

# M1D3: Prepare and treat cells for CometChip Experiment

09/19/18

1. Communication workshop 16-220
2. Determine # cells to load
3. Load CometChips with +DNAPKcs and –DNAPKcs cells
4. Induce DNA damage with  $\text{H}_2\text{O}_2$  and lyse cells

## Announcements

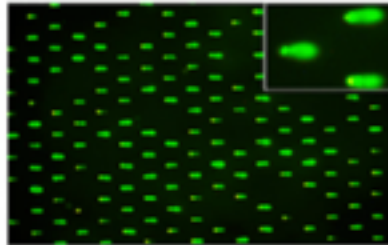
- \*Remember to spray & wipe benches with 70% ethanol before and after work
- \*Remember to empty benchtop buckets at end of day
- \*No lab on Friday!

# Overview of Module 1: Measuring Genomic Instability



## 1. Optimize comet chip assay ✓

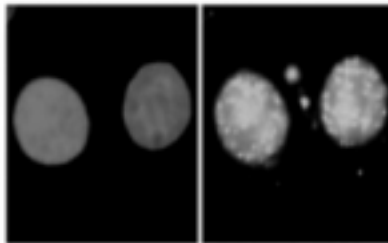
- Test loading variables



## 2. Use comet chip assay to measure DNA damage

- Measure effects of  $H_2O_2$  on +/- DNA-PK cell lines

TODAY

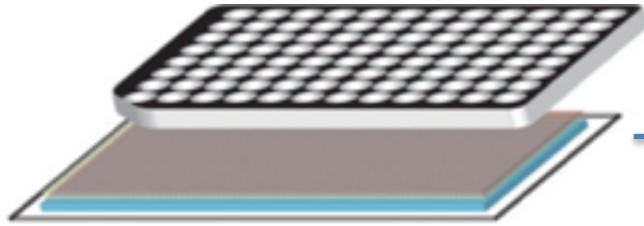


## 3. Use immuno-fluorescence assay to measure DNA damage

- Examine effect of  $H_2O_2$  on  $\gamma$ H2AX foci formation

# Assess DNA damage in tumor cells with & without DNAPKcs

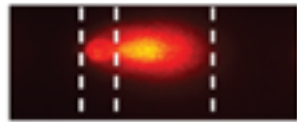
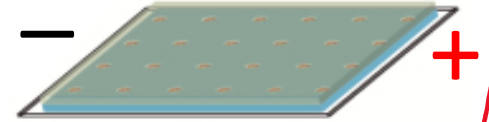
Treat captured cells in comet chip  
with  $\text{H}_2\text{O}_2$  (oxidative damage)



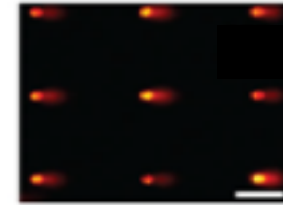
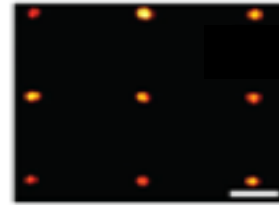
Lyse cells & unwind DNA  
(DNA still captured  
agarose in overlay)

Next week

Agarose Electrophoresis

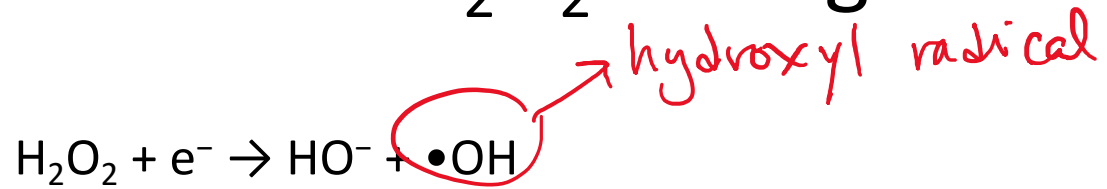


Analysis  
via  
Matlab

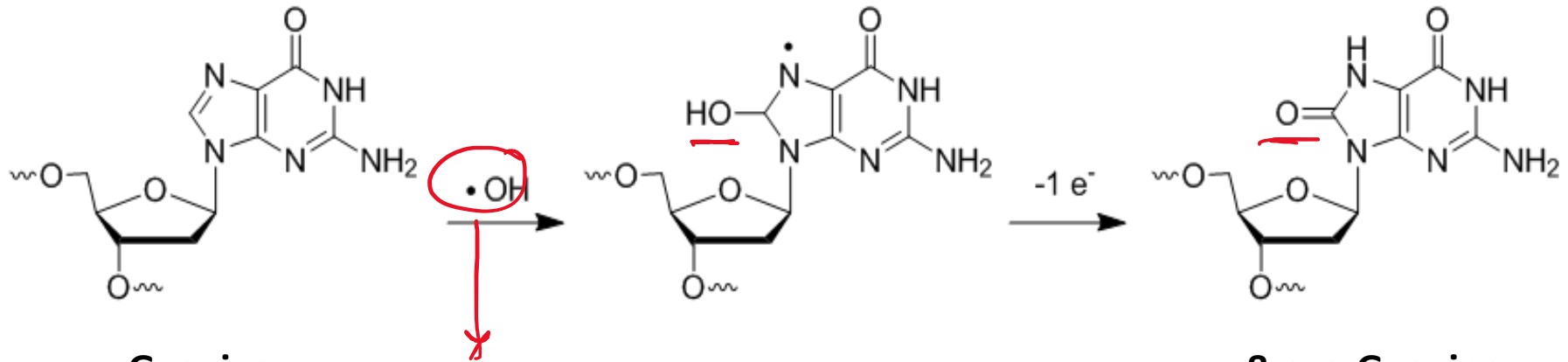
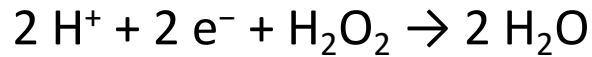


Stain DNA and image via  
fluorescence microscopy

# How does $\text{H}_2\text{O}_2$ damage DNA?



ROS = Reactive oxygen species

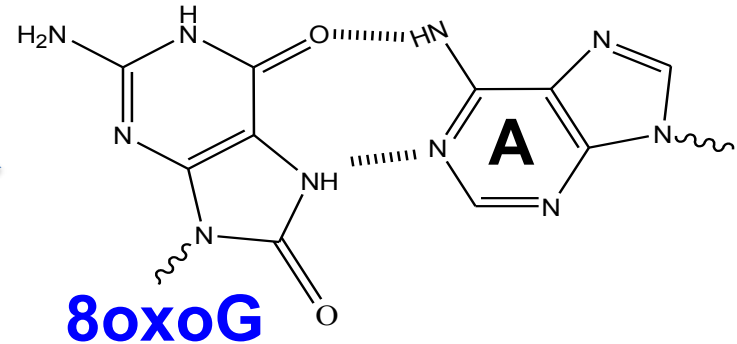
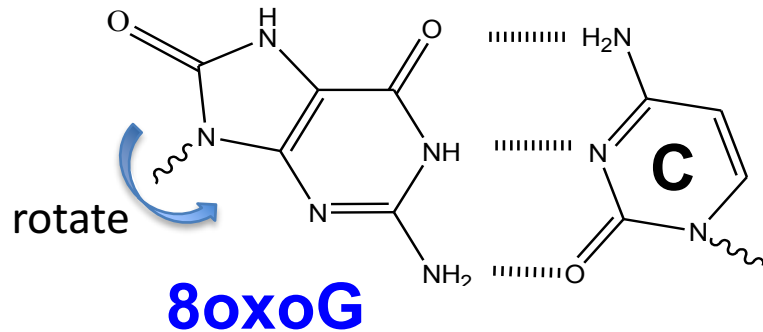
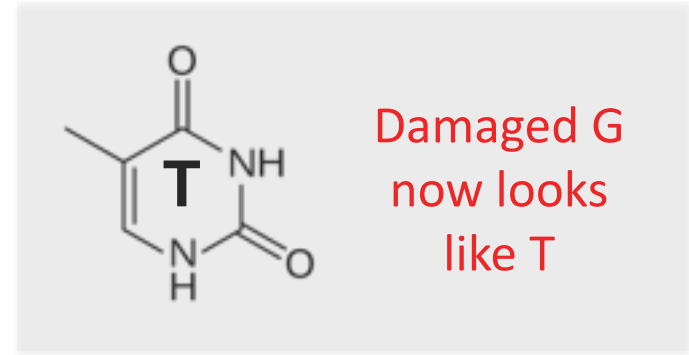
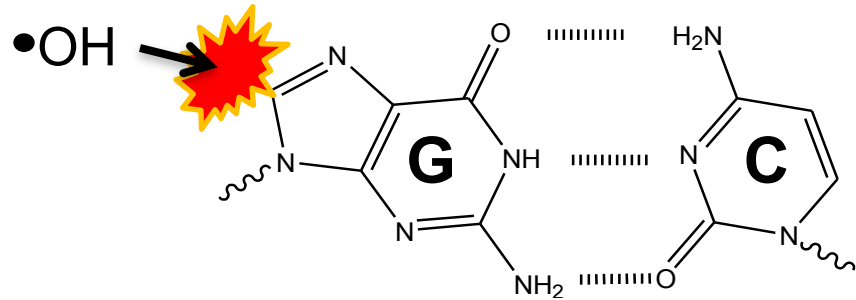


Guanine

8-oxo-Guanine

*hydroxyl radical attacks bond + adds carbonyl group*

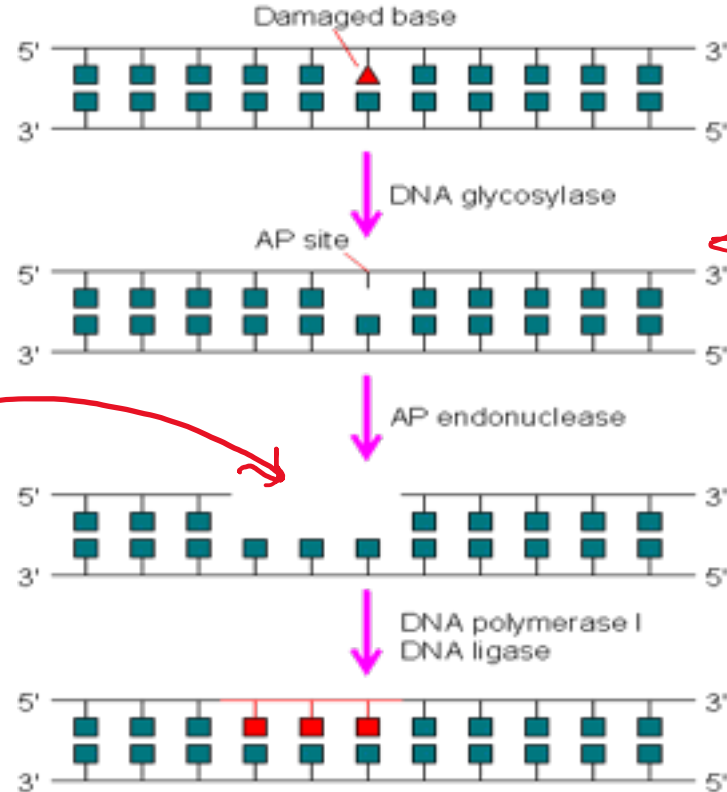
# How does $\text{H}_2\text{O}_2$ damage DNA?



Mutation if replicated GC → AT

# How do our cells respond?

## Typically, Base Excision Repair (BER) Pathway

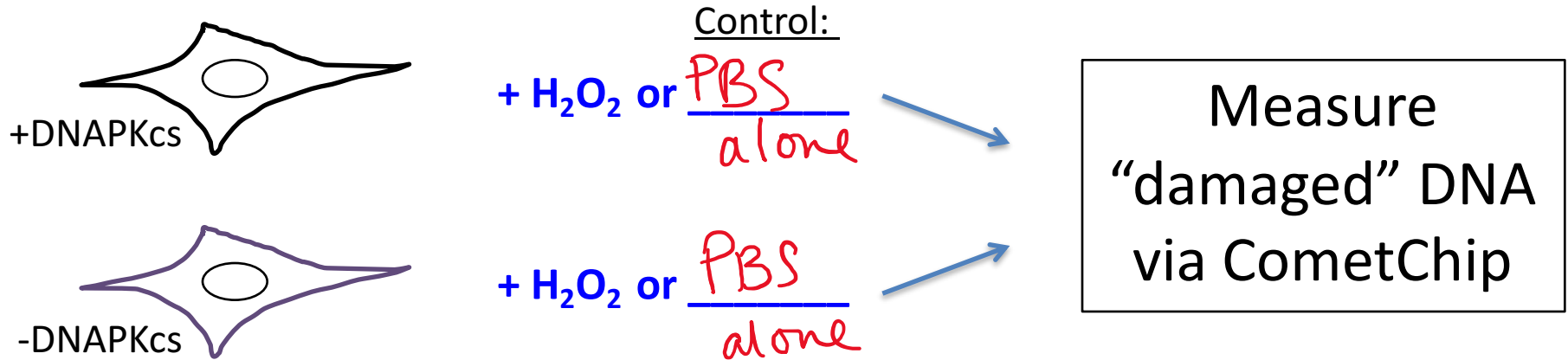


Single  
strand  
breaks  
(SSB)

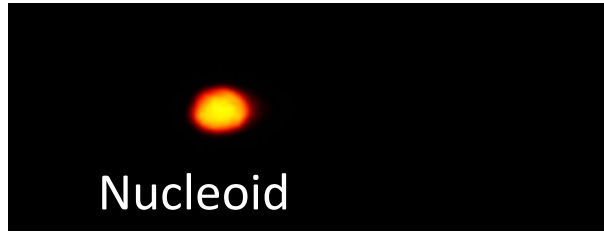
← AP site =  
a basic site  
(apurinic  
apyrimidinic)

# Does DNAPKcs have a role in DNA repair in response to oxidative damage?

- BER can lead to double strand breaks which can be repaired by NHEJ (nonhomologous end joining)
- DNAPKcs = DNA-dependent protein kinase, catalytic subunit
- DNAPKcs involved in NHEJ

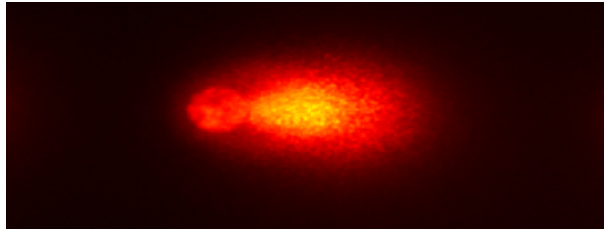


# Output of Alkaline CometChip Assay



## No Damage

- Supercoiled nucleoid
- Little or no migration



## High Damage

- SSBs, abasic sites, alkali labile sites
- forms a "Comet tail"

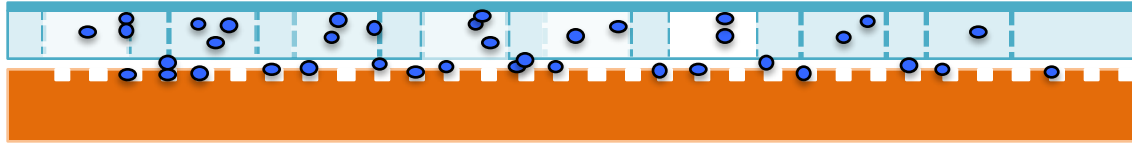
**Genomic damage from direct strand breaks**  
**AND** Repair Intermediates



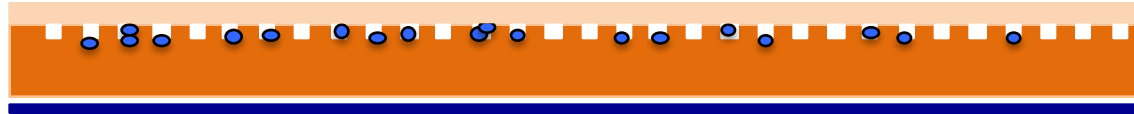
# Logistics of today's experiment

Be sure to mix  
your cells right  
before loading!!!

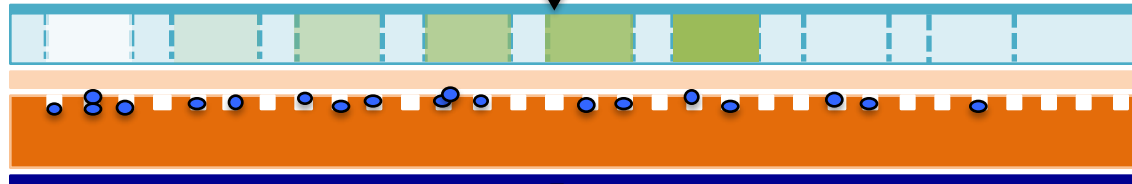
Load      TBD      cells per macrowell



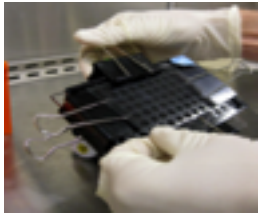
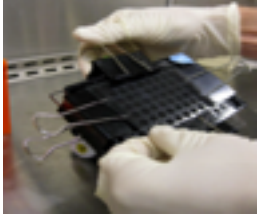
1% LMP agar



Treat with H<sub>2</sub>O<sub>2</sub>

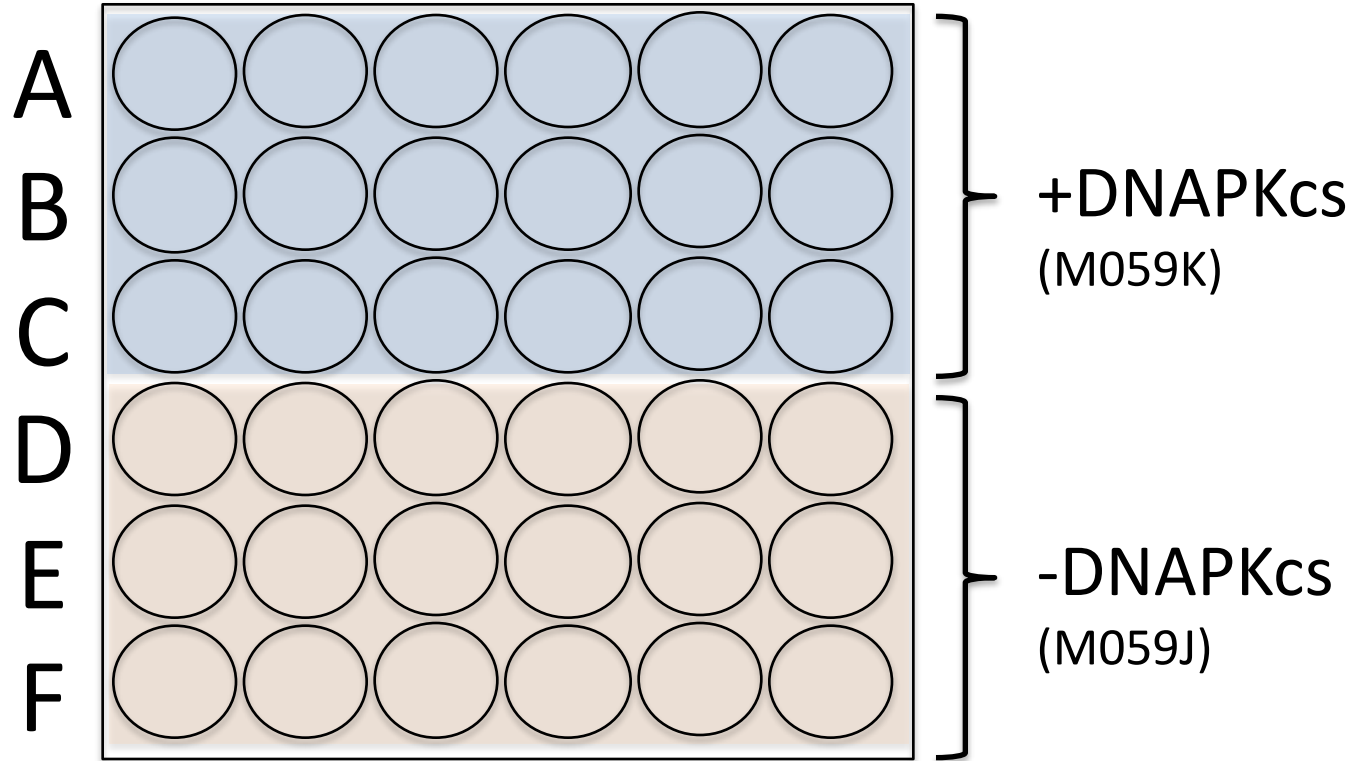


Wash off mutagen (carefully!) and leave CometChip in lysis buffer

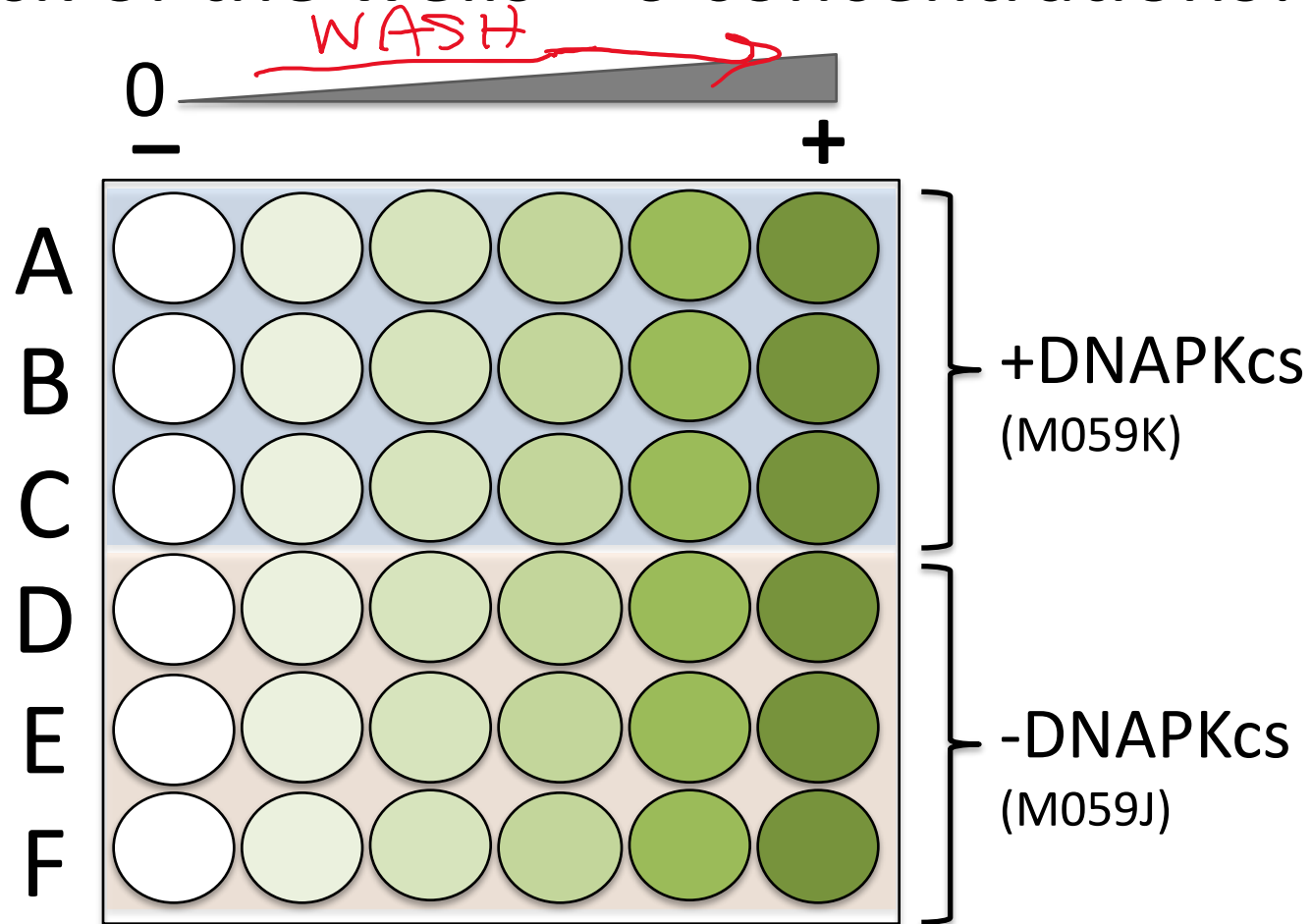


# Keep track of the wells – 2 cell lines!

WASH



# Keep track of the wells – 6 concentrations!



# Preparing H<sub>2</sub>O<sub>2</sub> dilution series:

Treat with: 0, 0.01, 0.1, 0.25, 0.5, 1  $\mu$ M H<sub>2</sub>O<sub>2</sub>

- 4°C for 20min
- Add **100 $\mu$ l** of drug dose to each macrowell
- **Triplicate**: each concentration will have three macrowells for each cell line
  - Make 1 mL of each concentration

**Stock 1: 10 M**

**Stock 2 Provided: 1 mM**

**Stock 3: 1  $\mu$ M**

Need  
2 mL total

1:1000

Note: 2  $\mu$ l  
minimum volume  
on your pipettes

## Handling tips:

- Concentrated H<sub>2</sub>O<sub>2</sub> (10M) should be left at front bench
- Keep H<sub>2</sub>O<sub>2</sub> **on ice** at all times
- Minimize waste!

$$C_1 V_1 = C_2 V_2$$

$$(1 \mu\text{M})(X) = (0.5 \mu\text{M})(1000 \mu\text{l})$$

↳ 500  $\mu$ l of Stock 3  
500  $\mu$ l of PBS

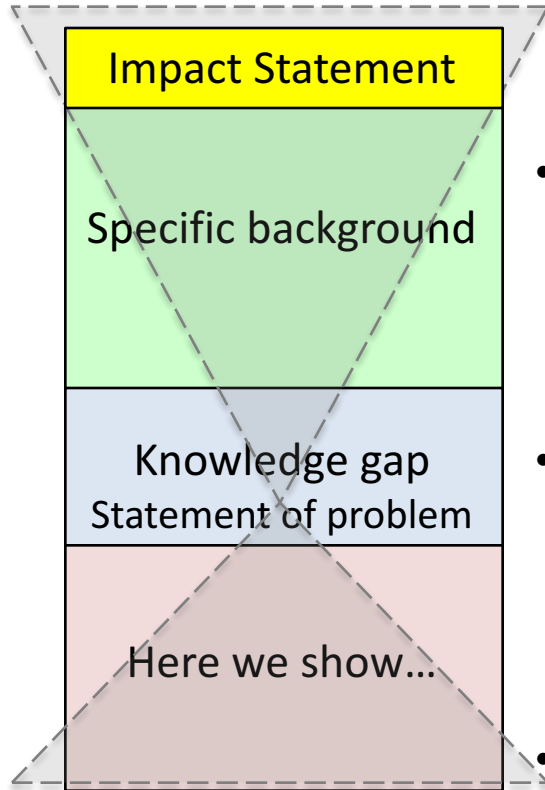
	0	0.01	0.1	0.25	0.5	1 $\mu$ M
Stock 3					500 $\mu$ l	1000 $\mu$ l
PBS					500 $\mu$ l	0

Let's combine our data from last time:

How many cells should we load per microwell? *25,000 cells per macrowell*

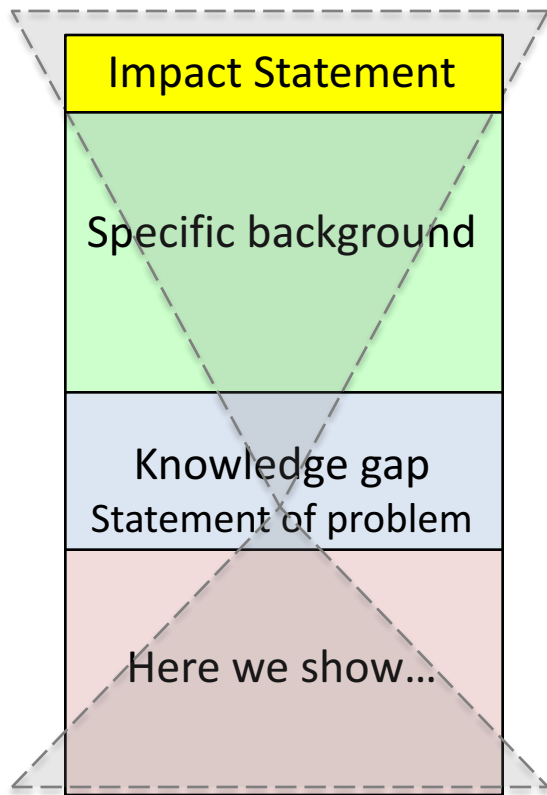
	M059J (-DNAPKcs)		M059K (+DNAPKcs)	
Team	Row B cell # loaded (% filled)	Row C cell # loaded (% filled)	Row B cell # loaded (% filled)	Row C cell # loaded (% filled)
Yellow	50K (32%)	250K (25%)	50K (25%)	250K (35%)
Pink	38K (48%)	100K (40%)	38K (36%)	100K (47%)
Green	25K (13%)	150K (28%)	25K (67%)	150K (62%)
Blue	5K (48%)	50K (40%)	5K (36%)	50K (47%)

# What goes into a background/motivation section?

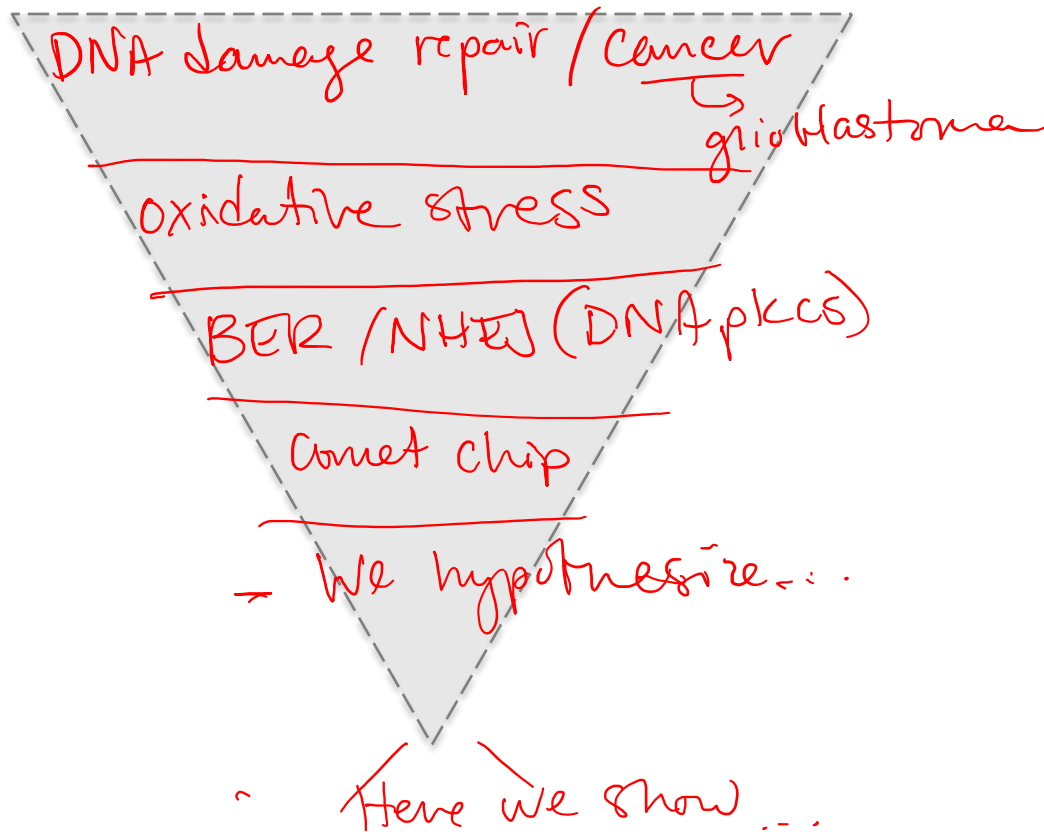


- Your research is anchored in a general topic that your audience cares about or could be interested in.
  - Focus on describing previous work in the field
- Specific background connects your project with the general background.
  - Minimum essential information
  - References current work in the field
  - Introduce specific technologies necessary for understanding the project
- The question you address is clearly articulated, connected to the background, and has appropriate scope for the project
  - Give evidence of incompleteness of current understanding, therefore motivating the investigation
  - Include a space holder for your hypothesis (or come up with one)
- A preview of your findings and their implications
  - Light on Methods

# What goes into your introduction?



*Choose one narrative*



# In lab today

1. Calculate volume of cells to load, obtain cells from instructors and load cells onto CometChip
2. Calculate volumes necessary for mutagen dilutions and check with instructors.
3. Treat CometChip with mutagen and carefully wash.
4. Leave CometChips in lysis buffer.

## M1D4HW (9/26/18)

1. Write topic sentences (1<sup>st</sup> sentence) for each paragraph that would be in your Data Summary's Background and Motivation section
    - Remember to include references with summary & why you chose it
- Reminder: Visit Comm Lab before M1D5
  - First Mod1 Quiz at 1:05pm M1D4