

# Reflection Activity

1. Why might we care about scientific communication?

When will we need to communicate science?

2. What makes you feel that any communication has been successful?

As a receiver?

As a sender?

What makes you feel that communication has been successful?

As a receiver?

Clear message, logic flows, you can find your way around, visual appeal

As a sender?

Reward (citation, grade, funding),  
good feedback: questions or criticism

# We often blame ourselves for struggling to understand talks or papers...

“I got stuck here. I feel like there was a huge logical leap I couldn't follow.”

“There's way too much going on in this plot. What am I supposed to be looking at?”

but poor communication is often the barrier, not your scientific understanding.

In these workshops, we'll turn your instincts as a reader of science into tools for identifying...

WHEN scientific communication is confusing

WHY it's confusing

HOW to fix the problem

...and start applying these tools to your 20.109 work.

## What we'll do in a workshop:

1. Discuss an example from the field
2. Derive principles and strategies
3. Practice strategies
4. Go home with a checklist/rubric

Practice with a fellow at the



[be.mit.edu/communicationlab](https://be.mit.edu/communicationlab)

# Designing Effective Figures

20.109 Communication Workshop 1

Dr. Prerna Bhargava and Dr. Sean Clarke



[be.mit.edu/communicationlab](http://be.mit.edu/communicationlab)

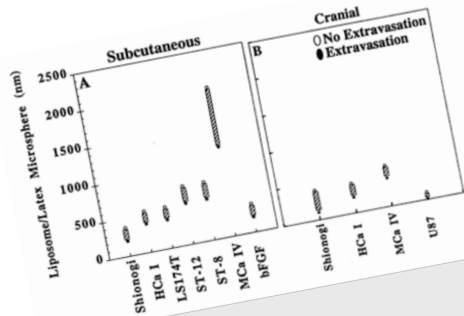
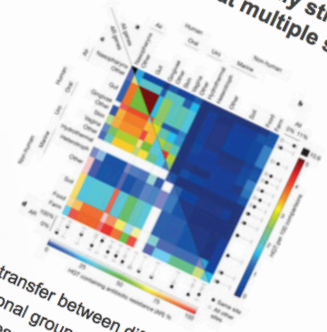


Figure 1

The vascular pore cutoff size for six different types of tumors grown in the dorsal window (A) and in the cranial window (B) was evaluated. The solid circles represent tumors grown in the cranial window (B) was evaluated. The size range below the hatched bar indicates the size range below which liposomes/latex beads did not extravasate and the first particle that did extravasate indicates the vascular pore cutoff size range. The majority of tumors have a vascular pore cutoff size range of approximately 1000-1500 nm when grown subcutaneously in the dorsal chamber. The interaction of the tumor with the subcutaneous microenvironment (A) leads to a smaller pore cutoff size than the interaction of the tumor with the cranial microenvironment (B). Comparison of bFGF-induced vessels (bFGF) with the subcutaneous pore sizes demonstrates that the presence of bFGF alone can lead to pores of approximately 1000-1500 nm.

Figure 3: HGT is ecologically structured by functional class and at multiple spatial scales



The frequency of transfer between different environments is shown for all functional groups (a, b) and for antibiotic resistance (AR) genes only (c, d). Box widths indicate the number of genomes from each environment. a, When all genes...

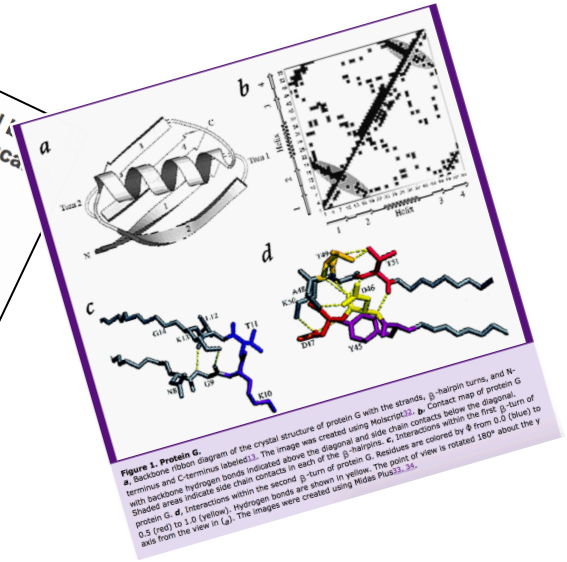


Figure 1. Protein G. a. Backbone ribbon diagram of the crystal structure of protein G with the strands,  $\beta$ -hairpin turns, and N-terminus and C-terminus labeled. The image was created using Molscript. b. Contact map of protein G with backbone hydrogen bonds indicated above the diagonal and side chain contacts below the diagonal. Shaded areas indicate side chain contacts in each of the  $\beta$ -hairpins. c. Interactions within the first  $\beta$ -turn of protein G. d. Interactions within the second  $\beta$ -turn of protein G. Residues are colored by  $\phi$  from 0.0 (blue) to 0.5 (red) to 1.0 (yellow). Hydrogen bonds are shown in yellow. The point of view is rotated 180° about the y-axis from the view in (a). The images were created using Molscript and PyMol.

# Figures (and captions)

Why start here?

# Figures must convince your audience of your data's impact and credibility.

Science AAAS

Article

Figures & Data

Info & Metrics

eLetters

PDF

PNAS

Abstract

Full Text

Authors & Info

Figures

Metrics

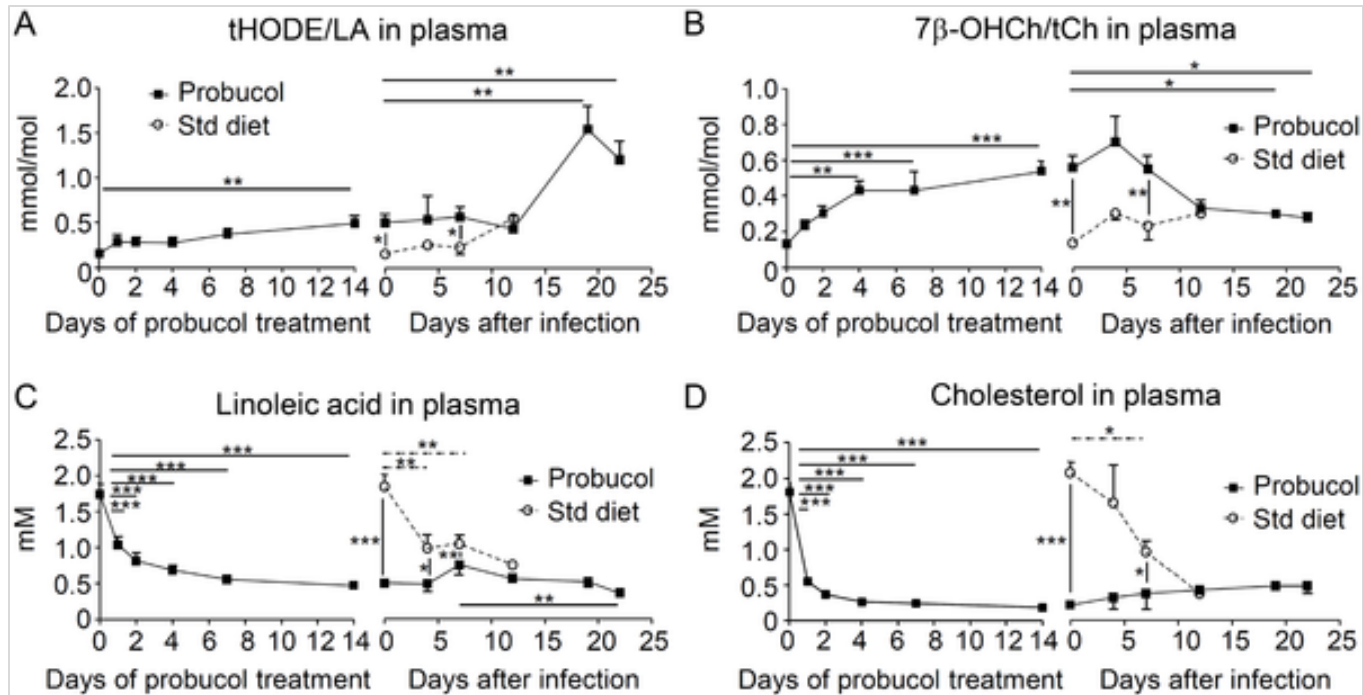
Related Content

PDF

- Expert audiences may ONLY read: Title  
Abstract  
FIGURES
- Figures help tell your story **compellingly** and **honestly**.
- Figures present your “naked” data for evaluation.



# Activity: Identify the basic figure components



**Fig 4. The ratios of lipid peroxidation products to parent lipids in plasma increased after probucol pre-treatment.**

Six-week-old C57BL/6J mice were treated with 1% w/w probucol in the diet for 2 weeks and then infected with 0.2 mL of  $1 \times 10^5$  erythrocytes/mL infected with *Plasmodium yoelii* XL-17. Plasma samples were obtained at day 0, 1, 2, 4, 7, and 14 after starting the probucol diet (n = 5 per group) and at day 0, 4, 7, 12, 19, and 22 post-infection (n = 2 to 7). The ratio of total hydroxyoctadecadienoic acid (HODE), a peroxidation product of linoleic acid (LA), to linoleic acid (tHODE/LA) in plasma (A) and the ratio of 7β-hydroxycholesterol (7β-OHCh), a peroxidation product of cholesterol, to total cholesterol (7β-OHCh/tCh) in plasma (B) were measured. The concentration of LA (C) and tCh (D) were measured by using gas chromatography-mass spectrometry (GC-MS). All data are expressed as mean  $\pm$  SE. Statistical analysis was carried out by analysis of variance (ANOVA). \* $p < 0.05$ , \*\* $p < 0.025$ , and \*\*\* $p < 0.001$ . The solid bars indicate the significant changes in probucol-treated groups and the dotted bars indicate the significant changes in standard (Std) diet-fed mice.

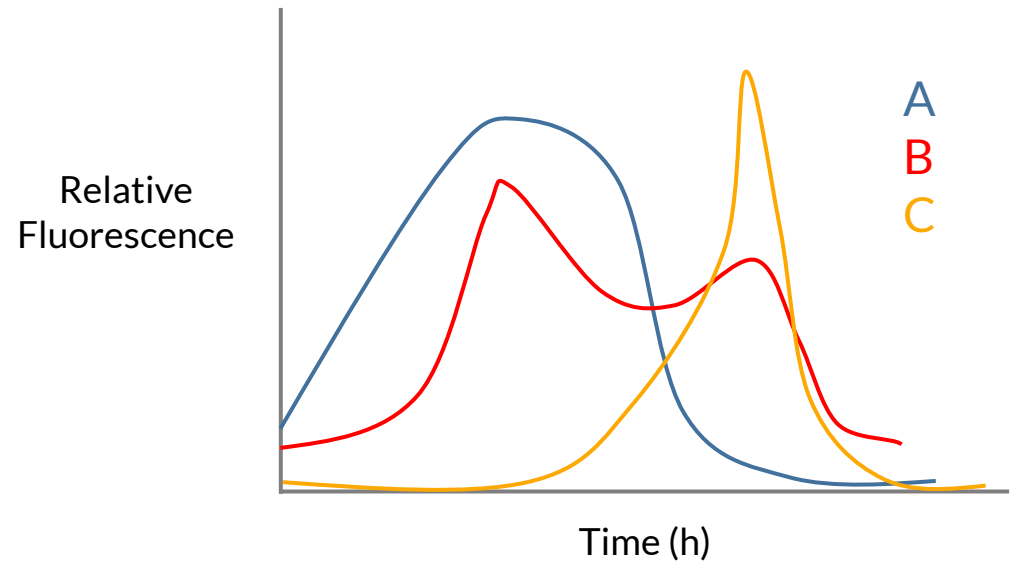
# Figure = message + data

## Choice of data

- Only data critical to the conclusion

## Presentation choices

- Type of graph or display, legends & labeling, design choices
- Uncluttered elements
- Allow quick evaluation of conclusions without relying on the legend or caption.

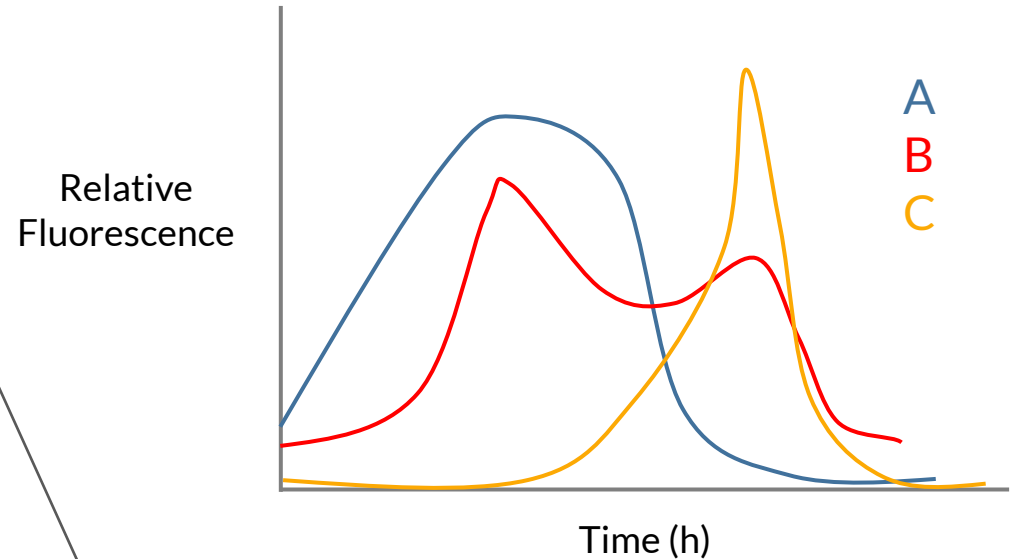


**Figure 1. A, B, and C have different dynamics under Condition X.** A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control.

# Figure titles express your message

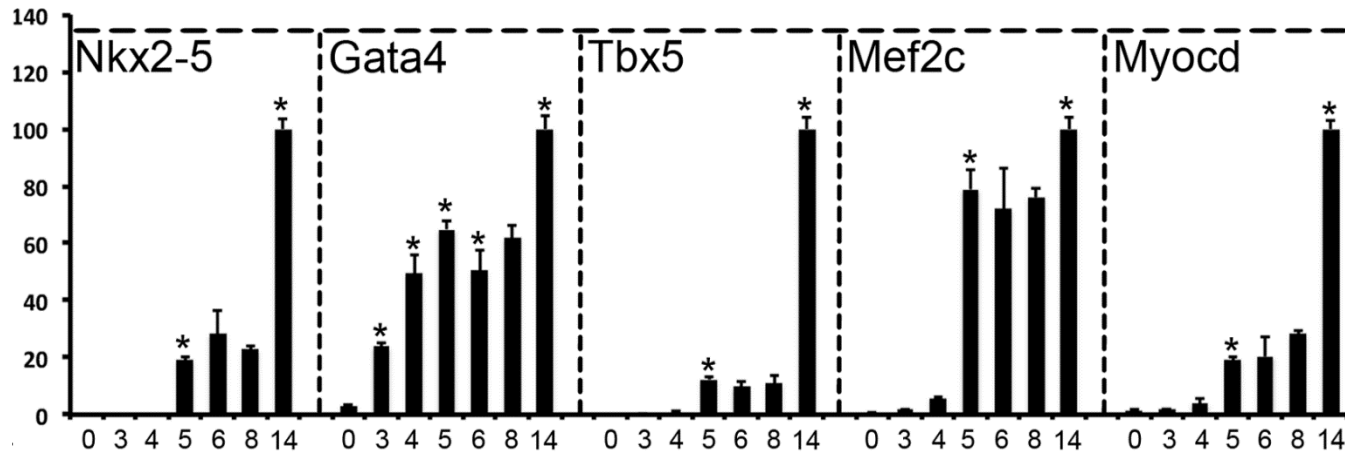
## Title

- Take-home *message* of the figure
- What conclusion should the reader evaluate when looking at the figure?



**Figure 1. A, B, and C have different dynamics under Condition X. A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control.**

# Message: Use titles to state a figure's message, not the method



Gene expression analysis performed on differentiating mouse iPS cells



Expression of early cardiac transcription factors increases over time in differentiating mouse iPS cells

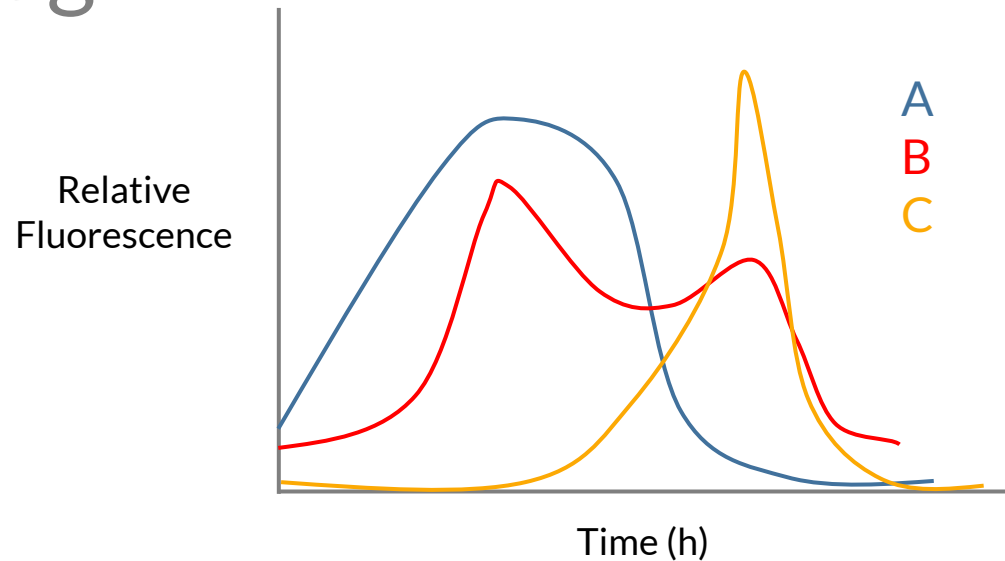
# The caption is how you got the data, without a message

## Title

- Take-home *message* of the figure
- What conclusion should the reader evaluate when looking at the figure?

## Caption

- Descriptive, not explanatory/interpretive
- Only enough methodological detail to make it clear how results were obtained.



**Figure 1. A, B, and C have different dynamics under Condition X. A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control.**

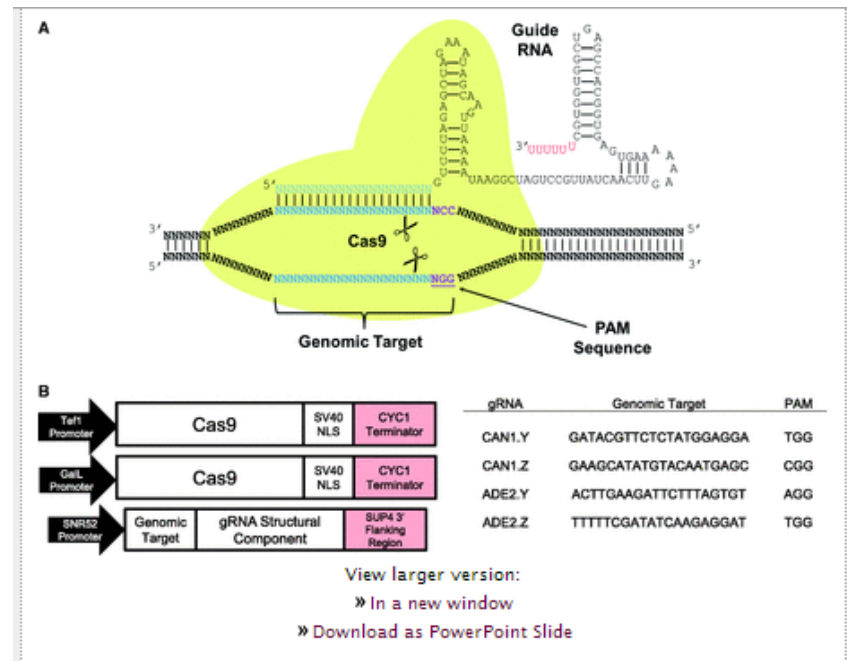
# All figures you make need all of these components

Schematics

Diagrams

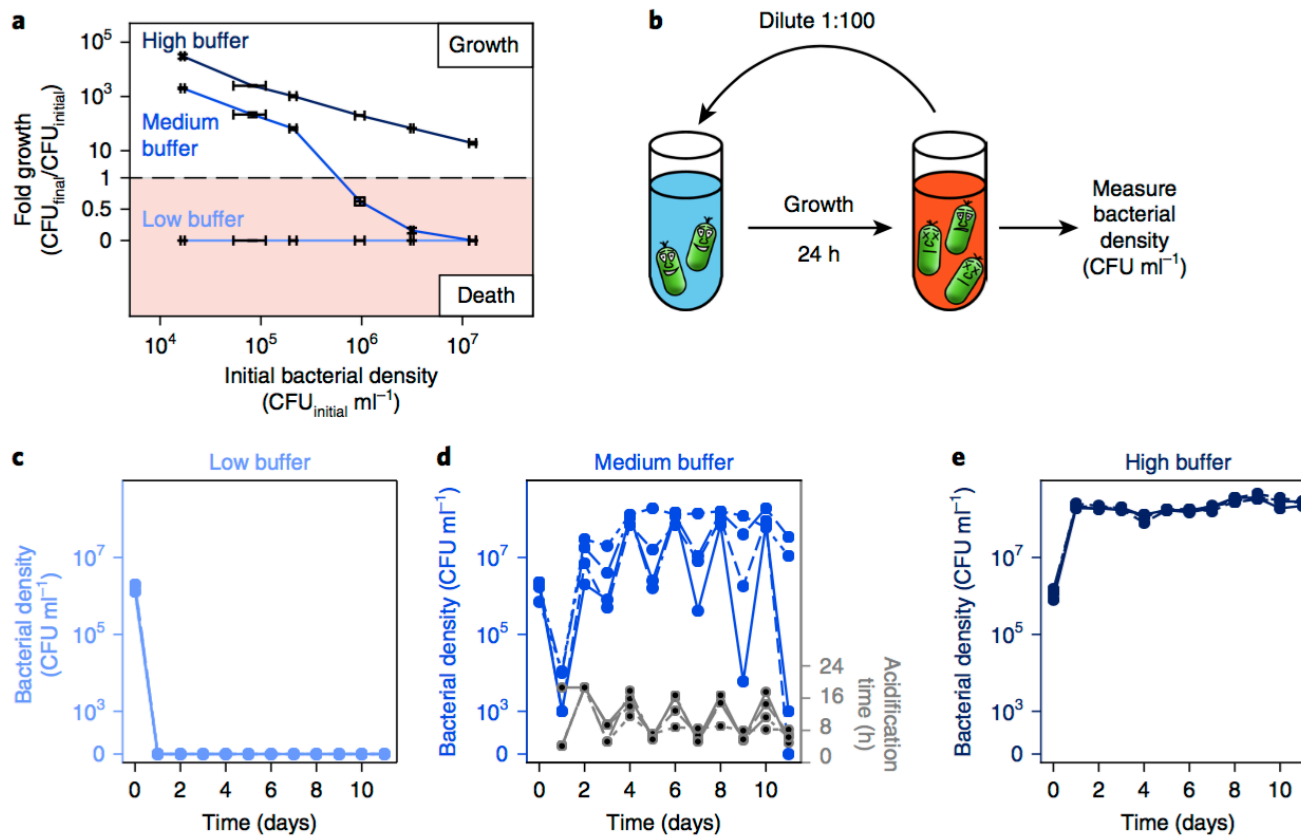
Photos

are figures too!



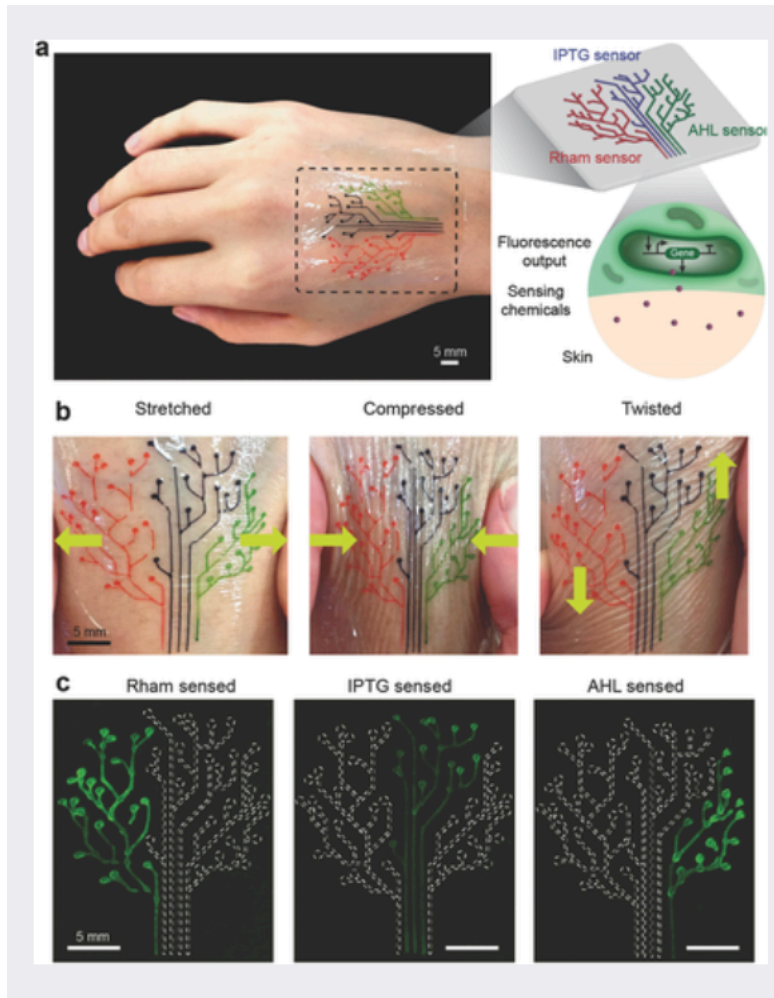
**Figure 1.**

Diagram of Cas9 complex and schematic of genetic constructs. (A) Illustration of Cas9 protein interacting with CRISPR gRNA to direct endonuclease activity proximal to the PAM sequence. (B) Design of the Cas9 and gRNA constructs. Cas9 gene contained a SV40 nuclear localization signal and was expressed under the Gal-L inducible promoter in CAN1 experiments and the TEF1 constitutive promoter in ADE2 experiments. The gRNA was



**Fig. 2 | Ecological suicide can cause oscillations in the population size over time.** **a**, At a low buffer concentration (10 mM phosphate), the bacteria commit ecologic suicide, whereas at a high buffer concentration (100 mM phosphate), the bacteria grow, in both cases independent of their initial density. However, at a moderate buffer concentration (26 mM phosphate), the bacteria die at high starting densities and grow at low starting densities. The fold growth at a high buffer concentration decreases for increasing initial bacterial densities, since the final bacterial density equals the carrying capacity and is therefore constant. Mean (solid lines) and s.e.m. (error bars) are shown for four replicates. The black horizontal dashed line corresponds to a fold growth of 1. **b**, To explore long time growth dynamics, the bacteria were grown in a daily dilution scheme with 24 h of incubation in well-mixed conditions followed by a 1:100 dilution into fresh medium. **c,e**, At low (10 mM phosphate; **c**) and high (100 mM phosphate; **e**) buffer conditions, the bacteria either die on the first day or grow to saturation every day. **d**, However, at medium buffer conditions, we measure oscillatory dynamics of the bacterial density. This is accompanied by oscillations in the time that the bacteria need to acidify the environment (acidification time, Supplementary Fig. 8). The exact type of oscillatory dynamics depends on the slope and shape of the curve in **a**, as discussed in more detail in the Supplementary Information. **c-e**, The four blue lines (solid, dashed, dotted, dashed-dotted; the separated curves can be seen in Supplementary Fig. 5) show different replicates. The strong differences between the replicates highlight the sensitivity of these oscillations to experimental conditions and that they probably do not show a limit cycle oscillation.





**Figure 5**

3D-printed living tattoo for chemical detection on human skin. a) The design of the living tattoo. The tattoo is printed as a tree-like pattern on a thin elastomer layer, which is then adhered to human skin. Hydrogels with different colors illustrate the different types of cells encapsulated. Inset: Schematic illustration of living sensors embedded in the tattoo, which can respond to different chemicals by emitting fluorescence. b) The living tattoo on skin in different states: stretched (left), compressed (middle), and twisted (right). Food dyes are added to facilitate visualization of the hydrogel pattern in (a) and (b). c) The response of the living tattoo on the skin smeared with Rham (left), IPTG (middle), or AHL (right).



# Steps to turn your pile of data into figures...

## 1 MESSAGE

What is the message of each figure?



## 2 DATA

What data do you include in each figure to convey your message?

How can you present your data to support your message?



## 3 DESIGN

What are some key design choices to think about?

# Organize your figures to build one storyline

Rearrange until you've created a **logical series** of conclusions.

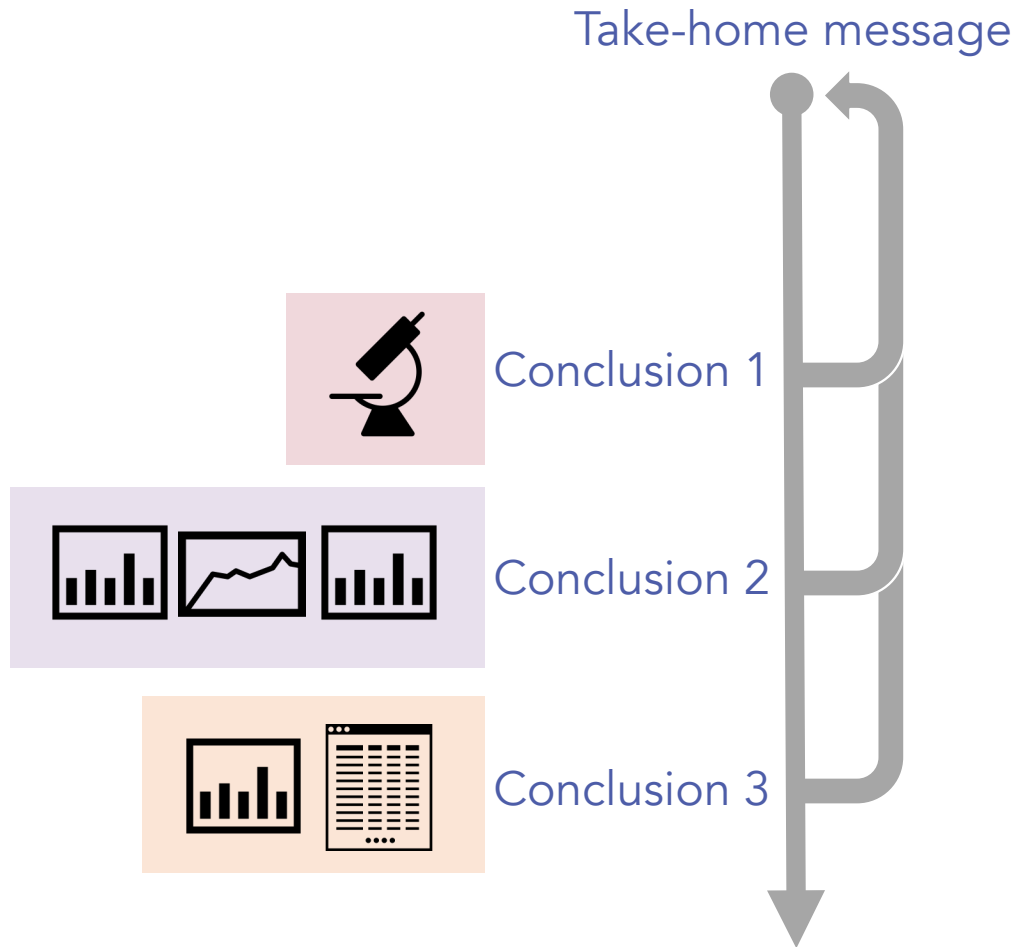


Identify **modules** that correspond to **conclusions**.

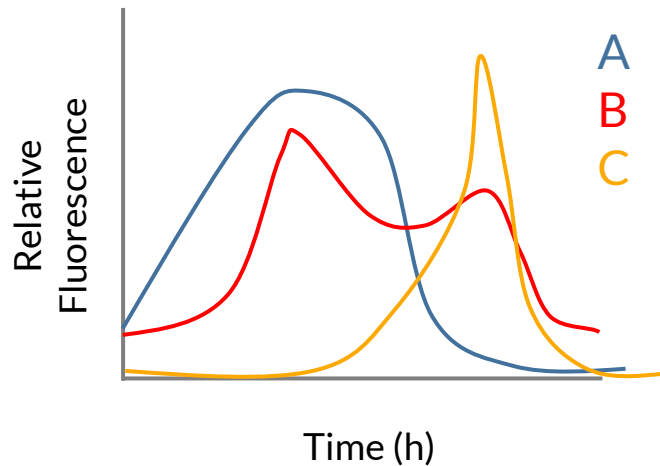


# Organize figures to build a single storyline

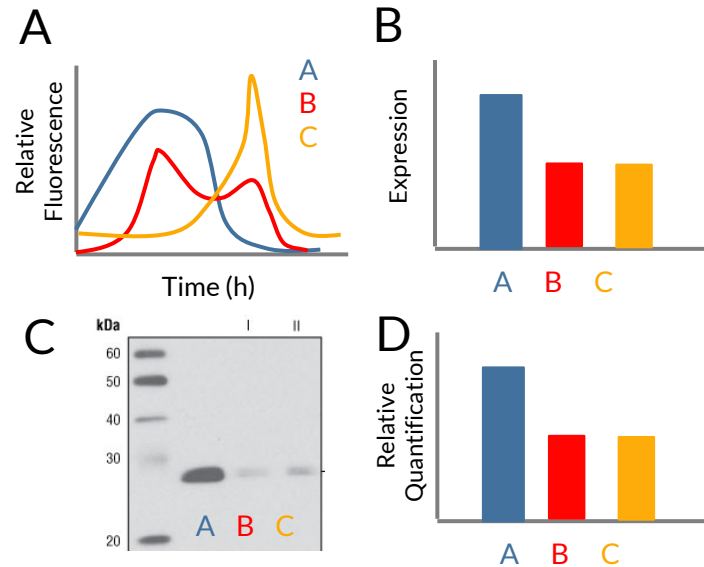
Identify modules that correspond to conclusions.



# Message: All the data in a figure should support one clear message.



**Fig. 1: A, B, and C have different dynamics under Condition X.** A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control.



**Fig. 1: A, B, and C have different dynamics under Condition X.** A) A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control. B) Gene expression data of samples A, B, and C, under condition X. Samples were collected at time T. C) Western blot analysis of samples A, B, and C, under condition X. D) Quantification of Western Blot.

# Steps to turn your pile of data into figures...

## 1 MESSAGE

What is the message of each figure?



## 2 DATA

What data do you include in each figure to convey your message?

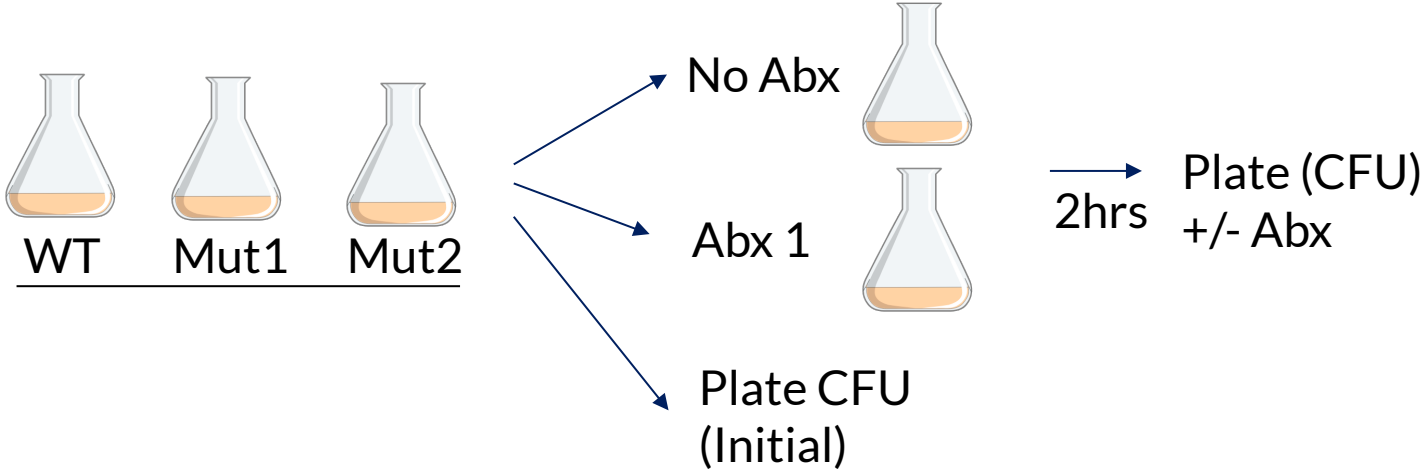
How can you present your data to support your message?



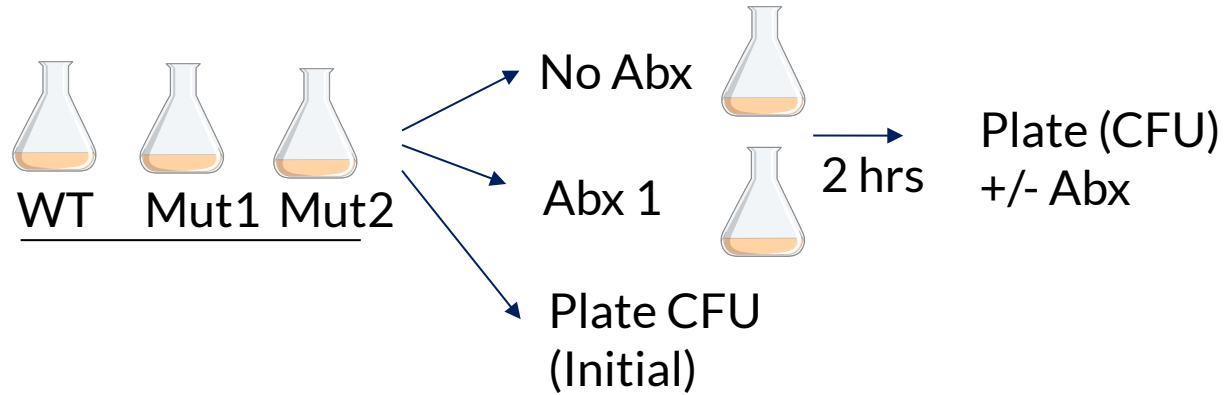
## 3 DESIGN

What are some key design choices to think about?

# Activity: Here's an experiment

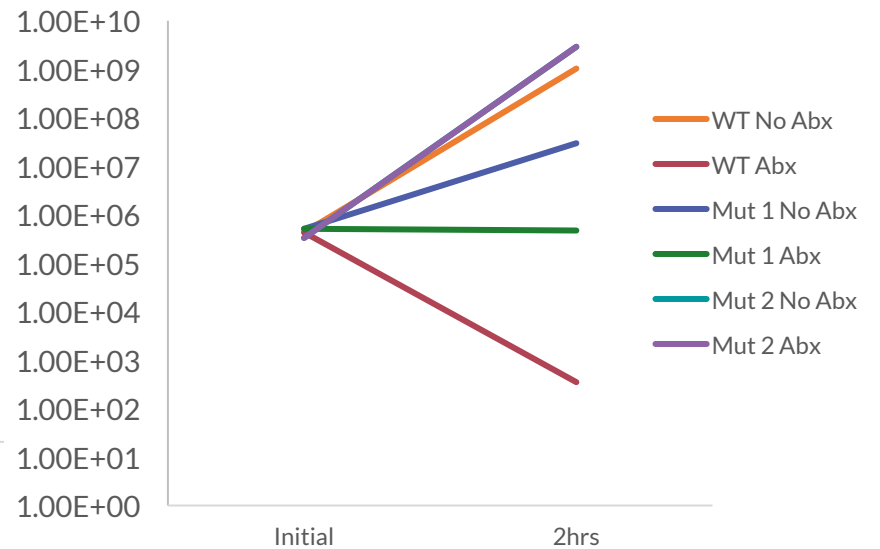
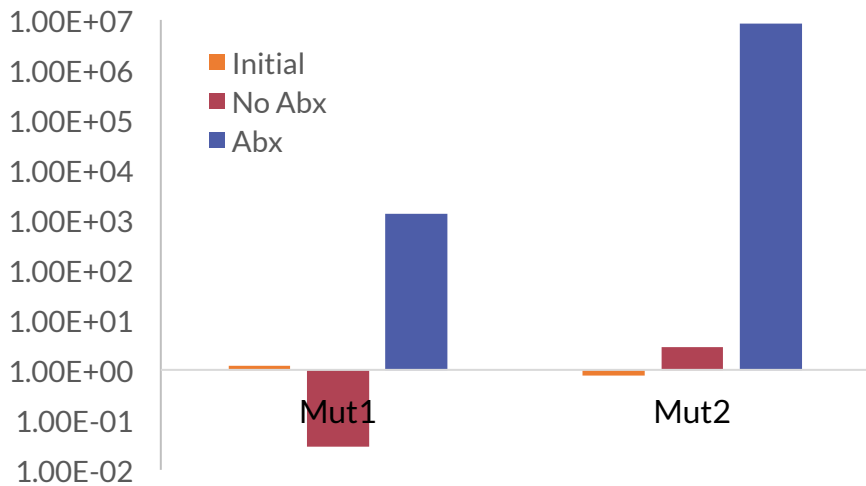
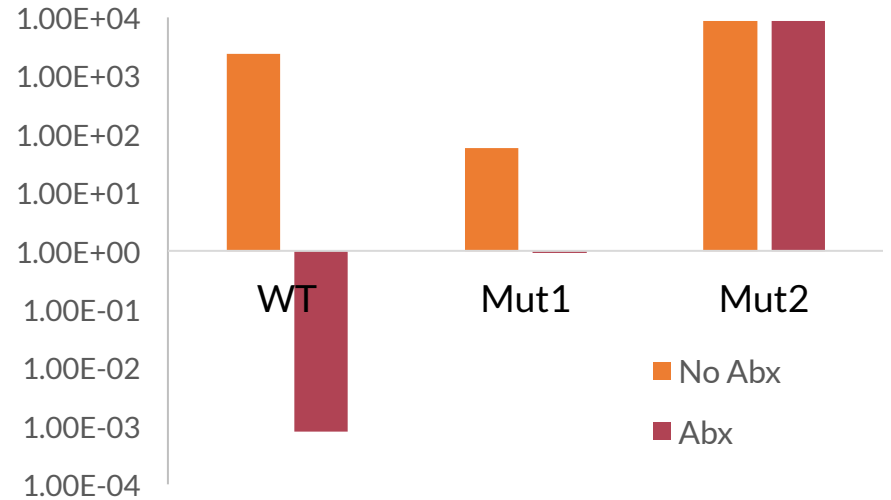
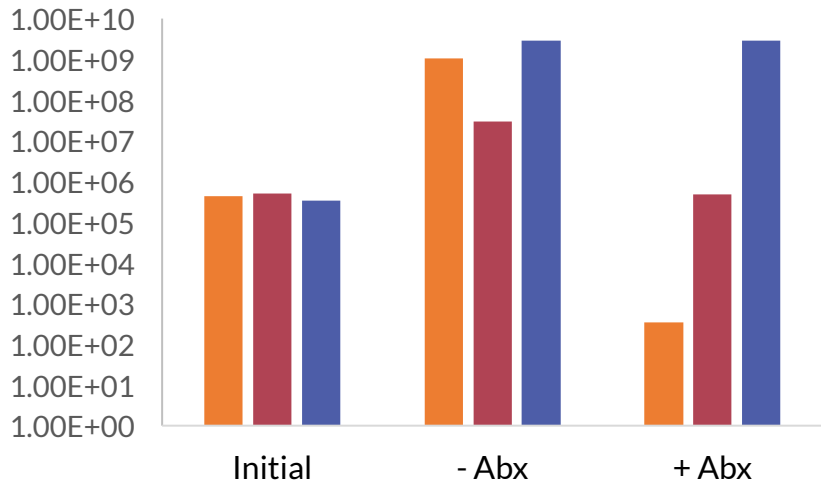


# Activity: How can we present this data?

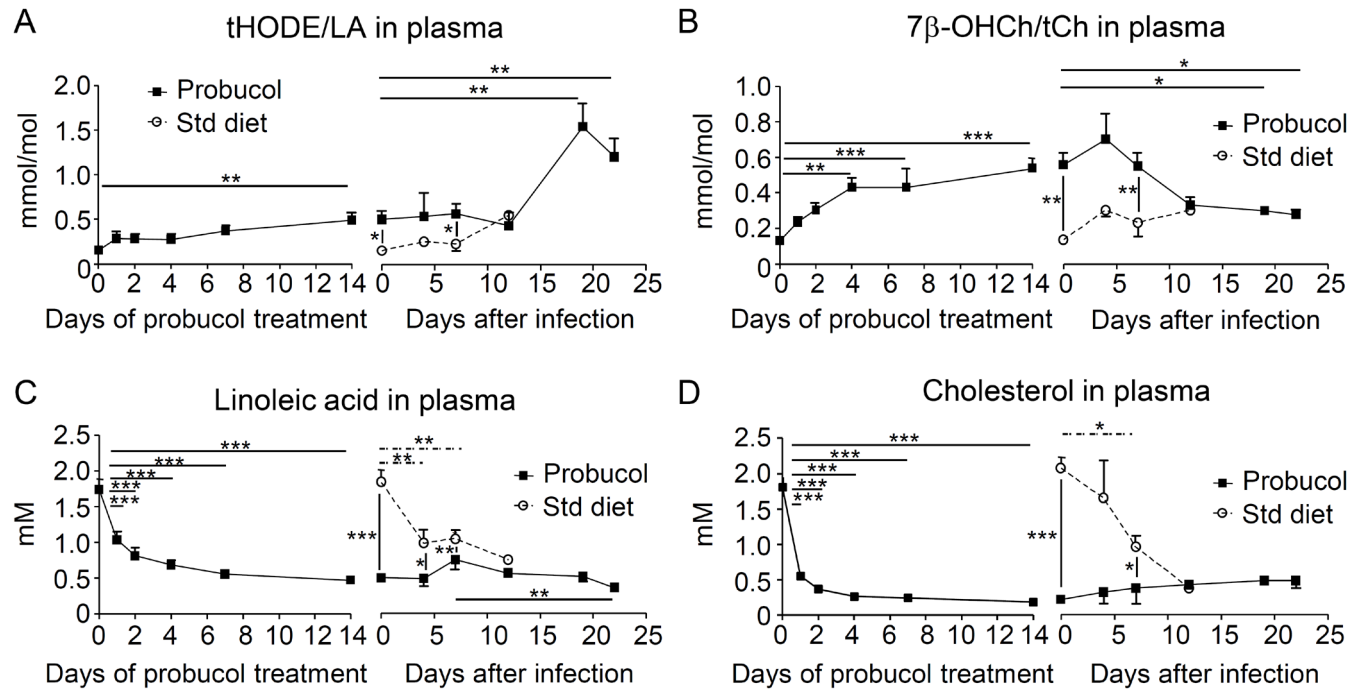


Strain	Condition	Replicate 1	Replicate 2	Replicate 3	Average
WT	Initial	1.8e5	3.2e5	7.8e5	4.3e5
WT	- Abx	1.0e9	1.3e9	8e8	1.0e9
WT	+ Abx	2.3e2	2.8e2	5.5e2	3.5e2
Mut1	Initial	2.5e5	8.3e5	4.6e5	5.1e5
Mut1	- Abx	5.5e7	2.3e7	1.1e7	3.0e7
Mut1	+ Abx	4.3e5	7.5e5	2.2e5	4.7e5
Mut2	Initial	5.3e5	3.2e5	1.3e5	3.3e5
Mut2	- Abx	3.4e9	2.2e9	3.0e9	2.9e9
Mut2	+ Abx	2.2e9	5.3e9	1.2e9	2.9e9





# Activity: Improve this published figure



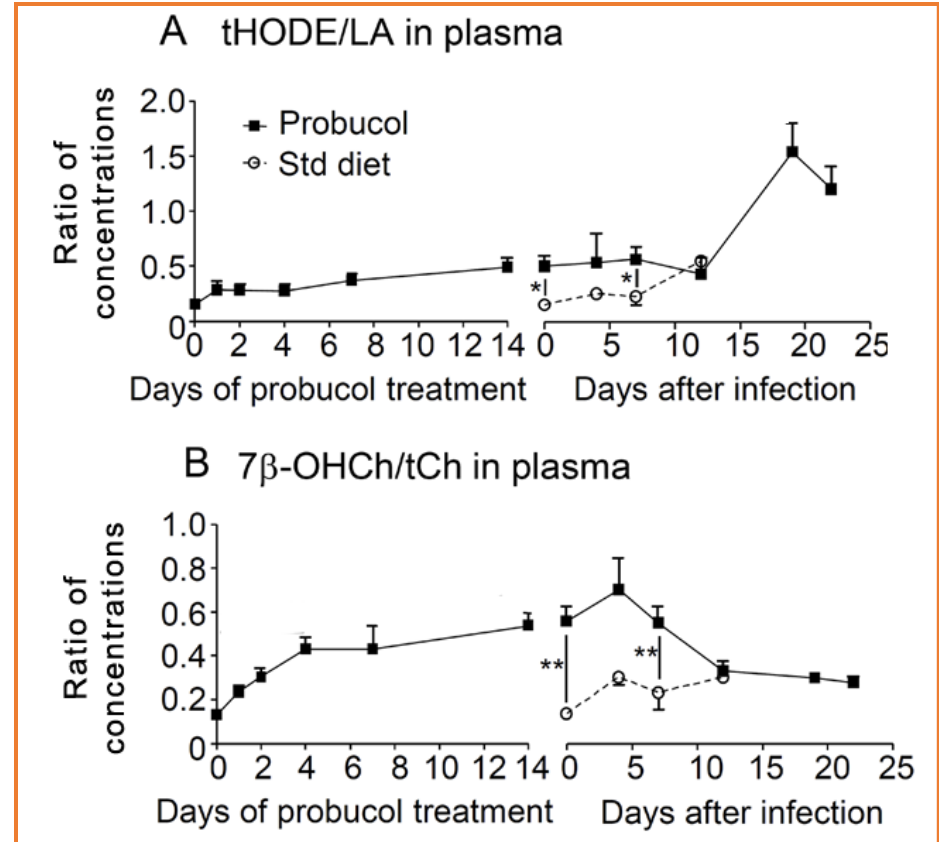
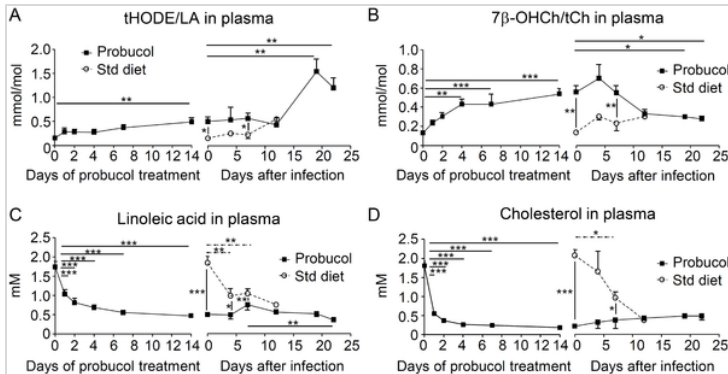
**Fig 4. The ratios of lipid peroxidation products to parent lipids in plasma increased after probucol pre-treatment.**

Six-week-old C57BL/6J mice were treated with 1% w/w probucol in the diet for 2 weeks and then infected with 0.2 mL of  $1 \times 10^5$  erythrocytes/mL infected with *Plasmodium yoelii* XL-17. Plasma samples were obtained at day 0, 1, 2, 4, 7, and 14 after starting the probucol diet (n = 5 per group) and at day 0, 4, 7, 12, 19, and 22 post-infection (n = 2 to 7). The ratio of total hydroxyoctadecadienoic acid (HODE), a peroxidation product of linoleic acid (LA), to linoleic acid (tHODE/LA) in plasma (A) and the ratio of 7β-hydroxycholesterol (7β-OHCh), a peroxidation product of cholesterol, to total cholesterol (7β-OHCh/tCh) in plasma (B) were measured. The concentration of LA (C) and tCh (D) were measured by using gas chromatography-mass spectrometry (GC-MS). All data are expressed as mean ± SE. Statistical analysis was carried out by analysis of variance (ANOVA). \*p < 0.05, \*\*p < 0.025, and \*\*\*p < 0.001. The solid bars indicate the significant changes in probucol-treated groups and the dotted bars indicate the significant changes in standard (Std) diet-fed mice.

# Evaluating figure choices

- Is the central message validated by the data shown?
- Which data are irrelevant?
- Are there any data/labels missing?
- What could be done to better highlight the most important data?
- Is there a better way to present the data?
- Do the statistics actually add anything here?

# Only include the minimum information necessary to draw a conclusion.



# Steps to turn your pile of data into figures...

## 1 MESSAGE

What is the message of each figure?



## 2 DATA

What data do you include in each figure to convey your message?

How can you present your data to support your message?



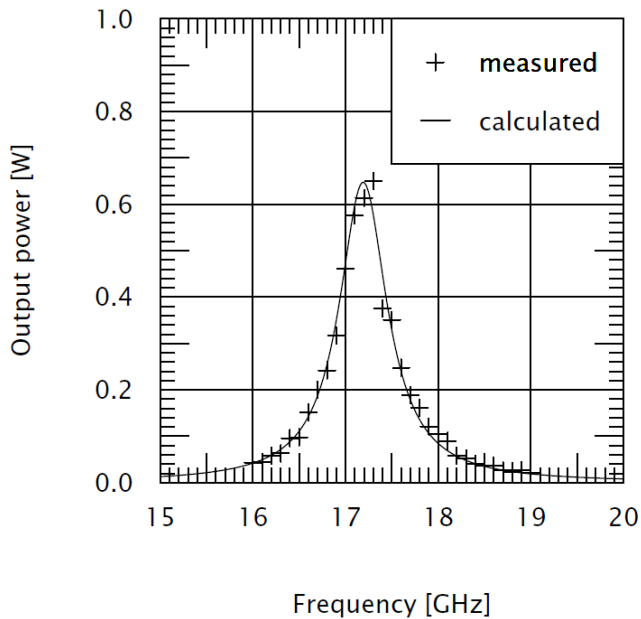
## 3 DESIGN

What are some key design choices to think about?

# Design: Maximize signal-to-noise

State your message.

Eliminate anything that distracts from it.



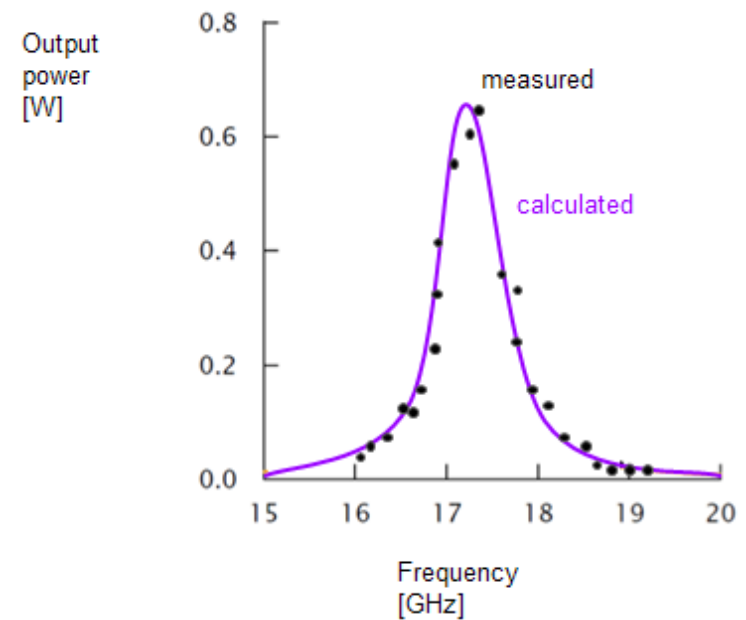
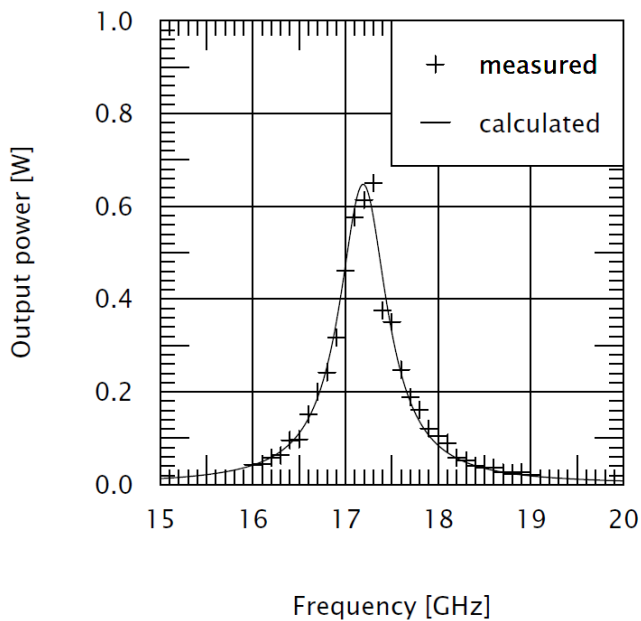
## Low signal-to-noise ratio

The background interferes with the data.

# Design: Maximize signal-to-noise

State your message.

Eliminate anything that distracts from it.



## Low signal-to-noise ratio

The background interferes with the data.

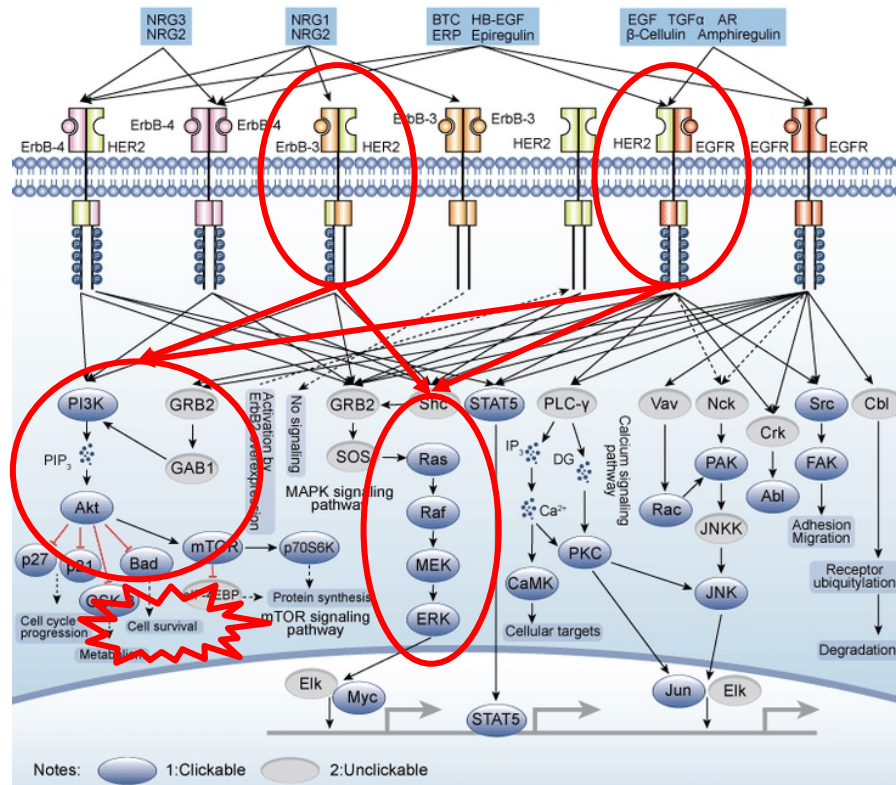
## High signal-to-noise ratio

Only the necessary information is shown.

# Design: Schematics should include info that reinforces your message

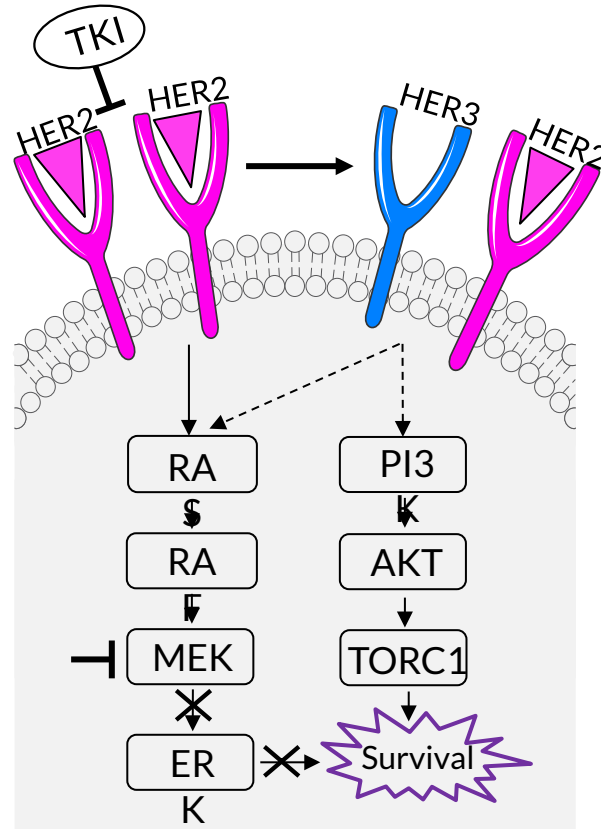


receptor tyrosine kinase signaling pathway





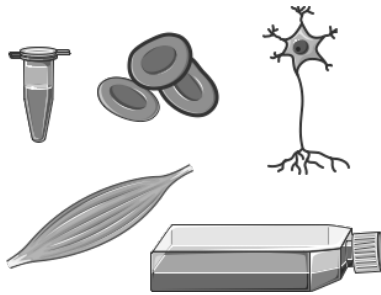
# Design: Schematics should include info that reinforces your message



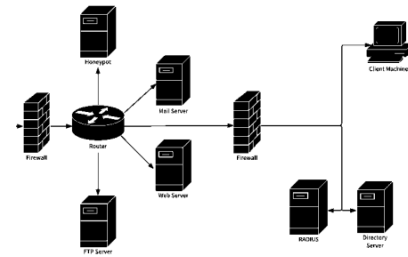
# Design: You don't need to reinvent the wheel

Servier Powerpoint Image Bank &  
Biorender

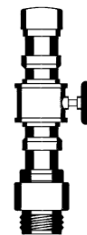
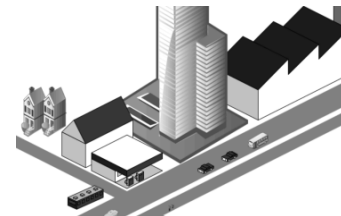
Biology, lab equipment  
(free)



MS Visio & Lucidchart  
Networks, engineering, circuits, charts  
(\$\$) & (free)



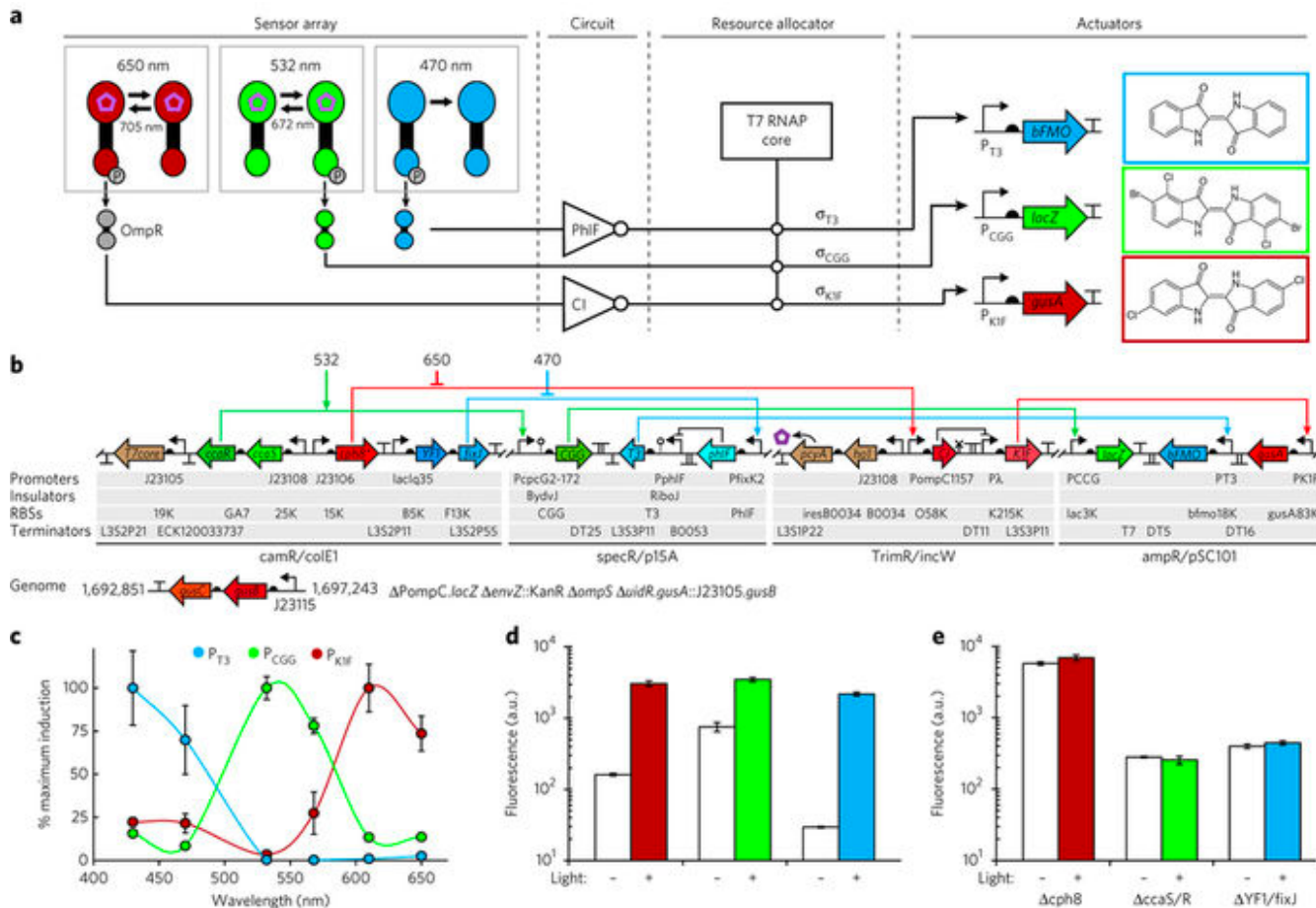
Noun Project  
Everything  
(free)



# Design: Maximize signal-to-noise

## Minimize # style choices and be consistent

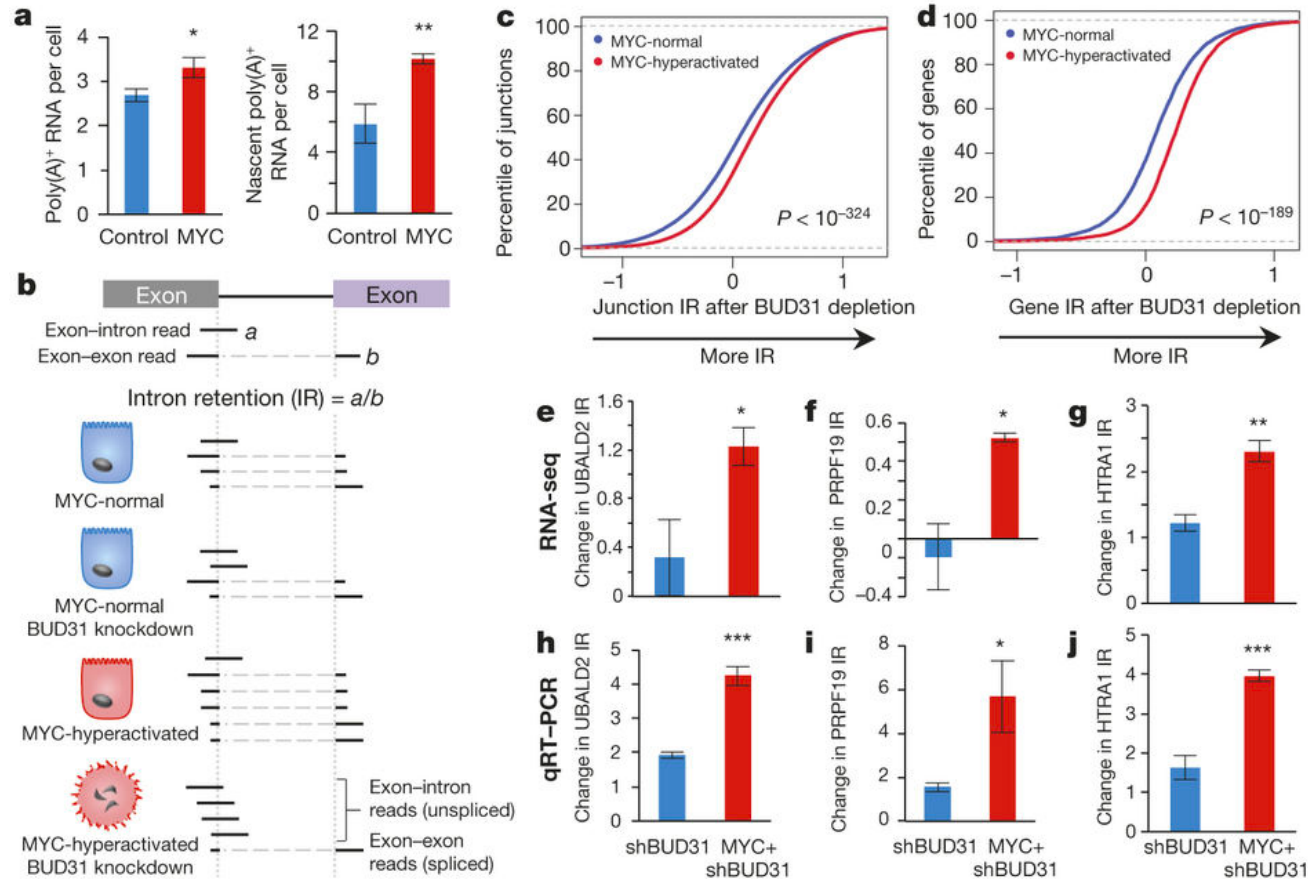
- Grayscale or **a few consistent colors**



# Design: Maximize signal-to-noise

Minimize # style choices and be consistent

- S, M, L font sizes
- Consistent fonts and line thickness
- Consider alignment in making figures



# These are our next steps

- Slides and tips will be on the wiki
- Put these tips to work on your 109 figures today and beyond
- Make an appointment with a Comm Fellow by M1D5
- We're happy to help with all parts of data summary drafts
- In 2 weeks... Titles and Abstracts!

# Optimize your figures with these reminders

## High-level questions

- *Strategic purpose:*
  - What do you want to convey?
  - How will you and/or your audience use this figure?
- *Organizational structure:*
  - Where does this figure fit into the communication?
  - Why?

## Checklist

- Choice of data
- Title/caption
  - Can figure stand alone?
- Consistent layout
  - Fonts, spacing, colors
- Text amount and placement
- Scale, axes, tick marks
- Error analysis
- Ink-to-whitespace ratio

