# M2D7: Induce CRISPRi system

- 1. BE Communication workshop
- 2. Pre-lab
- 3. Analyze sequencing results
- 4. Prep for dCas9 induction and mixed-acid fermentation

### Major assignments for M2

- Research Article
  - Due by 10pm on Mon., November 20<sup>th</sup>

#### **Research Article content**

- 1. Title
- 2. Abstract
- 3. Introduction
- 4. Materials and Methods
- 5. Figures and Results
- 6. Discussion
- 7. References

• Blog post for M2 due by 10pm on Tues., Nov. 21st

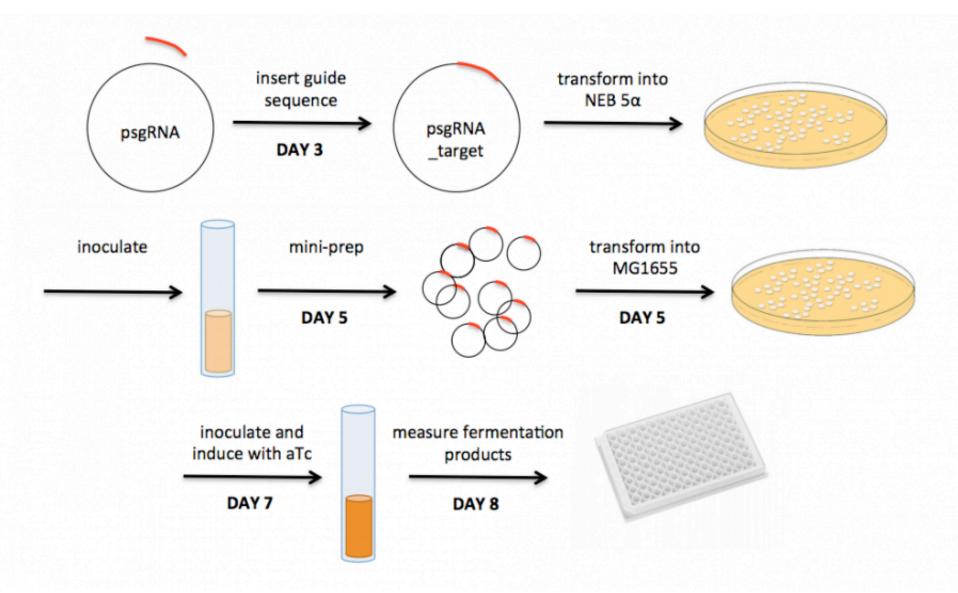
### Extra office hours

- Saturday Nov. 18<sup>th</sup>
  12pm-5pm
- Monday Nov. 20<sup>th</sup> 11am-5pm

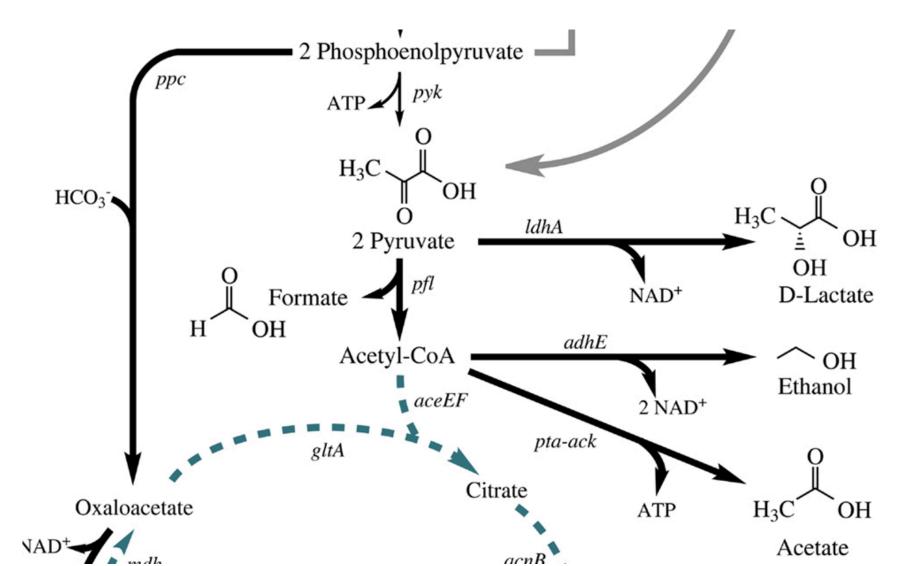
## **Regular office hours**

- Noreen: Mon. 2-5pm (16-317)
- Leslie: Fri. 9am and 3pm (56-341c)
- Josephine: Mon. 1pm, Thurs. 2pm (56-341c)

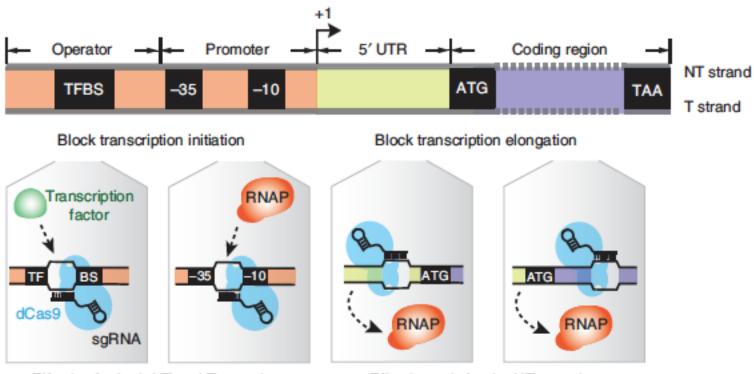
### M2 experimental overview



## Using CRISPRi manipulate the *E. coli* fermentation pathway



# Design of gRNA for CRISPRi system



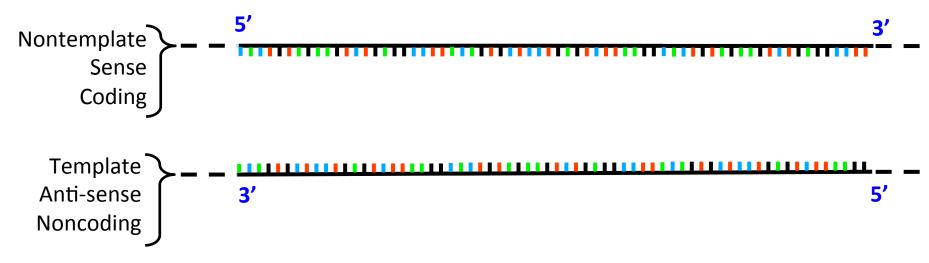
Effective for both NT and T strands

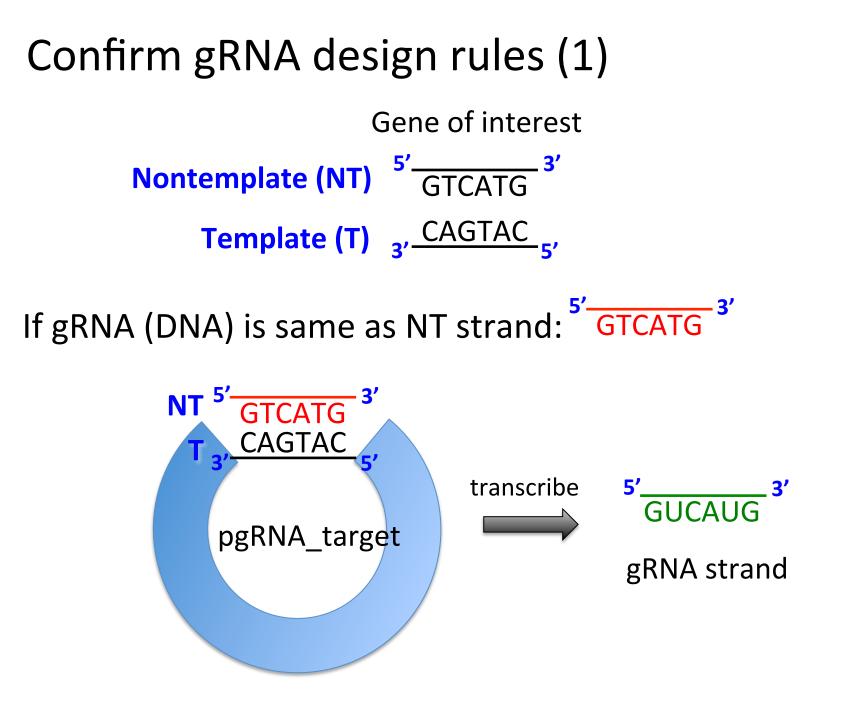
Effective only for the NT strand

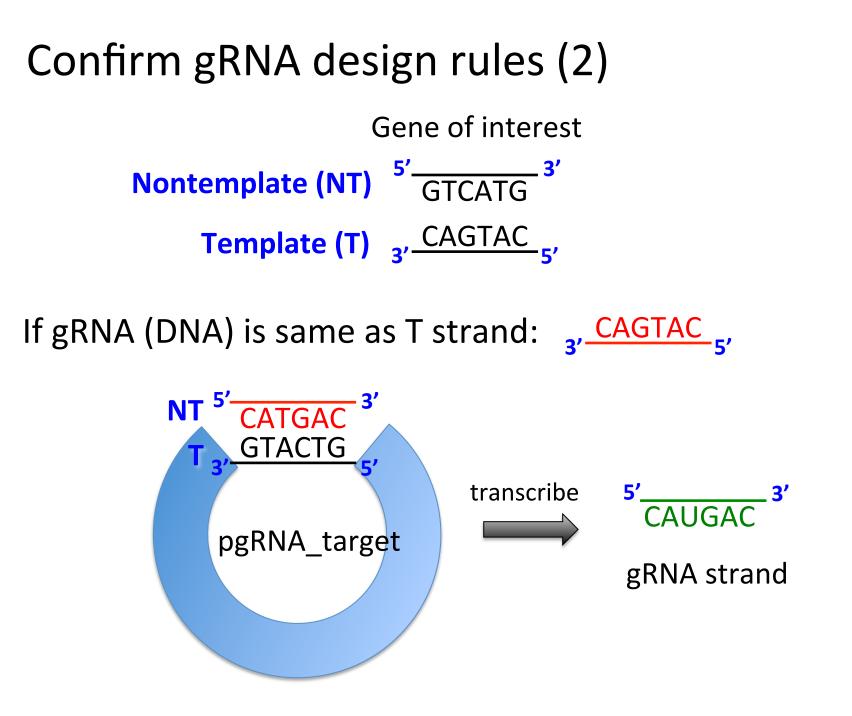
### Design of gRNA for CRISPRi system

(1) If you target the template DNA strand, the gRNA (DNA) sequence will be the same as the transcribed (nontemplate) sequence.

(2) If you target the nontemplate strand, the gRNA (DNA) sequence will be the reverse-complement of the transcribed (template) sequence.







# Please add your targeting info to the wiki today

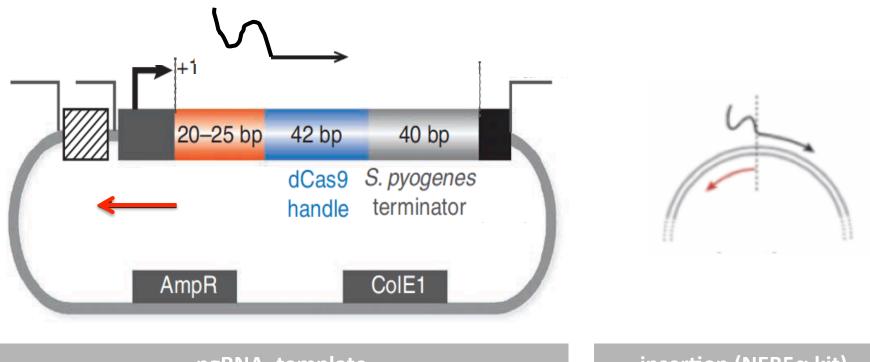
#### -> On the Mod2 Overview page, discussion tab

W/F

Team	Ethanol (E) or Acetate (A)	Gene targeted by CRISPRi gRNA	gRNA sequence end)
red	Ethanol	ack (indirectly, pta)	GTTTTTTTAGCC
orange	Ethanol	ldhA	ATTCAACATCAC
blue	Ethanol	ackA	TTTTTAGCCACG

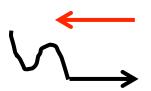
	Target template or nontemplate strand
ГТ	

## M2D3: Generated pgRNA\_target by SDM





insertion (NEB5α kit)

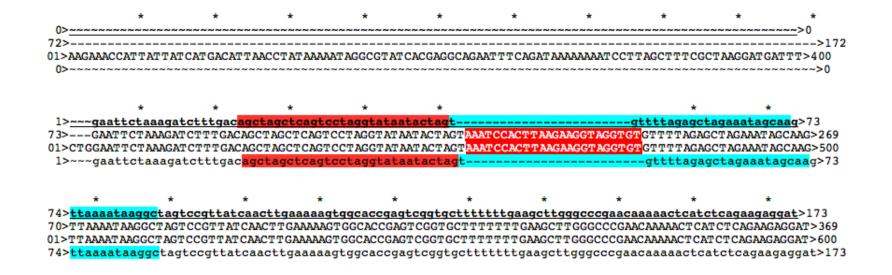


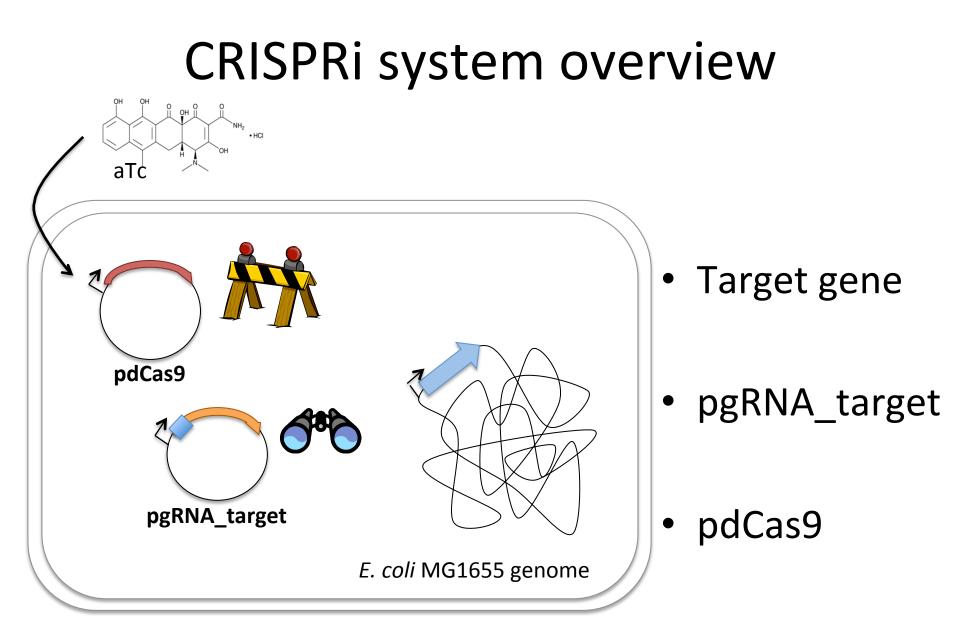
CRISPRi universal *amplification* reverse primer forward primer including crRNA to be inserted ( $\bigcirc$ )

dCas9 handle ( -----> )

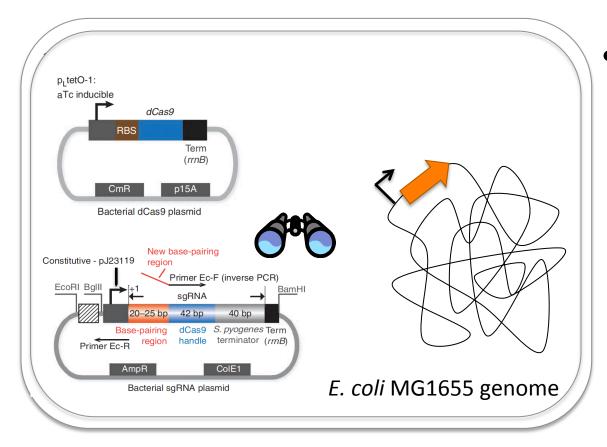
# Analyzing Sequence Information

- Remember to reverse and complement your reverse primer sequence before alignment
- Consider importing your entire gRNA expression vector seq. with your target sequence if alignment is not working well

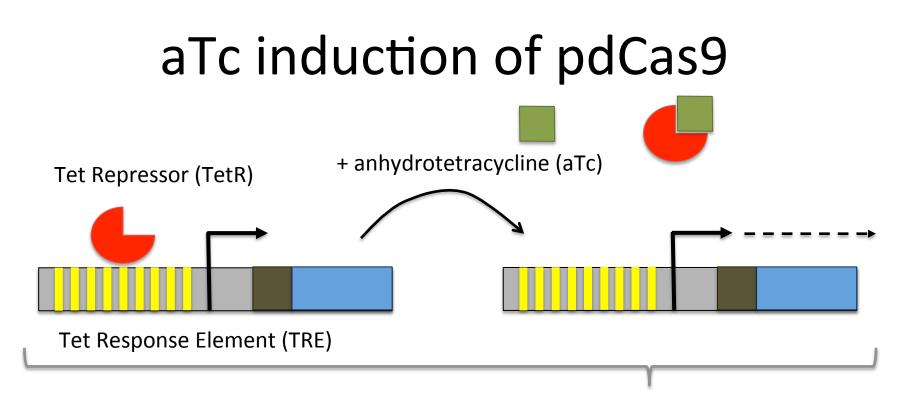




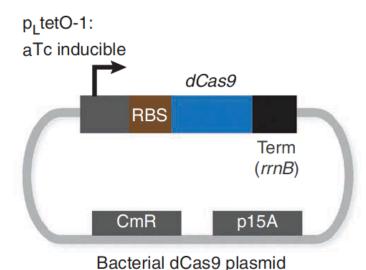
# CRISPRi 'inactive' in absence of inducer



pgRNA\_target
 expressed
 constitutively
 Always
 transcribed and
 binding to
 target gene



 Tet promoter regulates expression of dCas9 gene



#### CRISPRi 'blocks' gene expression in presence of inducer HCI aTc pdCas9 p<sub>1</sub> tetO-1: aTc inducible expressed when dCas9 RBS aTc added Term (rrnB) CmR p15A – When Bacterial dCas9 plasmid New base-pairing transcribed Constitutive - pJ23119 region Primer Ec-F (inverse PCR) EcoRI Ball BamHI saRNA associates with 42 bp 40 bp dCas9 S. pyogenes Term Base-pairing handle terminator (rrnB) region pgRNA target / Primer Ec-R CoIE1 AmpR E. coli MG1655 genome Bacterial sgRNA plasmid target gene

# Media for mixed-acid fermentation and pdCas9 induction

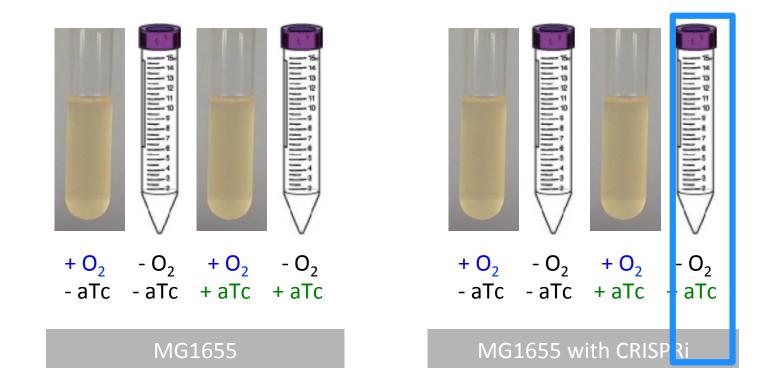
• What are the necessary components?

 $\mathsf{IR}$ 

Antibiotics - AMP, CAM aTc

# Media for mixed-acid fermentation and pdCas9 induction

• Where do we expect most ethanol if hypothesis confirmed?



# Today in lab...

- Download your sequencing data from discussion tab and align using ApE software
- Prepare media for mixed-acid fermentation inoculations

## M2D8 Assignments

- Quiz on Tues (M2D8) Nov. 7<sup>th</sup>
- Peer-review (see wiki for details)