this is why residuals $Y - Y_{model}$ provide qualitative and quantitative goodness of fit!



Part 2: fit K_d and n

$$Y = \frac{L^n}{K_d^n + L^n}$$



KD2_wt = 0.5025 μM
n_wt = 5.2508

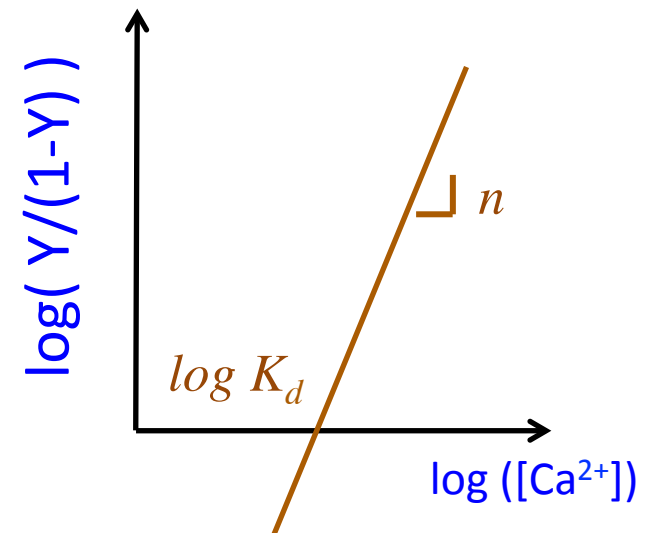
KD2_m1 = 0.0737 μM
n_m1 = 1.3250

- Is the fit any better?

Part 3: fit K_d and n by Hill analysis

- Work only on linear transition region
 - linear fit (polynomial of degree 1)
 - x-intercept = $\log(K_d)$
 - slope = n
- Will need to change indexes in MATLAB algorithm
 - then work with *cell arrays* to parallelize analysis

$$\log\left(\frac{Y}{1-Y}\right) = n \log(L) - n \log(K_d)$$



```
L_wt = L(9:10); Y_wt = Y_wt(9:10); Yp_wt = Y_wt./(1-Y_wt);  
L_m1 = L(2:10); Y_m1 = Y_m1(2:10); Yp_m1 = Y_m1./(1-Y_m1);  
L_m2 = L(6:10); Y_m2 = Y_m2(6:10); Yp_m2 = Y_m2./(1-Y_m2);  
  
% Create cell arrays to concatenate elements of different size:  
L = {L_wt; L_m1; L_m2};  
Y = {Y_wt; Y_m1; Y_m2};  
Yp = {Yp_wt; Yp_m1; Yp_m2};
```

Make a story out of your M2 results

