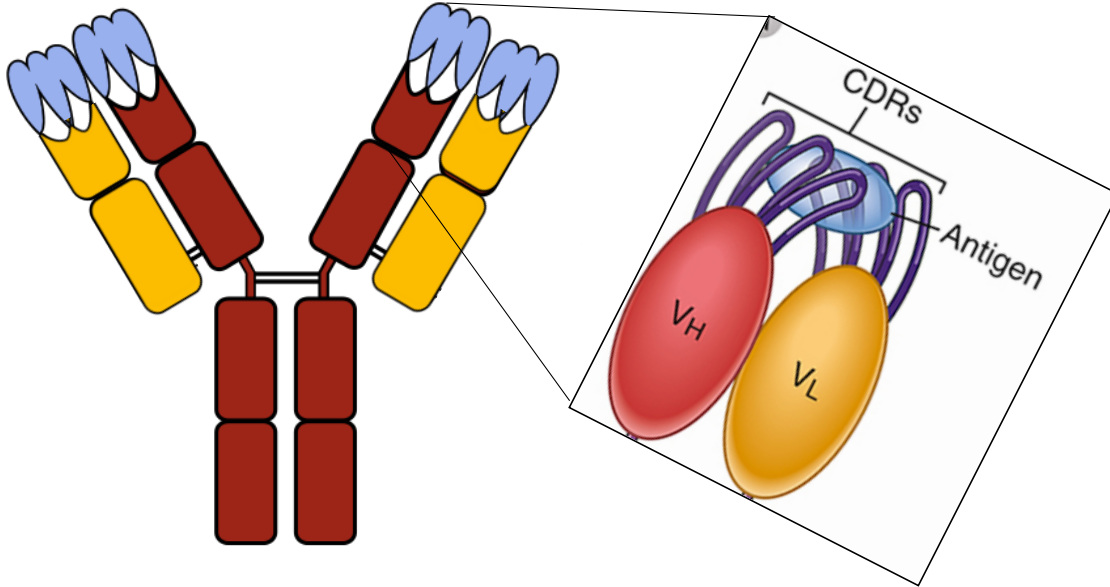


# The antigen- antibody interaction

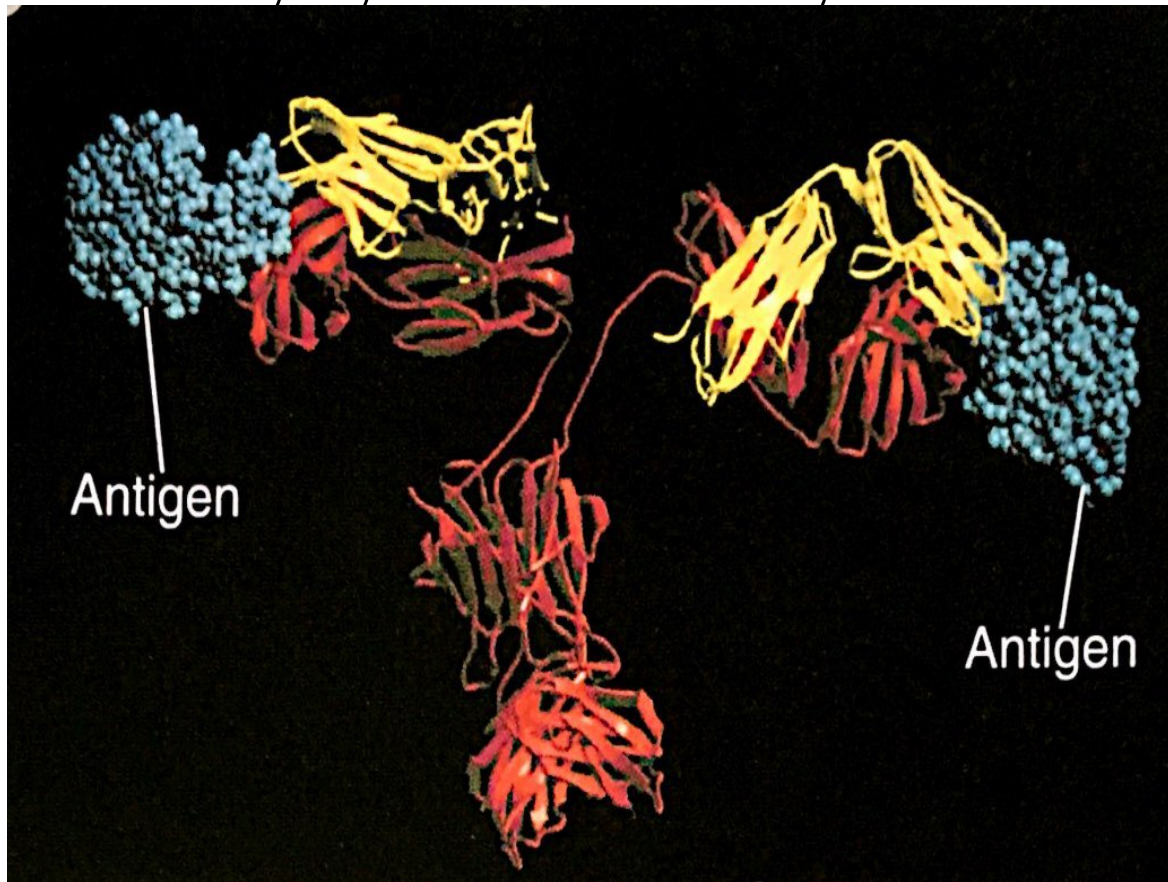
# Characterization of scFvs that bind lysozyme



- The goal of this screen is to find a scFv clone with improved binding to lysozyme
- Antibody with a lower  $K_d$  for its antigen means a more stable interaction and a higher affinity
- We sorted a library of scFv yeast that bind to lysozyme
- Today will determine the dissociation constant of a single clone scFv with lysozyme

# CDRs generate antigen binding site specificity

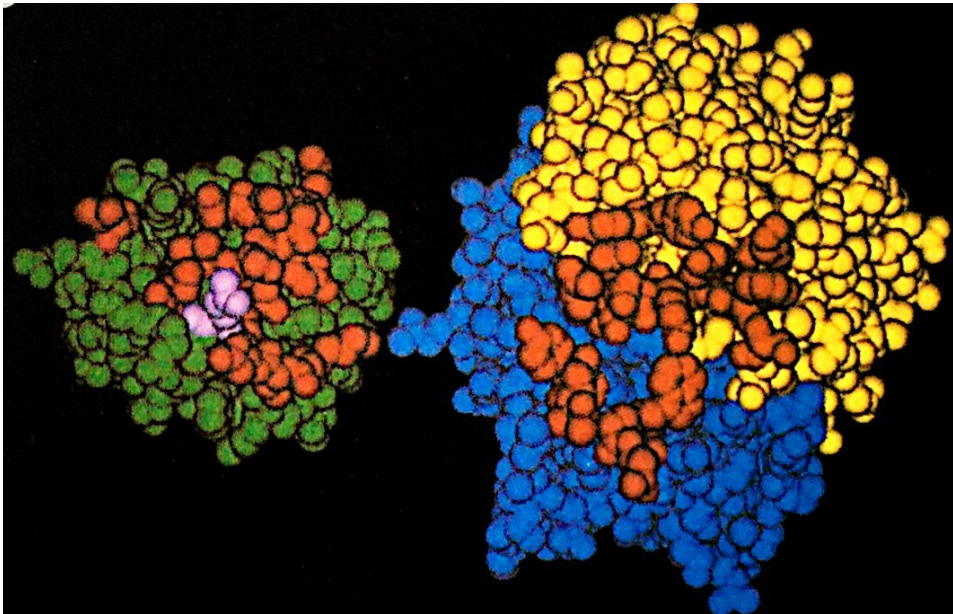
Lysozyme bound to antibody



- Specificity, degree to which an antibody differentiates between different antigens
- Finger-like CDRs usually recognize 15-22 amino acids
- Basic antibody structure maintained ( $\beta$  strands) when variability confined to CDR loops

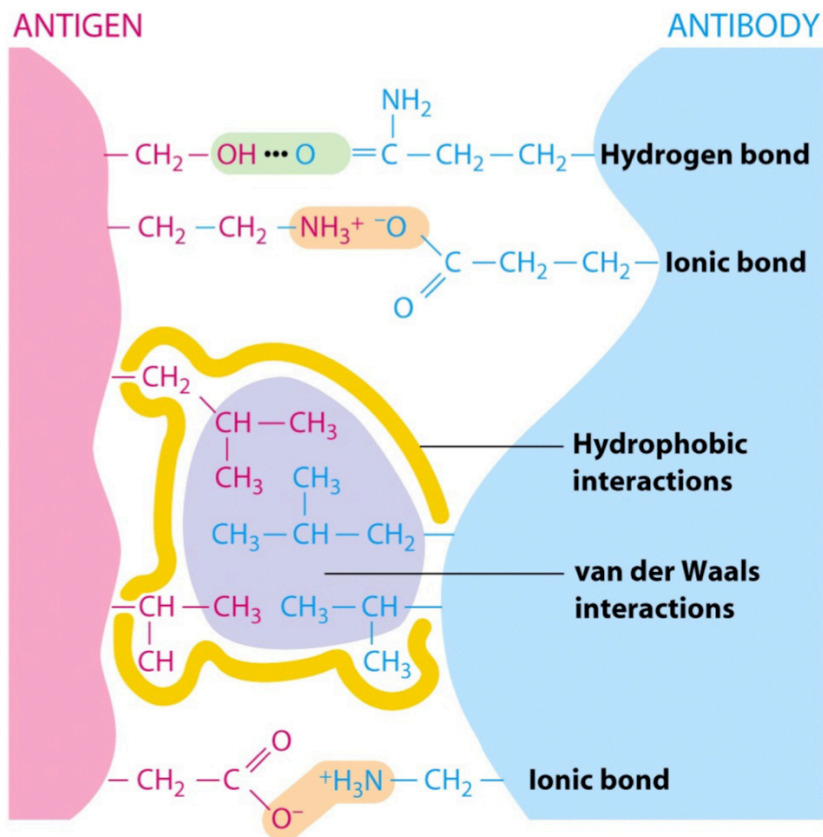
# The Antigen - Antibody interaction forms multiple contacts

3D: Lysozyme bound to variable region



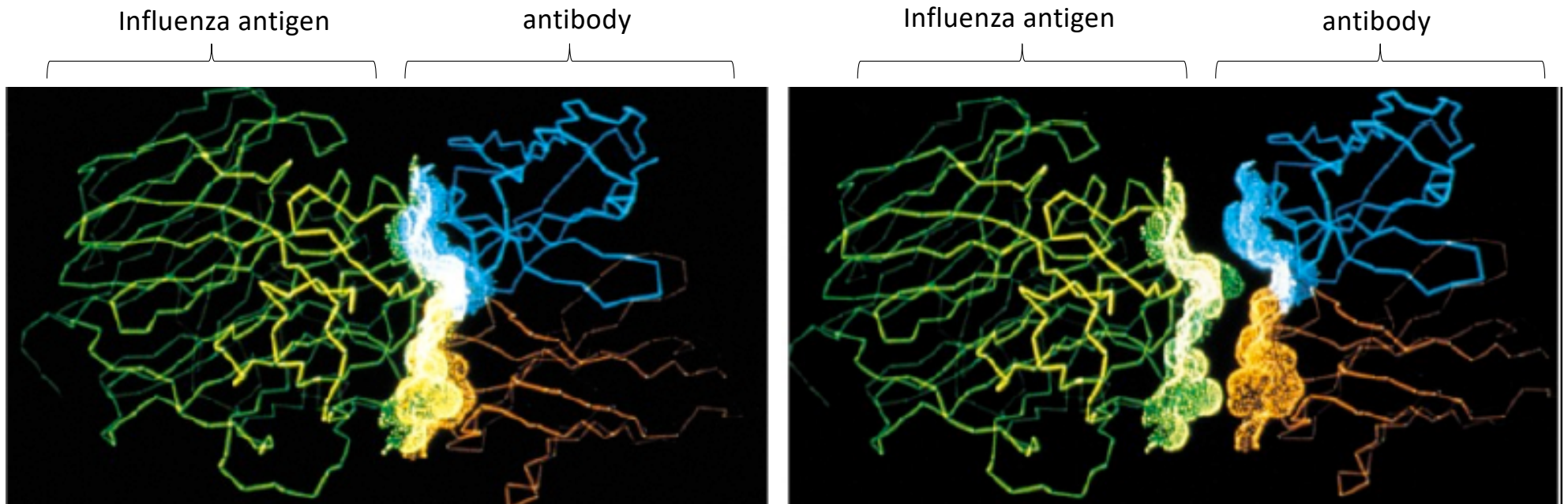
- Green: lysozyme
- Blue/Yellow:  $V_L$  and  $V_H$
- Red amino acids that interact
- Pink critical glutamine residue fits into cleft of CDR

# Noncovalent bonds form the basis of the antibody binding site

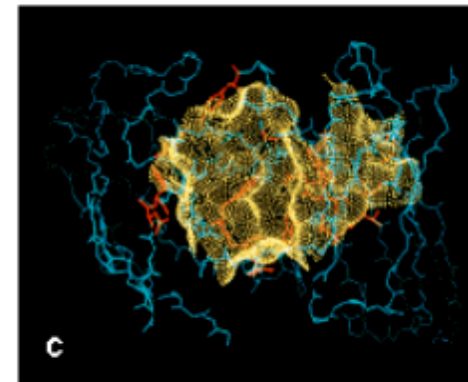
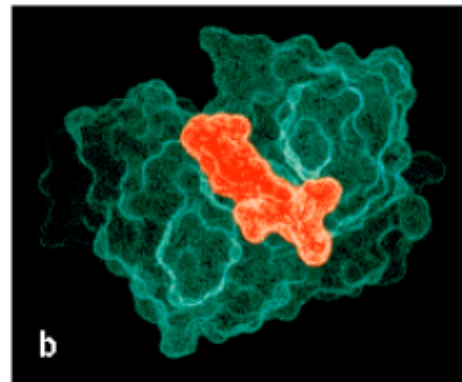
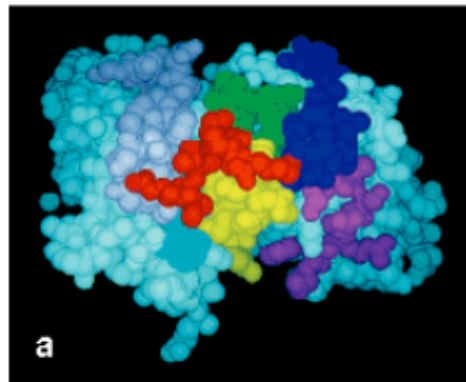
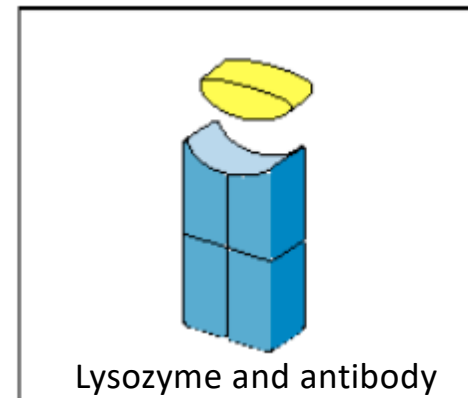
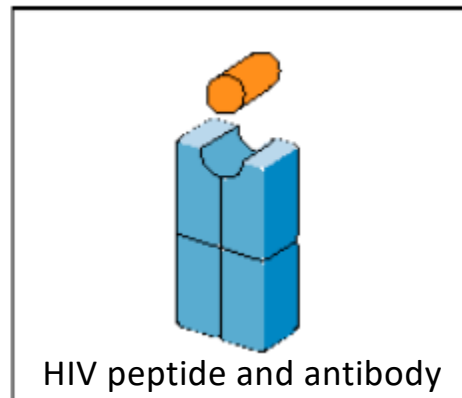
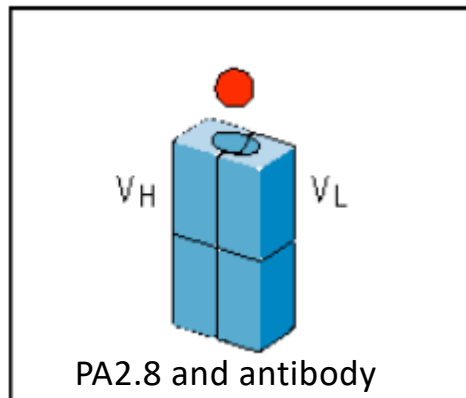


- Strength of each of these noncovalent interactions is weak
  - Many noncovalent bonds are required to form a strong interaction
- Each of these interactions operates over a very small distance ( $\sim 1 \text{ \AA}$ )
- This requires a high degree of complementarity between the CDR of the antibody and the antigen

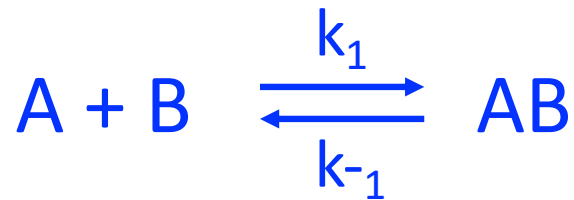
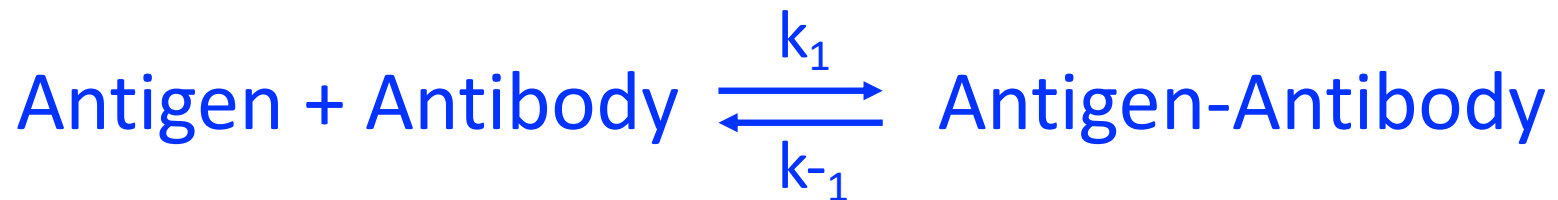
# Influenza antigen and antibody binding illustrates complementary when separated by 8 Å



# Large variation in antibody binding pockets



Binding a monovalent antigen by an antibody can be described by a bimolecular equation

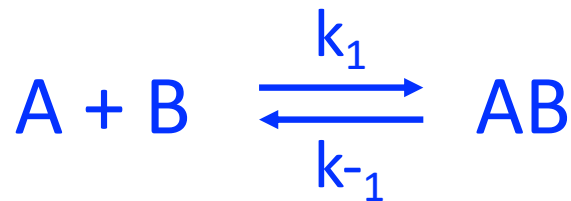



$k_1$ =rate of association

$k_{-1}$ =rate of disassociation



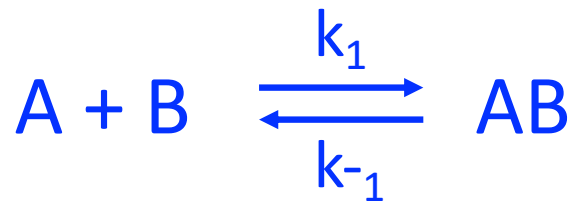
The equilibrium association constant ( $K_a$ ) is a good indicator for antibody affinity




$$K_a = \frac{[AB]}{[A][B]}$$


- Ratio of products to reactants
- Affinity, the strength of the total noncovalent interactions between one antigen and antibody
- Units of  $K_a$  are concentration<sup>-1</sup>
- Example: nM<sup>-1</sup>

Equilibrium dissociation constant ( $K_d$ ) is an indicator of the stability of a complex



$$K_d = \frac{[A][B]}{[AB]}$$


- Ratio of reactants to products
- Antibodies produced in a typical immune response usually varied from  $K_d = 10^{-7}$  (~100nM) to  $10^{-9}$  (~1nM)
- Units of  $K_d$  are concentration
- The smaller the  $K_d$  the more stable the interaction

## Prof. Koehler Mod1 Lecture 4

### Range of biologically important interactions

Antibody-antigen  
interactions

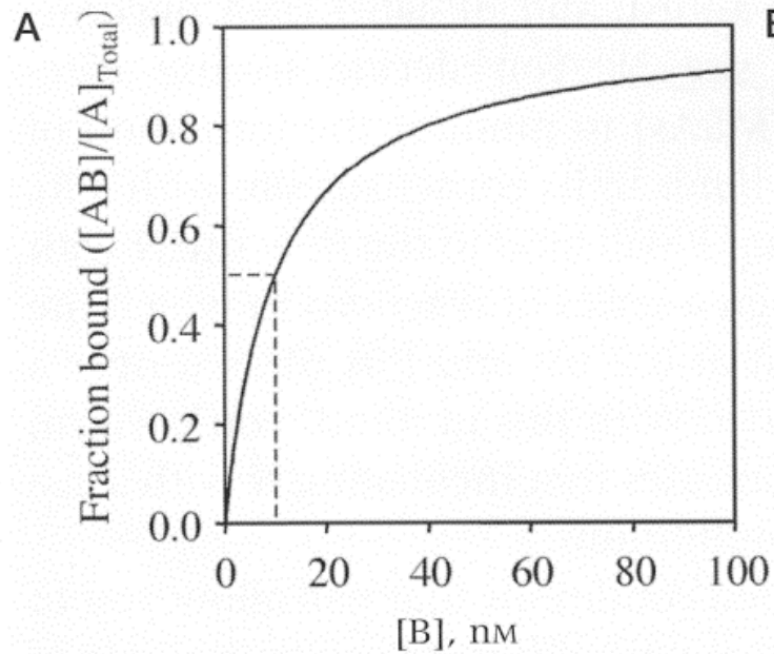
Type of Interaction	$K_D$ (molar)	$\Delta G_{bind}^0$ (at 300K) kcal/mol
Enzyme:ATP	$\sim 1 \times 10^{-3}$ to $\sim 1 \times 10^{-6}$ (millimolar to micromolar)	<b>-4 to -8 kcal/mol</b>
signaling protein binding to a target	$\sim 1 \times 10^{-6}$ (micromolar)	<b>-8 kcal/mol</b>
Sequence-specific recognition of DNA by a transcription factor	$\sim 1 \times 10^{-9}$ (nanomolar)	<b>-12 kcal/mol</b>
small molecule inhibitors of proteins (drugs)	$\sim 1 \times 10^{-9}$ to $\sim 1 \times 10^{-12}$ (nanomolar to picomolar)	<b>-12 to -17 kcal/mol</b>
biotin binding to avidin protein (strongest known non-covalent interaction)	$\sim 1 \times 10^{-15}$ (femtomolar)	<b>-21 kcal/mol</b>

higher  $K_D$  value  
weaker interaction

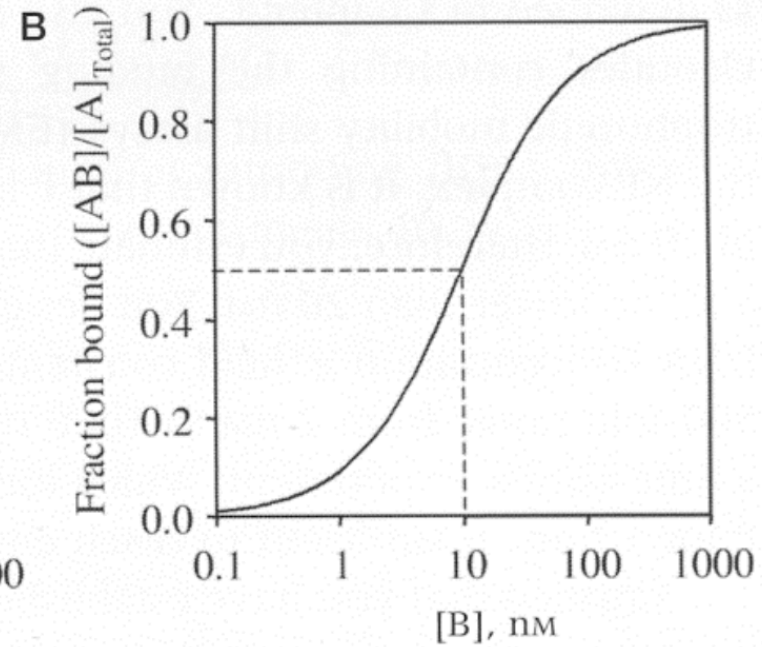
lower  $K_D$  value  
stronger interaction

# Logarithmic vs. Linear display of data

*historic convention*

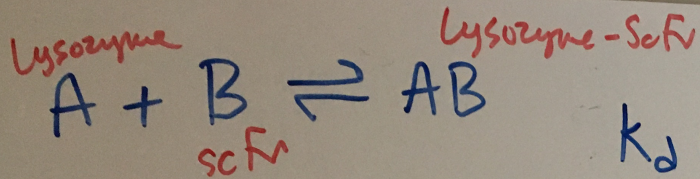


*current convention*



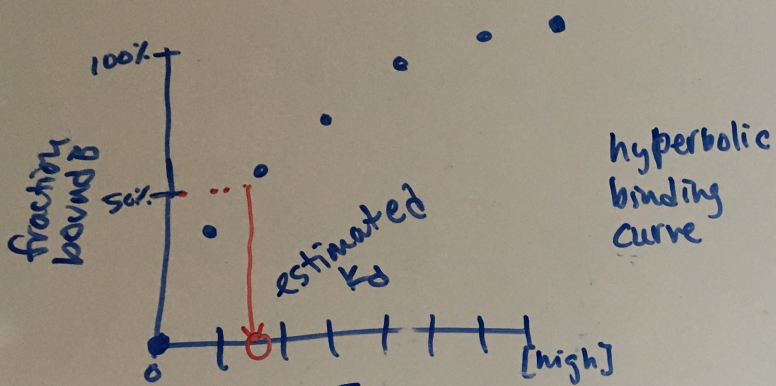
**Biomolecular binding interaction at equilibrium:**  
Why is antibody dissociation constant ( $K_d$ ) equal to the antigen concentration at which 50% antibody is bound to antigen?





Lysozyme-ScFv

$$K_d = \frac{[A][B]}{[AB]}$$



$$\text{fraction bound B} = \frac{[AB]}{[B] + [AB]} = \frac{1}{\frac{[B]}{[AB]} + 1} = \frac{1}{\frac{K_d + [A]}{[A]}} = \frac{[A]}{K_d + [A]}$$

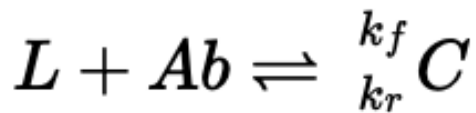
$$\frac{[B]}{[AB]}$$

$$K_d = \frac{[A][B]}{[AB]} \Rightarrow \frac{1}{\frac{[A]}{[AB]}} = \frac{[B]}{[AB]}$$

$$\text{fraction bound B} = \frac{[A]}{K_d + [A]} = \frac{1}{2} \text{ or } 50\%$$

$$K_d = [A]$$

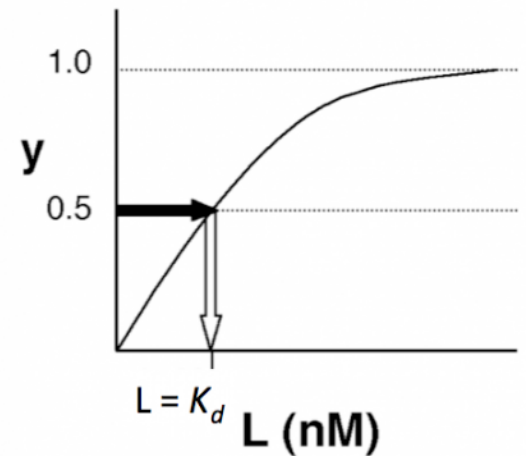
Mathematical relationship between fraction bound and free reactant makes estimations easy



$$y = \frac{[L]}{[L] + K_d}$$

If  $L$  in excess (in solution), and  $[L] = L$  constant

- at  $L = K_d$   $y = 0.5$
- if  $L \ll K_d$  then  $y \approx \frac{[L]}{K_d}$  (linear relationship)
- if  $L \gg K_d$  then  $y \approx 1$  (at saturation)





# Alternative methods to measure binding dynamics without necessitating equilibrium

Binding of the antibody to the antigen alters the resonance readout and can translate to affinity

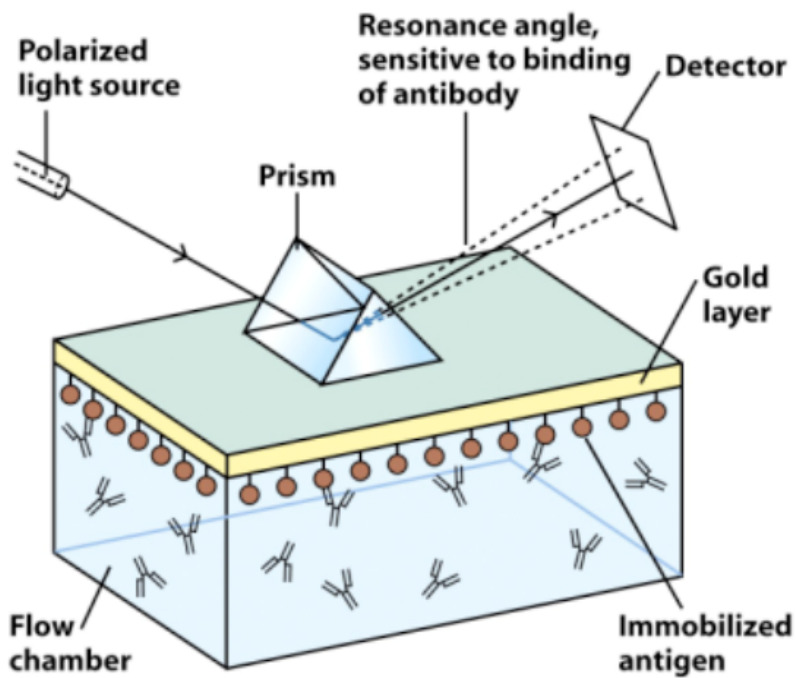
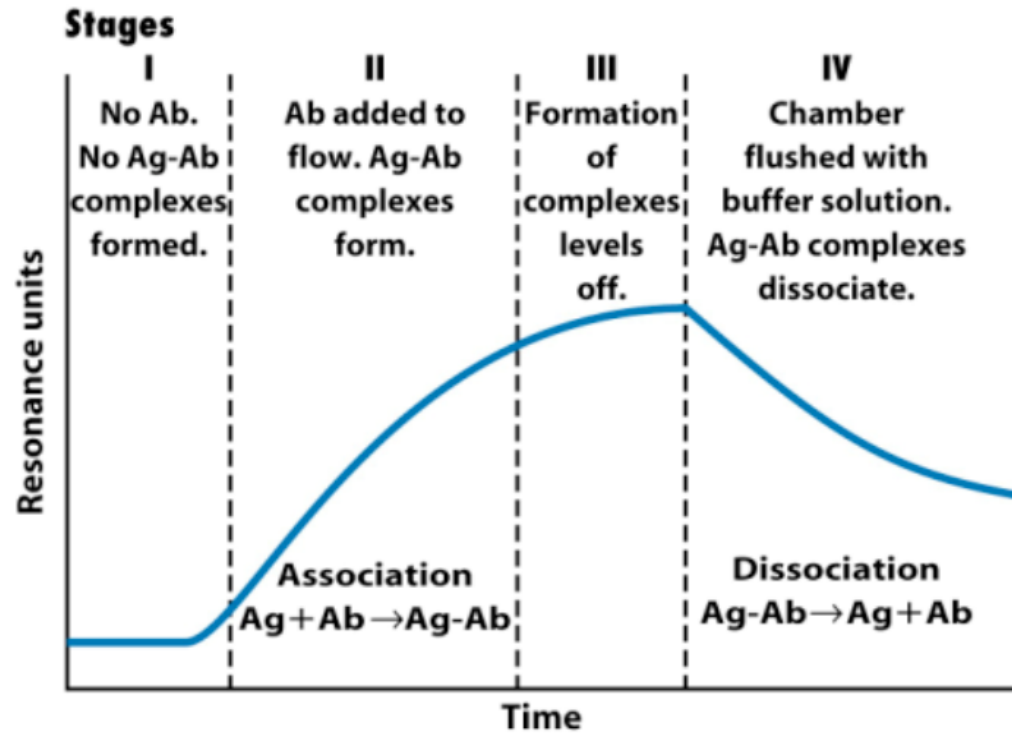
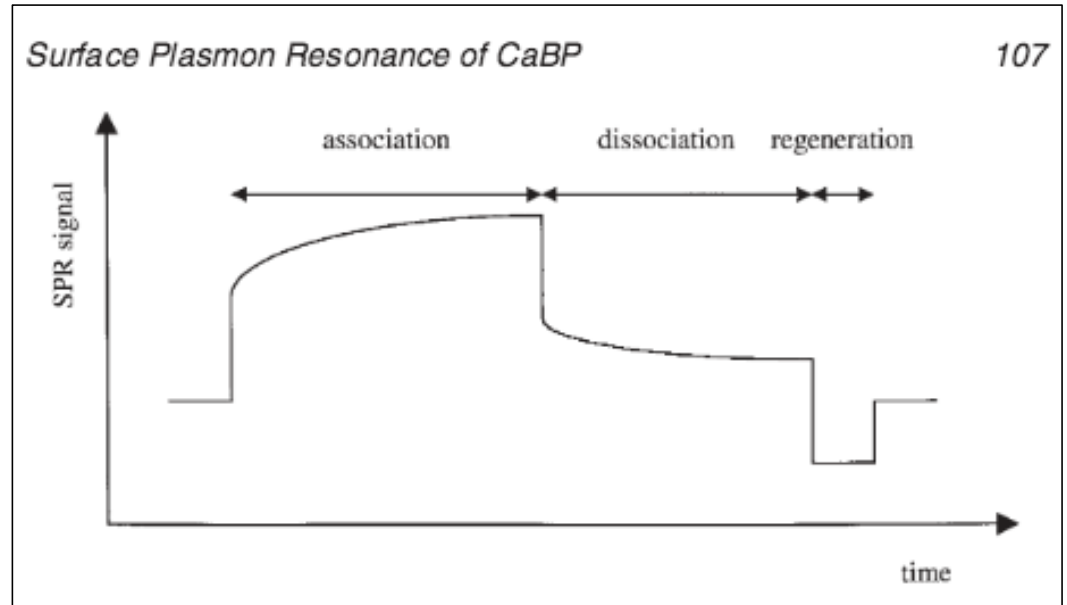
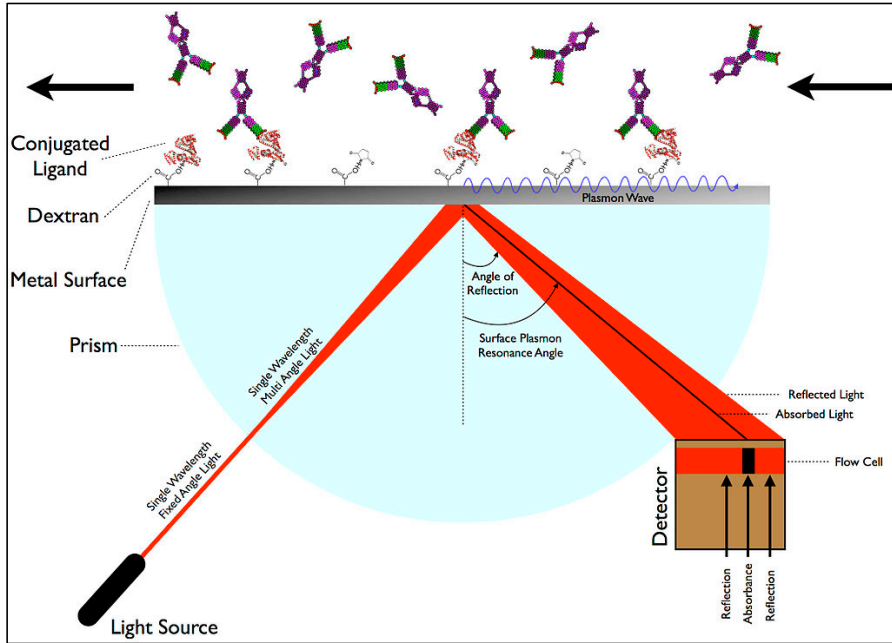


Figure 6-4a  
Kuby IMMUNOLOGY, Sixth Edition  
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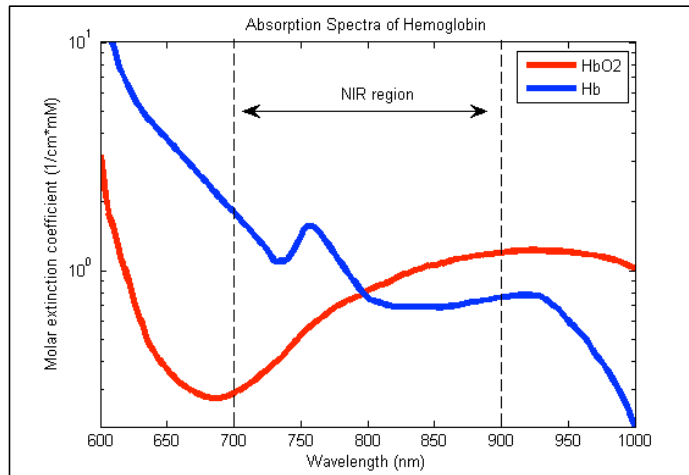


# Surface plasmon resonance (Biacore)

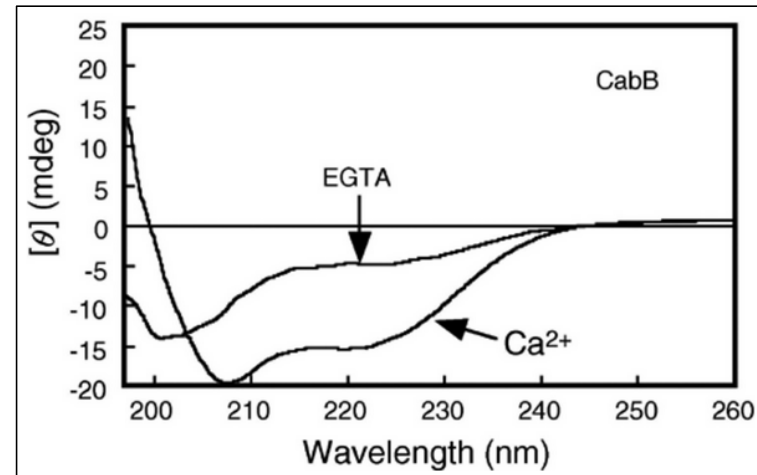


# Binding may be quantified using methods other than fluorescence

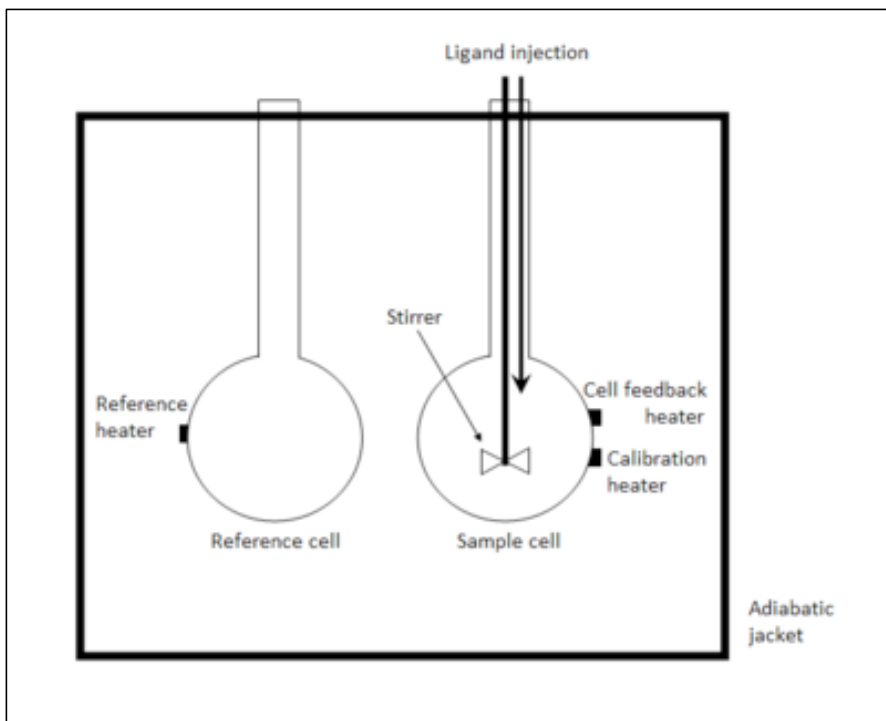
- absorbance spectroscopy  
*e.g.* hemoglobin binding to O<sub>2</sub>



- circular dichroism  
*e.g.* Ca<sup>2+</sup> binding to CabB

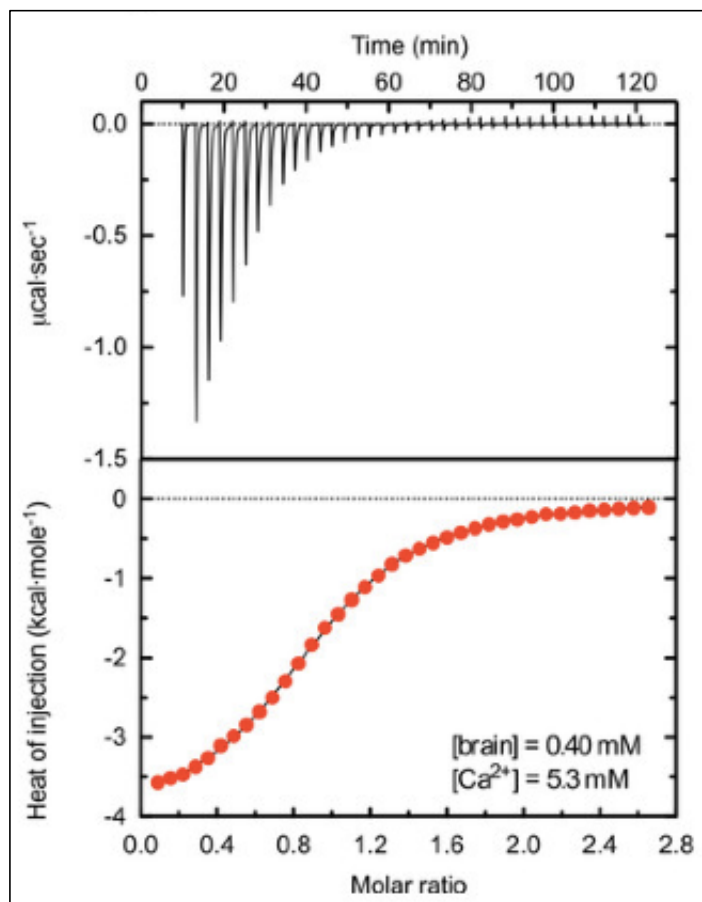


# Isothermal titration calorimetry measures thermodynamic parameters of interactions



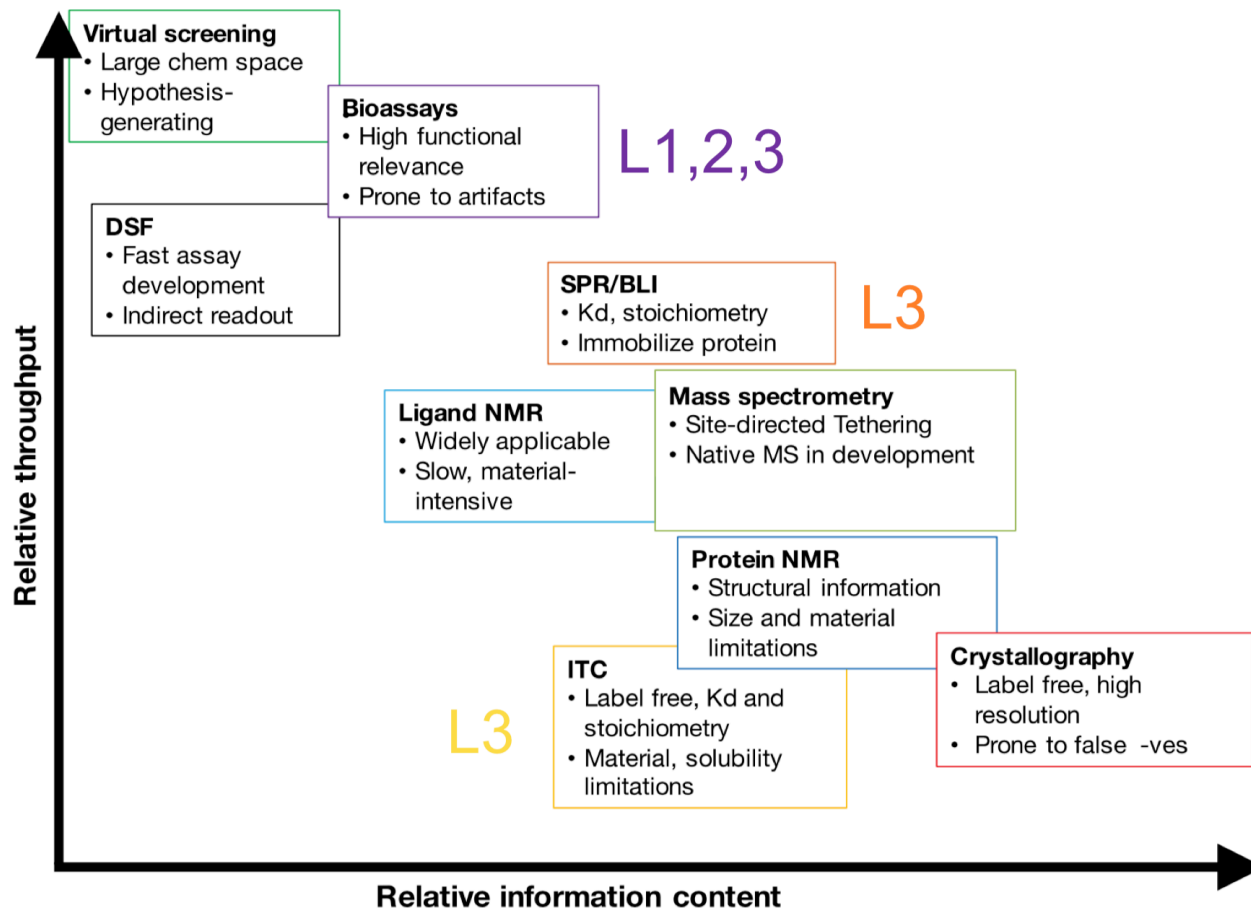
$$\Delta G = -RT \ln K_a = \Delta H - T\Delta S$$

Backman (2015) *PeerJ* 3: e944



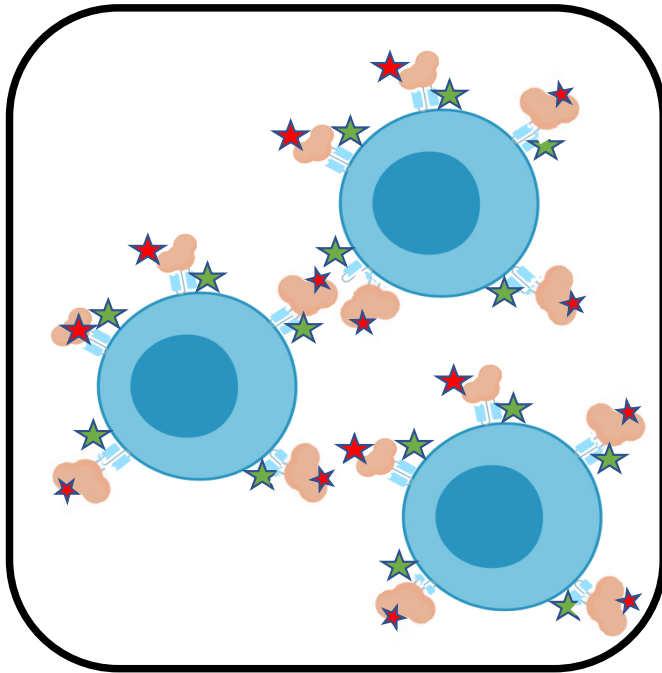
## Prof. Koehler Mod1 Lecture 4

# Methods to evaluate binding interactions

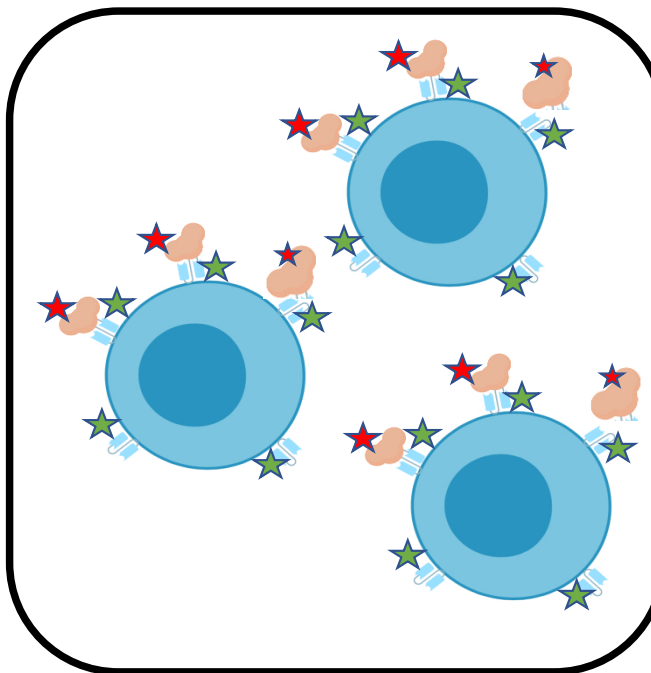


Practically: how will we measure equilibrium binding with different antigen concentrations?

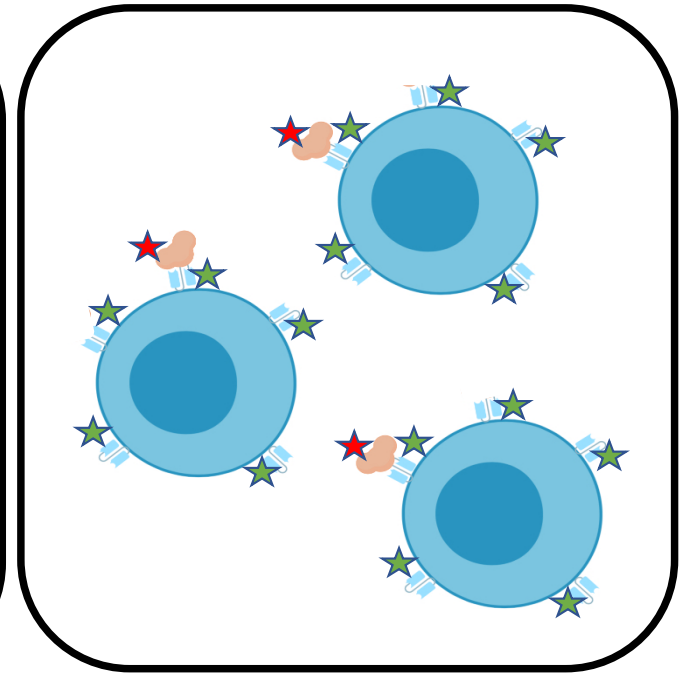
Tube #1



Tube #2

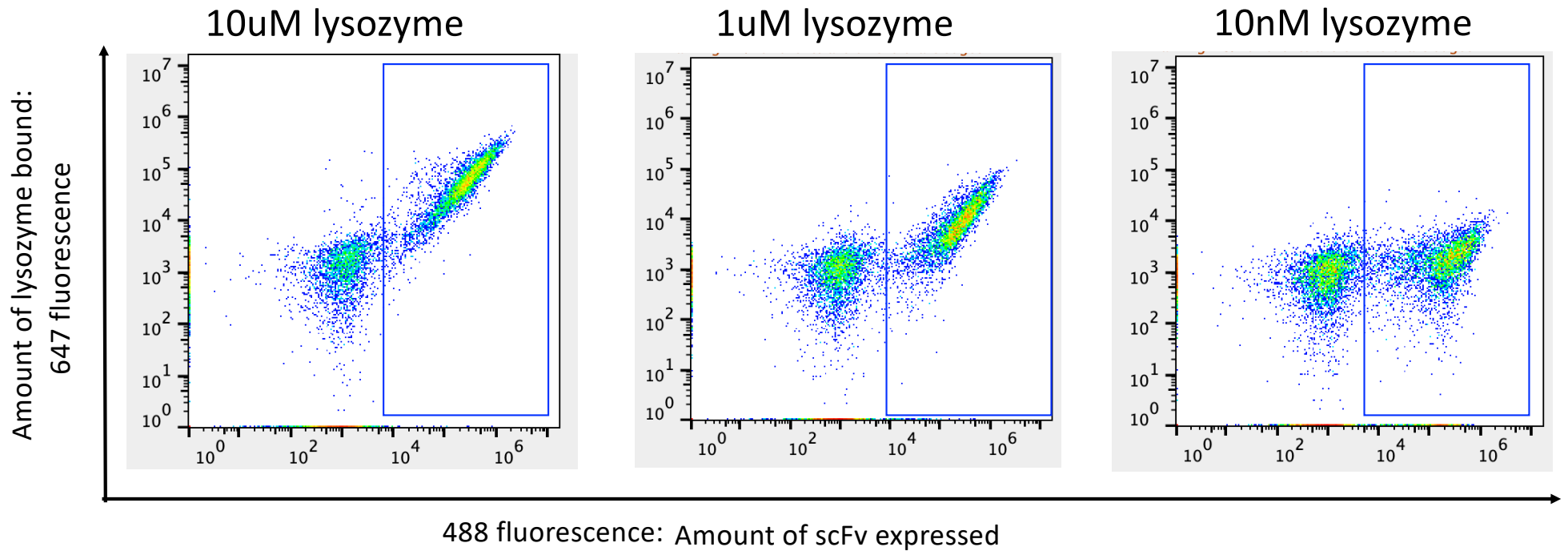


Tube #3

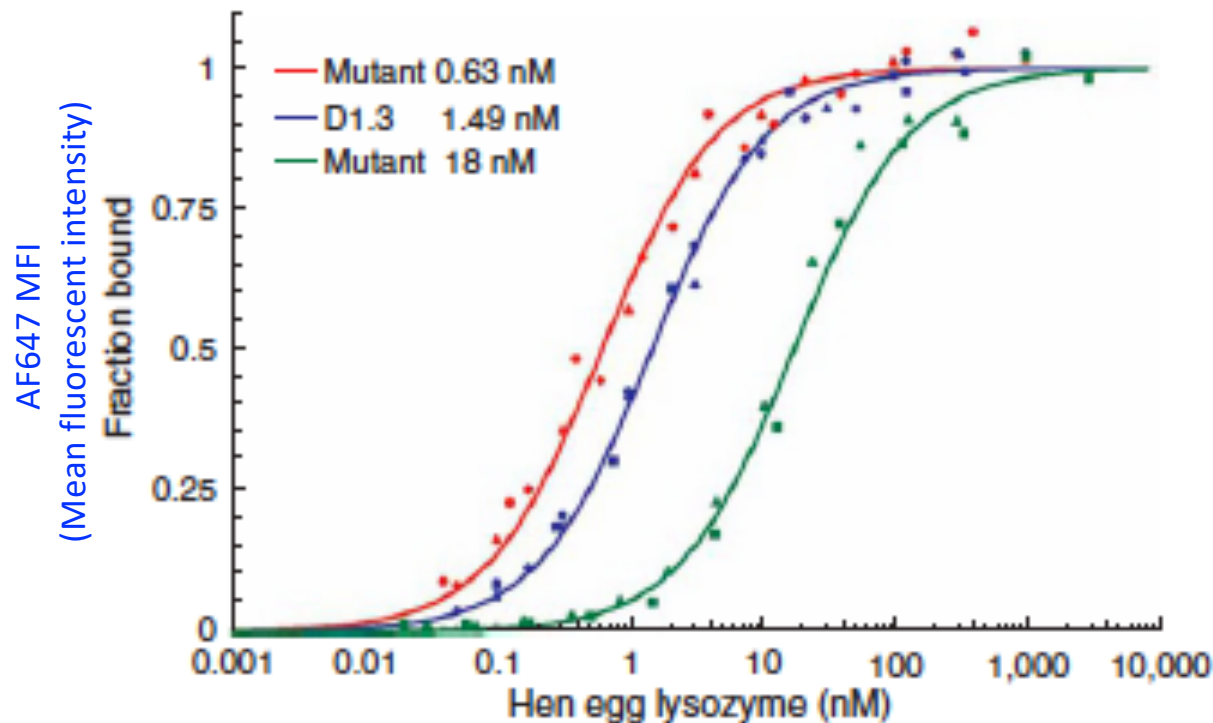


# Mean fluorescent intensity of gated scatterplot is to fraction bound

scFv Clone 14989 (650nM  $K_d$ ) incubated with:



Plotted MFI illustrates fraction of antigen bound to antibody

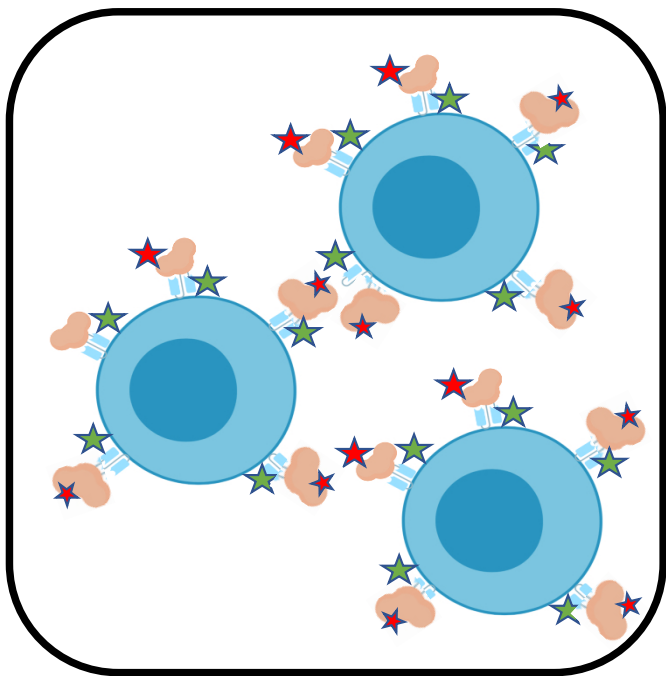


Do you agree with the  $K_d$ ?



# Today in “lab”

1) Set up titration of equilibrium binding reactions



2) Analyze flow cytometry data

