

# M3D1:Growth of phage materials

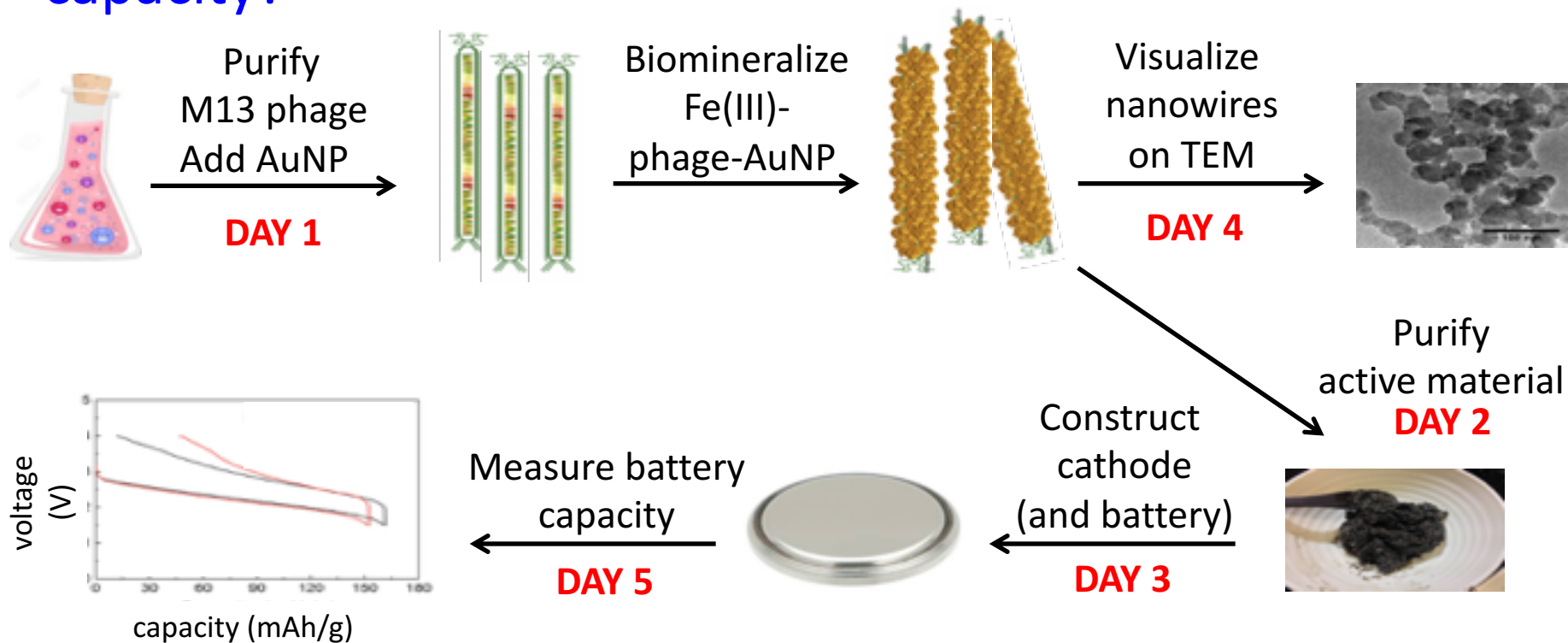
1. Purify M13 bacteriophage (phage)
2. Prelab during 60min incubation
3. Finish M13 purification and measure concentration
4. Incubate phage with nanoparticles (AuNP)



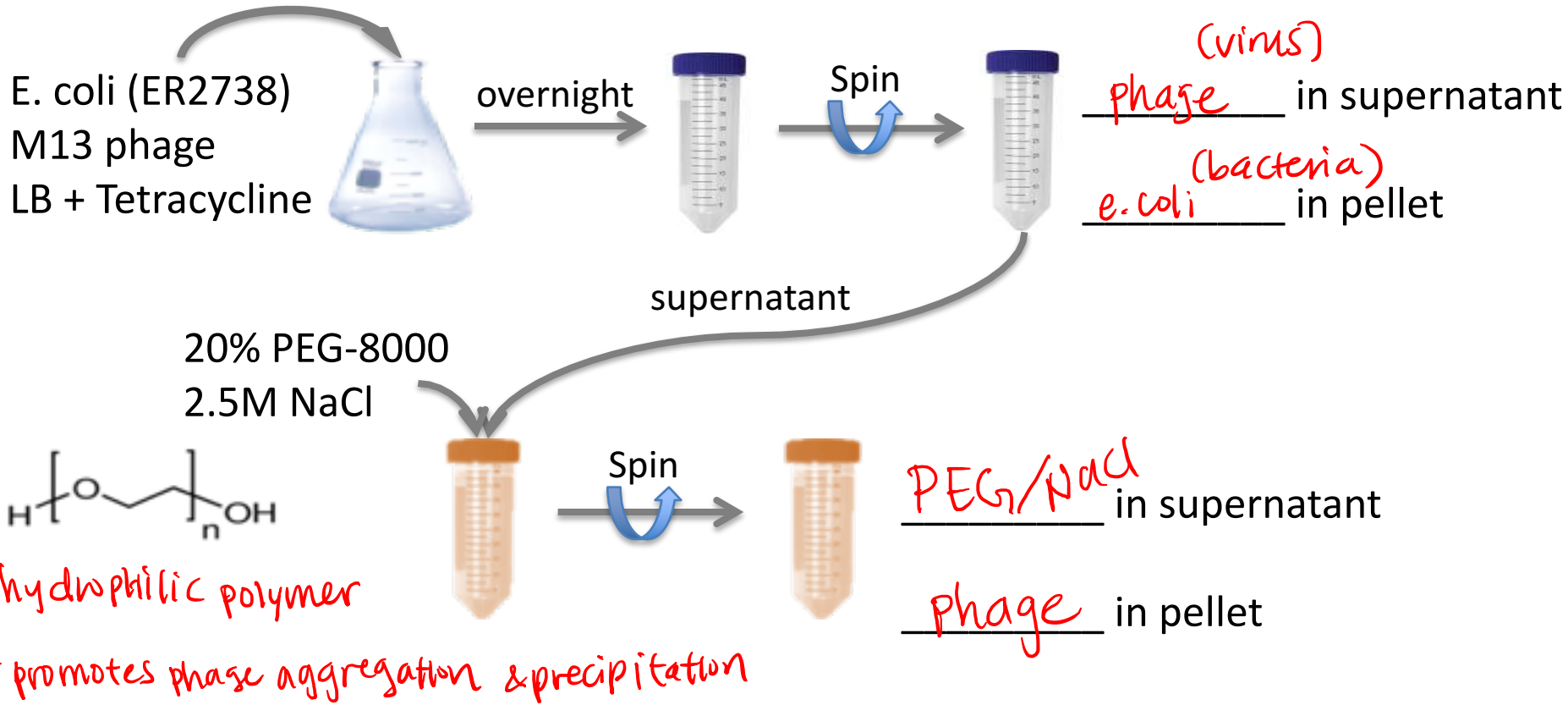
Thank you, Jifa Q. (Belcher Laboratory)!

# Module 3: biomaterials engineering

## How do nanoparticle size and quantity affect battery capacity?



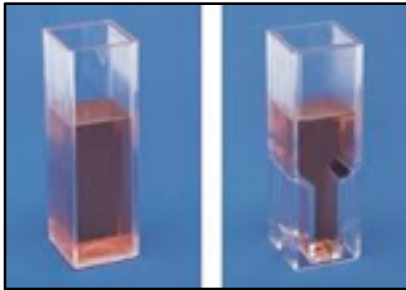
# Phage purification using polyethylene glycol (PEG) in 2.5M NaCl



# Determining phage titer (number of virus):



- By plating: plaque assay
  - Phage slows *E. coli* growth = plaque (cleared zone)
  - Plaque-forming units: PFU/mL



- By spectrophotometry

$$\# \text{ phage / mL} = \frac{(6 \times 10^{16}) (A_{269} - A_{320})}{\# \text{ bases in phage genome}}$$

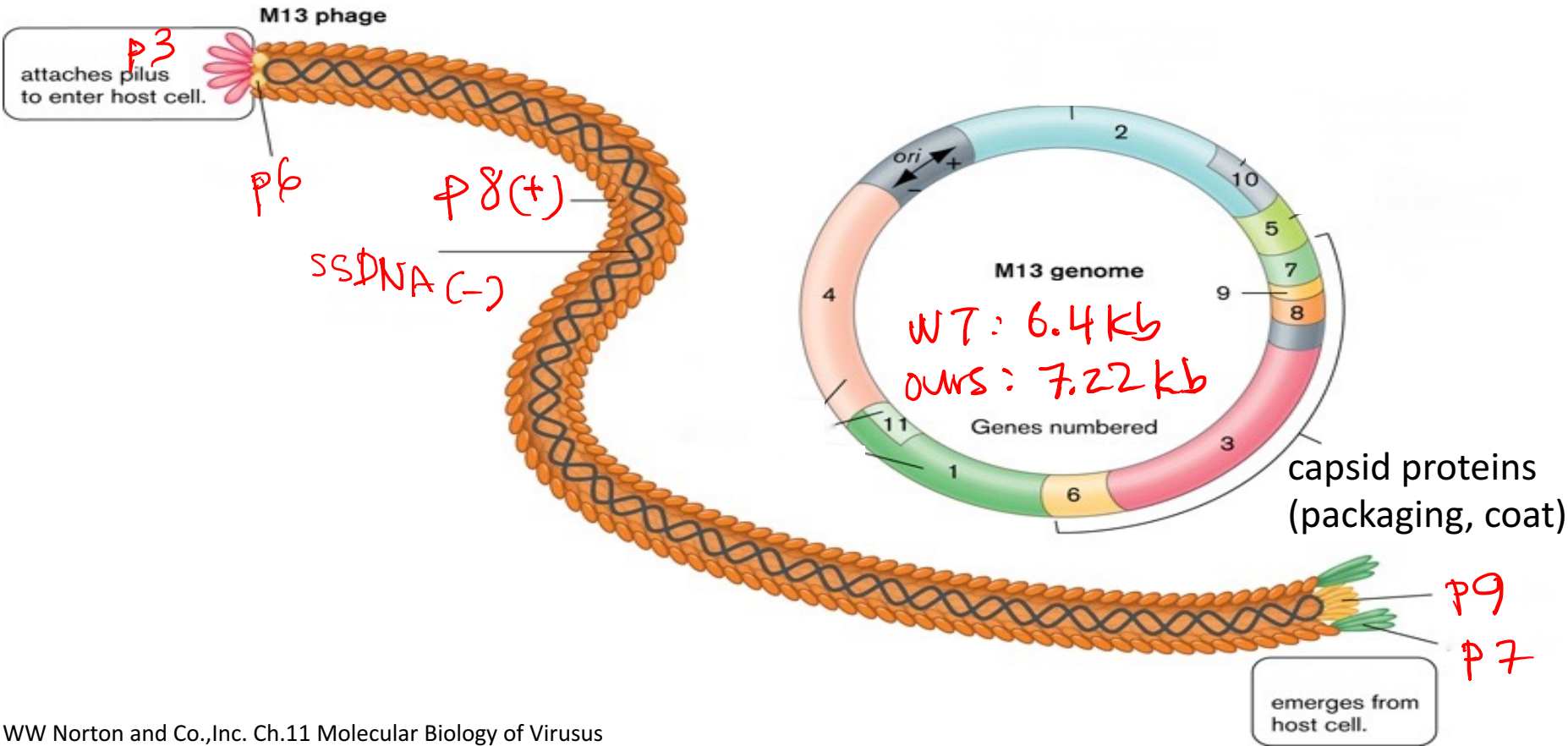
\* dilution factor

❖ Quartz cuvettes are expensive!

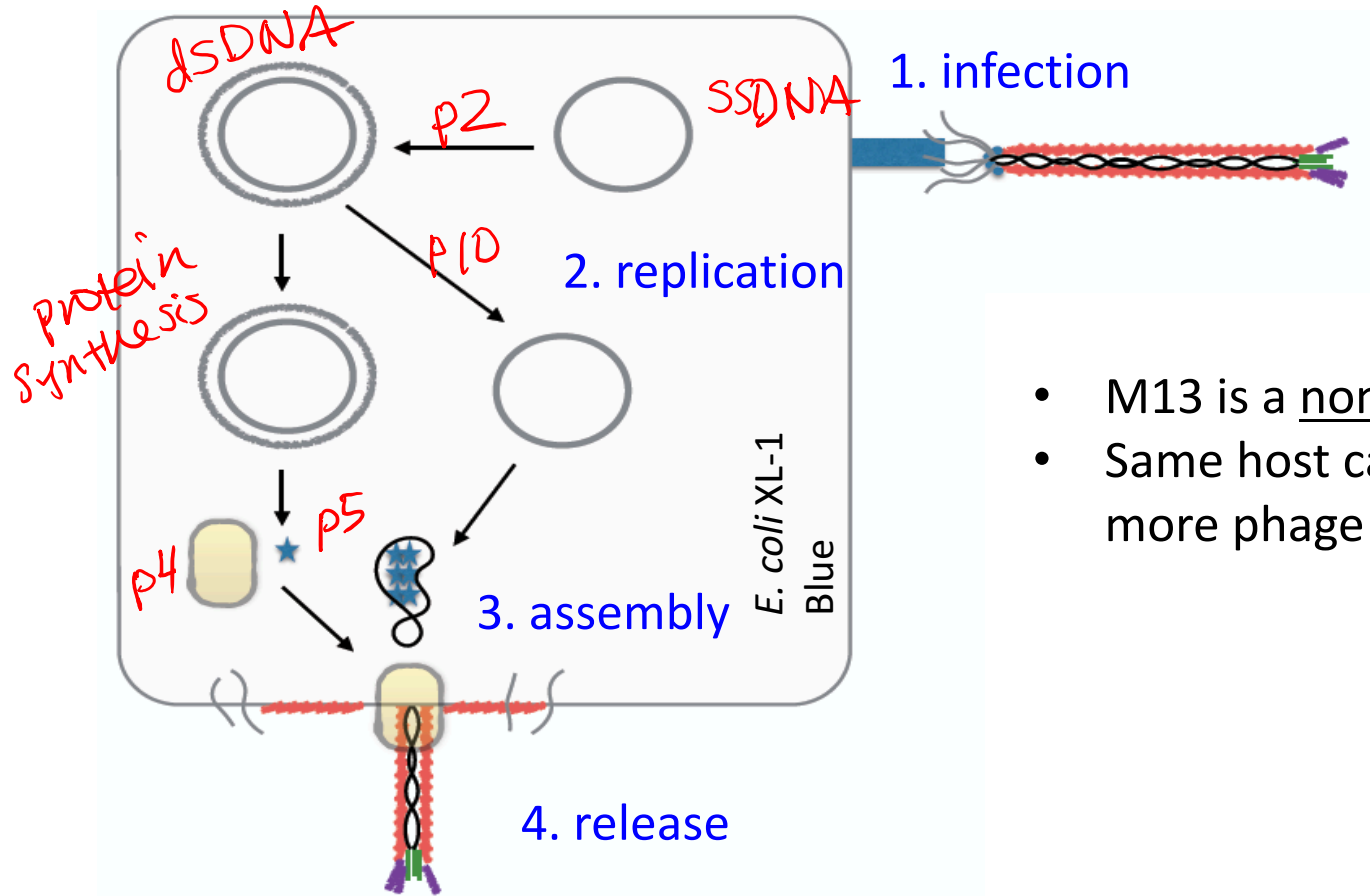
# M13 is a high aspect ratio phage

## coated in proteins encoded by ssDNA loop

*~ 900 nm long*  
*~ 6 nm wide*



# M13 virus life-cycle has four essential steps

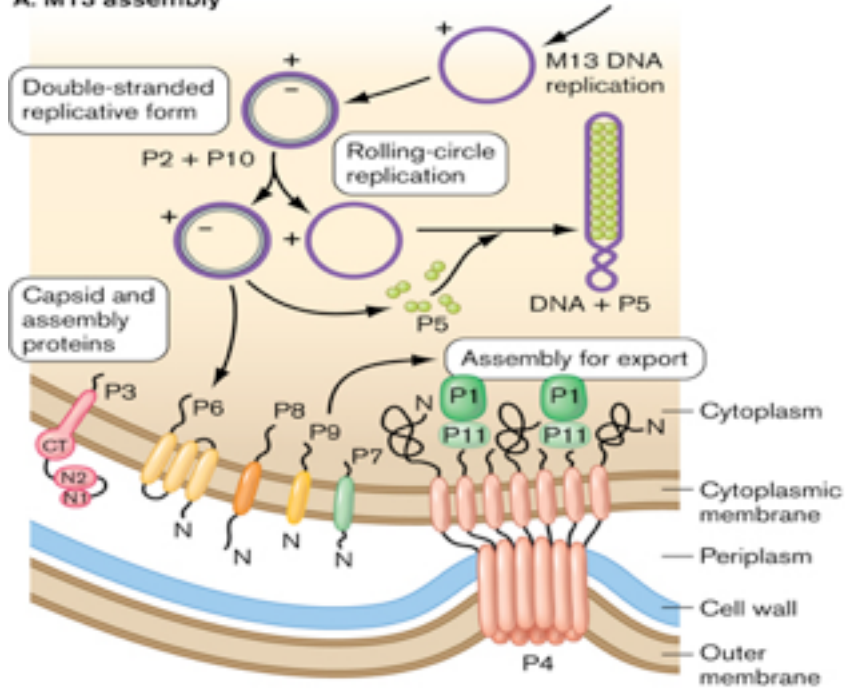


- M13 is a nonlytic phage
- Same host can keep producing more phage

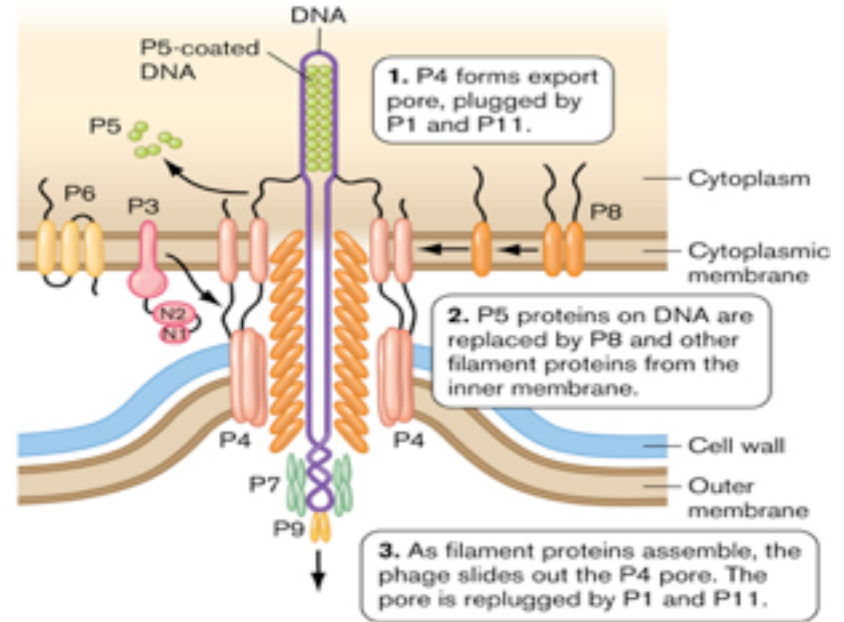
# M13 is a nonlytic bacteriophage

(so we can easily get lots of it)

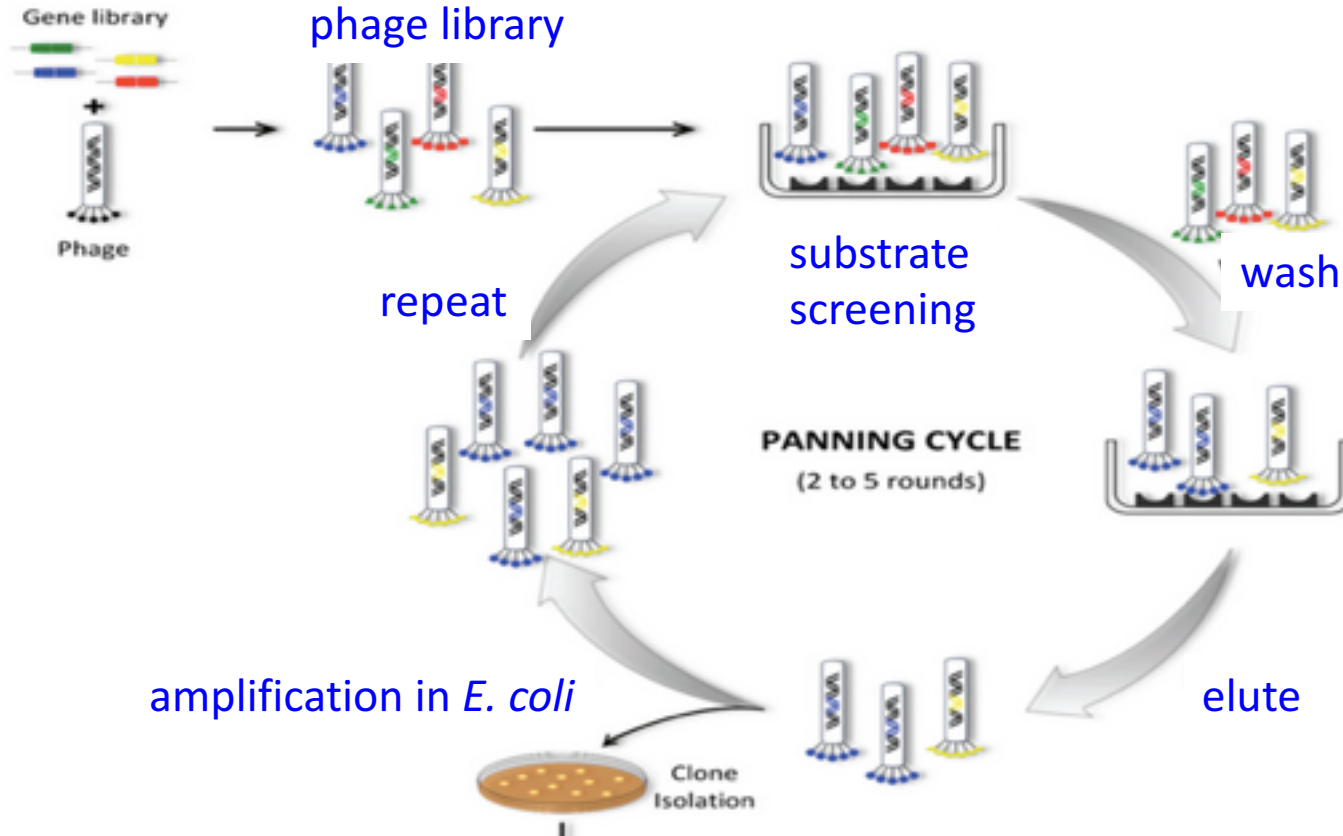
A. M13 assembly



B. M13 export



# Phage display allows agnostic selection of useful peptide sequences (typically binding)

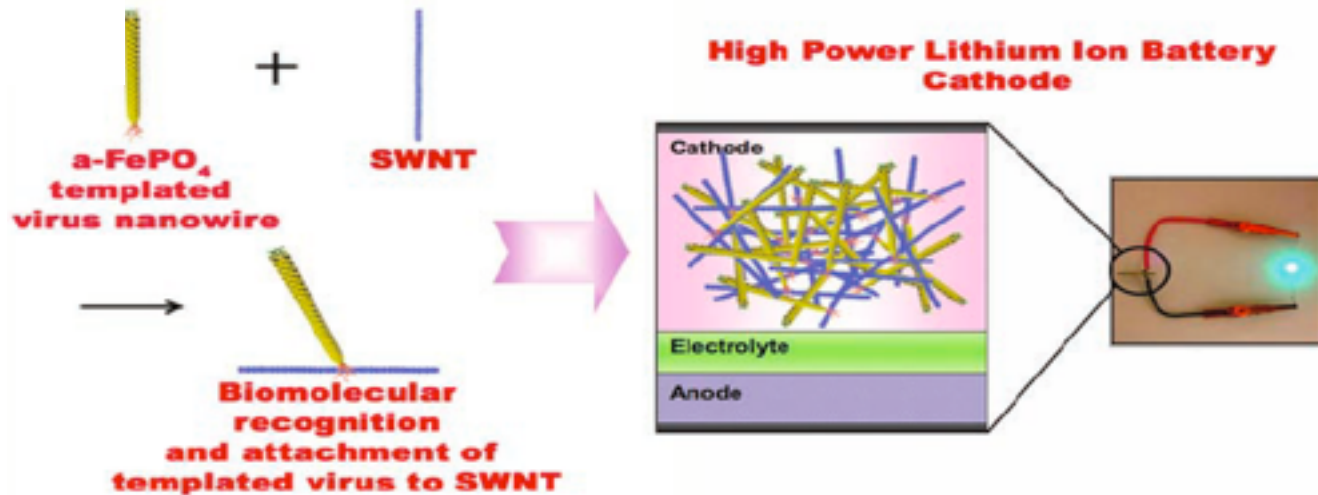




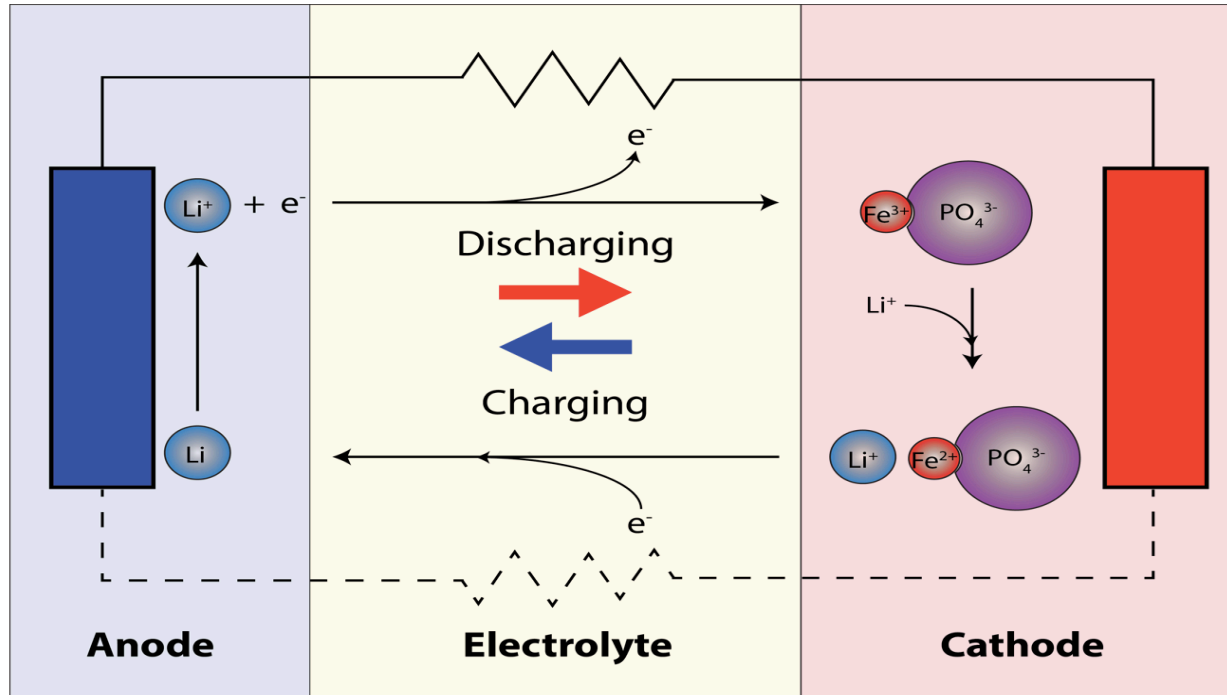
# M13 are engineer-able biomaterials

*negatively charged*

- Our p8 coat protein was mutated to contain sequence **DSPHTELP**
- Modified p8 proteins bind single wall carbon nanotubes (SWCNT), iron, gold, and other cationic metals
- Example of this virus in literature (Science, 2009):



# M13 nanowires as battery cathode



Cathode needs to be a good conductor of:

- ions
- electrons

# You will make a “gold standard” battery and an experimental battery

- Gold standard: 3.8 nm AuNPs, 40 AuNP/phage
- Experimental: choice of sizes & quantities
  - Size: 3.8 nm, 5 nm, 9 nm diameter AuNP (can mix them)
  - Quantity of AuNPs
  - Constraint: up to 50 mL total volume (phage + NPs)

# Considerations for experimental battery: nanoparticle material and size

- Surface area to volume ratio of AuNPs
  - Conductivity (consider volume of Au in cathode)
  - More surface area may be beneficial if Au has a catalytic function (Au may facilitate intercalation of  $\text{Li}^+$  in  $\text{FePO}_4$  cathode)
- Phage surface area available for Au and Fe binding
  - Too many AuNPs may reduce # binding sites for  $\text{FePO}_4$

Design with your lab partner. What is your **hypothesis**?

# You will make two flasks—one for each battery

## Gold standard



- 1)  $4 \times 10^{13}$  Phage
- 2) 3.8 nm Au NPs  
(40 NPs/phage)
- + 3) Water

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Final volume 50 mL

## Experimental



- 1)  $4 \times 10^{13}$  Phage
- 2) \_\_\_\_\_ nm Au NPs  
(\_\_\_\_\_ NPs/phage)
- + 3) Water

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Final volume 50 mL

# Today in lab

1. Finish phage purification
2. Calculate phage number
3. Begin construction of phage-NP-FePO<sub>4</sub> nanowires (2 flasks, one per battery)
  - **Choose Au NP size, quantity**

M3D2 HW: Describe **FIVE** recent findings that could potentially define an interesting research question.

- Formally cite the finding
- Write 3-5 sentences summarizing the finding

# RNA-seq vs. qPCR

## RNA-seq:

- Measures every expressed gene
- You enrich for mRNA
- Requires sequencing

## qPCR:

- Measures single gene
- No mRNA enrichment
- Does not require sequencing

**Q1:**

Why are you more likely to observe sequence from the 3' end of a gene in RNA-seq data (relative to sequence from the 5' end)?



Why are you more likely to observe sequence from the 3' end of a gene in RNA-seq data?

- What is special about the 3'-end? *↳ of mRNA only*

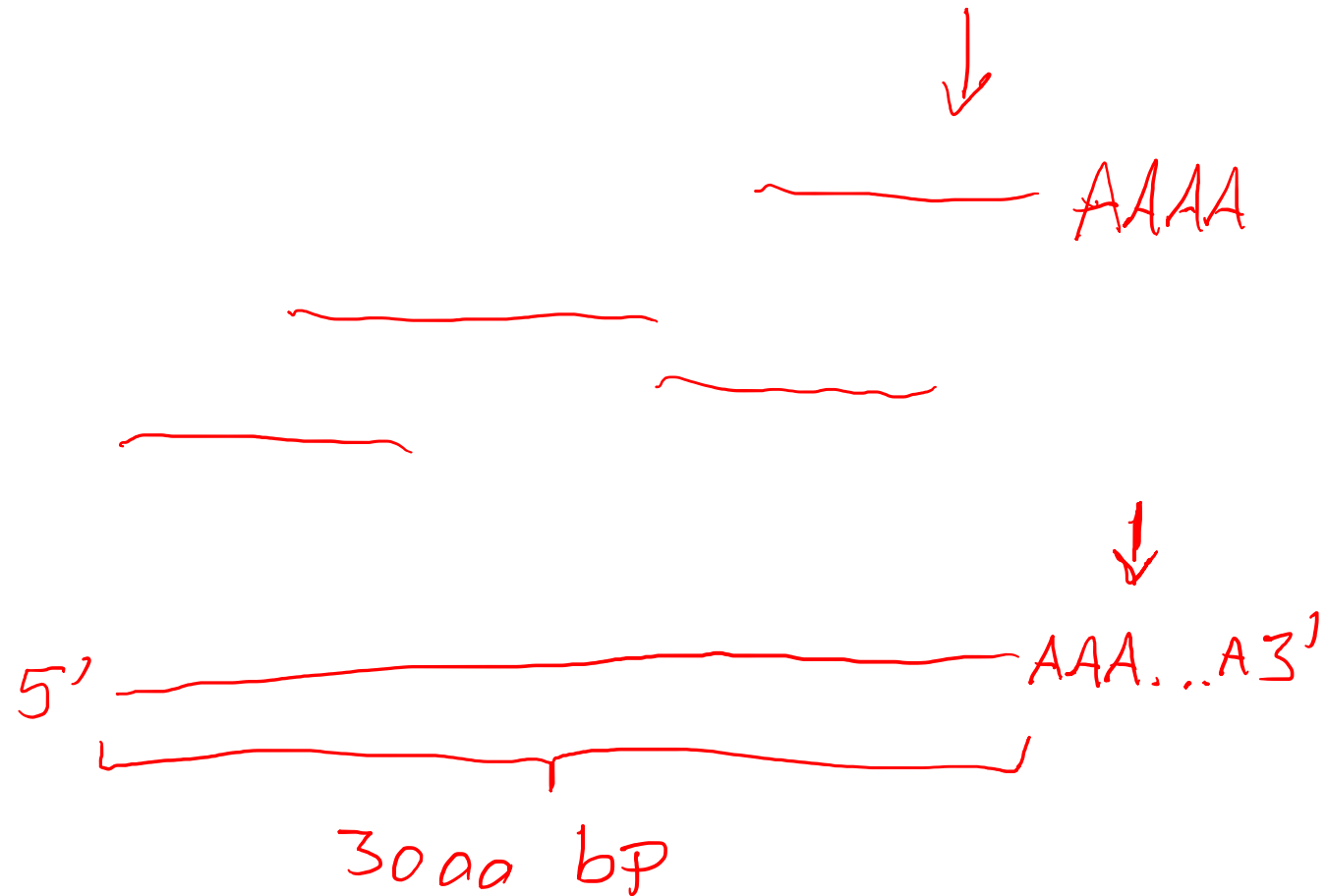
*Has a polyA-tail*

- What is special about mRNA?

*Has the  $\gamma$ (A) tail*

- How do you enrich for mRNA?

*Use oligo(dT)*



## Q2:

To compare two sets of RNA-seq data, you first normalize the results by calculating the RPKM value for each gene. What are the two factors to which you normalize (hint: how do you normalize between experiments AND how do you normalize between genes)?

# Calculating the RPKM

- RPKM = Reads Per Kilobase Million

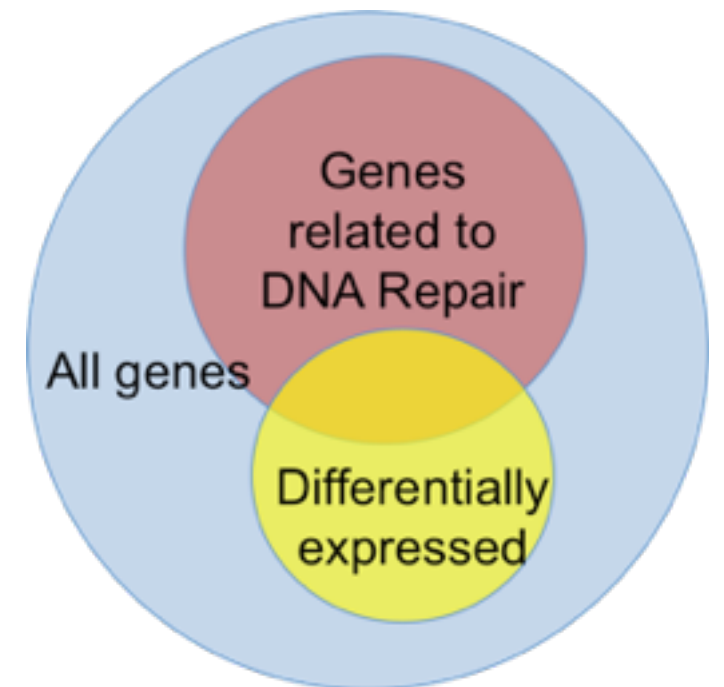
Total reads / 1,000,000 = **per million (PM)** scaling factor

Reads / **PM** = RPM

RPM / gene length in Kb = RPKM

**Q3:**

When analyzing RNA-seq data you identify a group of differentially expressed genes (yellow circle). You already know which genes are involved in DNA repair (red circle).w



# Which probability distribution will tell you the probability of overlap?

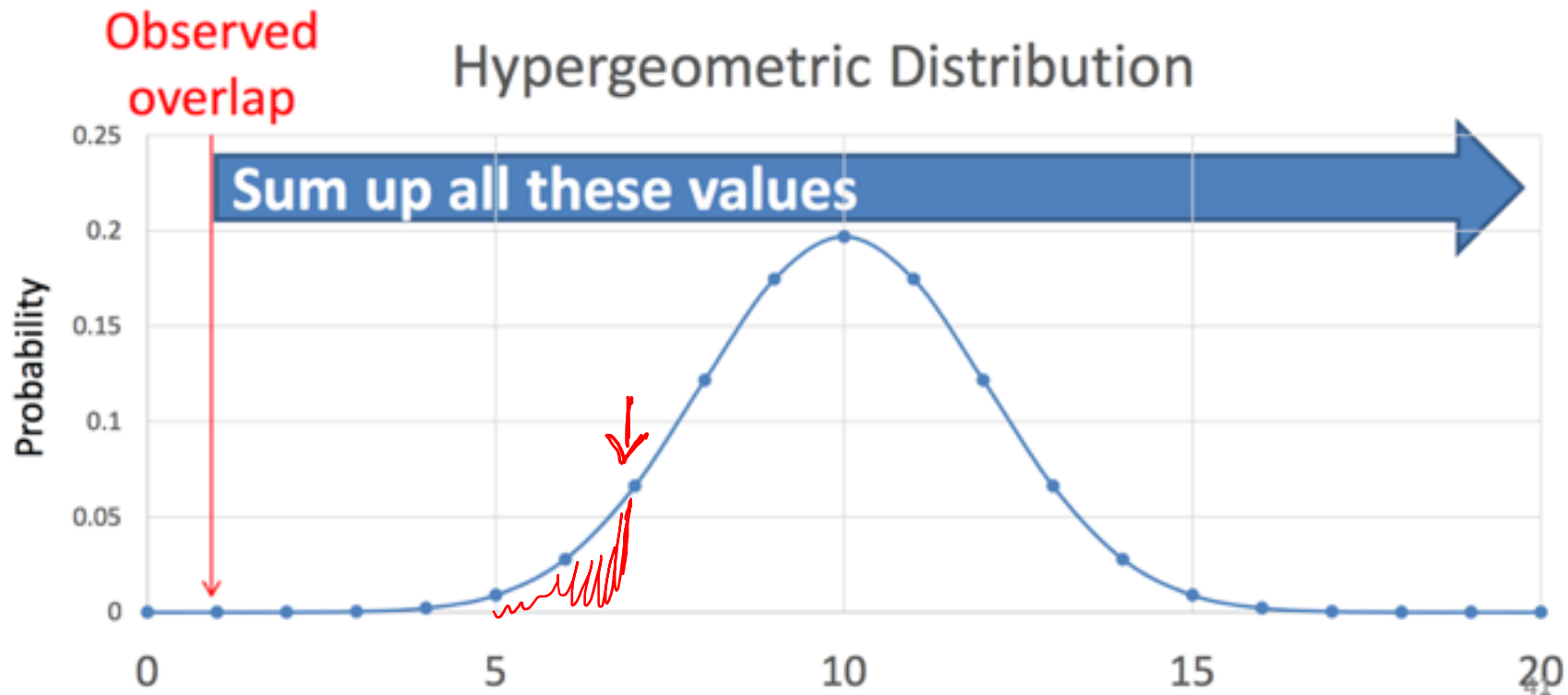
- Hypergeometric distribution

$$P(\text{overlap}) = \frac{\binom{DNA\ repair}{Overlap} \binom{Genome - DNA\ repair}{Diff.\ expr. - overlap}}{\binom{Genome}{Diff.\ expr.}}$$



# What statistical function can you use to test if the overlap is significant?

- Cumulative density function (CDF)
- Fisher's Exact Test



**Q4:**

qPCR is used to measure expression levels of specific genes.

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### Why measure p21?

It lets us track the cell cycle in response to DNA damage

### Why measure GAPDH?

- Normalize p21 expression
- "Housekeeping" gene



Q5:

Briefly describe “synthetic lethality” and how it applies to your cell viability experiment

# Briefly describe “synthetic lethality” and how it applies to your cell viability experiment

## What is synthetic lethality?

The combination of  
2 or more gene defects  
→ cell death.

Individual mutations do  
not kill the cell

## How does it apply to our experiment?

If we knock out NHEJ and HR  
we expect cells to die when we  
induce DNA damage. We do not  
expect cells to die from DNA damage  
if only a single pathway has been  
knocked out