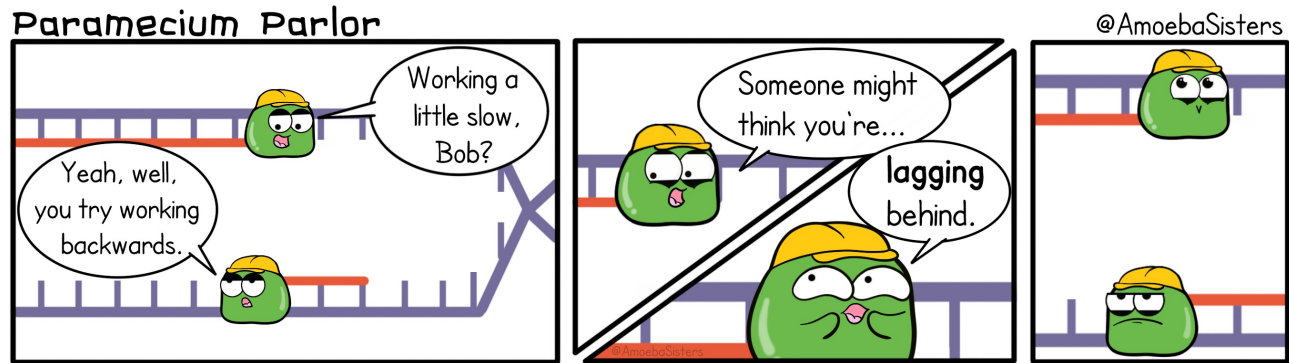


# Finish this (start PCR) before prelab

1. Make 100  $\mu$ M gRNA stock (e.g. 20 nmol  $\rightarrow$  200  $\mu$ L nuclease-free water)
2. Make primer mix (10  $\mu$ M of each primer in 20  $\mu$ L volume)
  - 2  $\mu$ L gRNA (forward primer)
  - 2  $\mu$ L RevP (reverse primer)
  - 16  $\mu$ L nuclease-free water
3. Take PCR tube and label with your team
  - 10.25  $\mu$ L nuclease-free water
  - 1.25  $\mu$ L primer mix (from #2)  $\rightarrow$  OK to use P20
  - 1  $\mu$ L pgRNA plasmid
  - 12.5  $\mu$ L Q5 Hot Start 2x Master Mix

# M2D3: Clone psgRNA expression plasmid

1. Set up reaction to generate gRNA\_target plasmid
2. Pre-lab discussion
3. Continue generation of psgRNA
4. Transform pgRNA\_target into bacteria



# Mod2 Overview

Research goal: Increase the yield of commercially valuable byproducts in *E.coli* using CRISPRi technology to target genes involved in mixed-acid fermentation pathway.

## Last Lab:

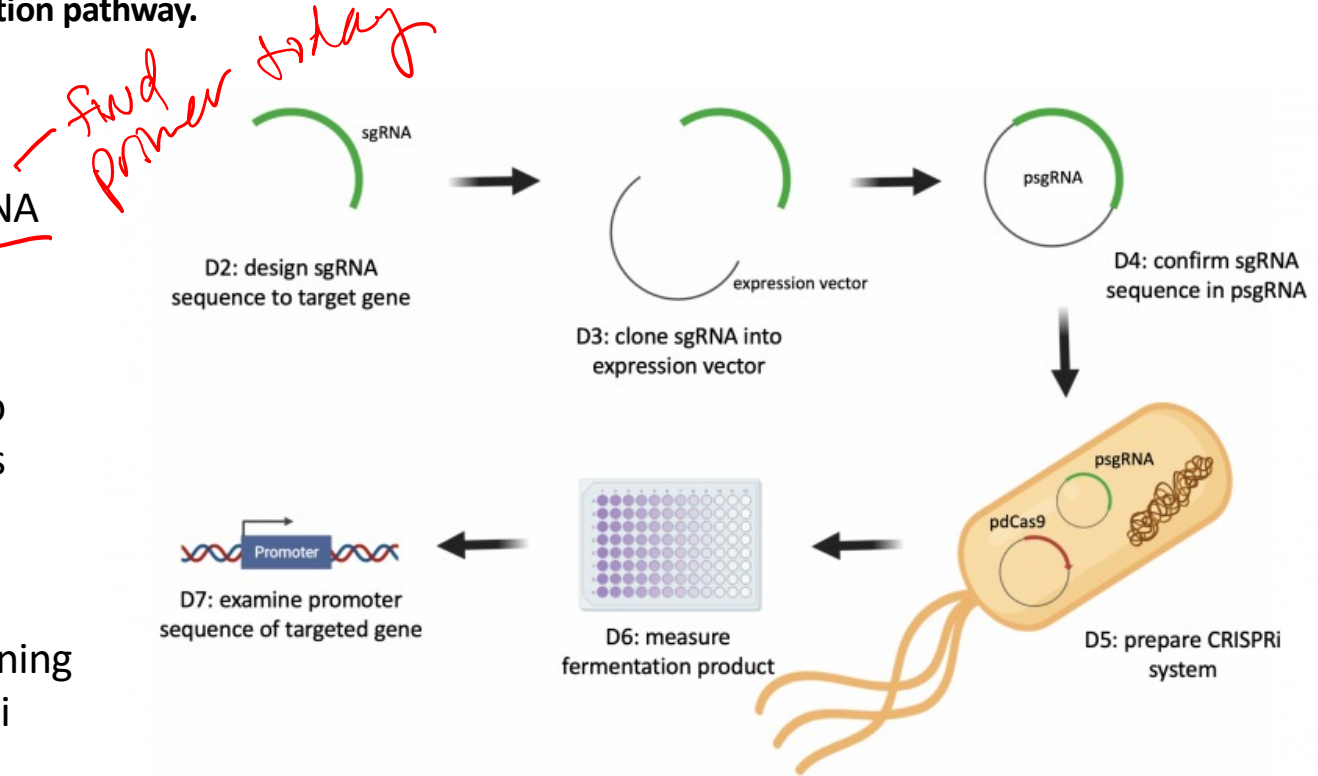
Chose metabolic gene of interest and designed sgRNA to target it

## This Lab:

Clone sgRNA into vector to create plasmid that targets gene of interest

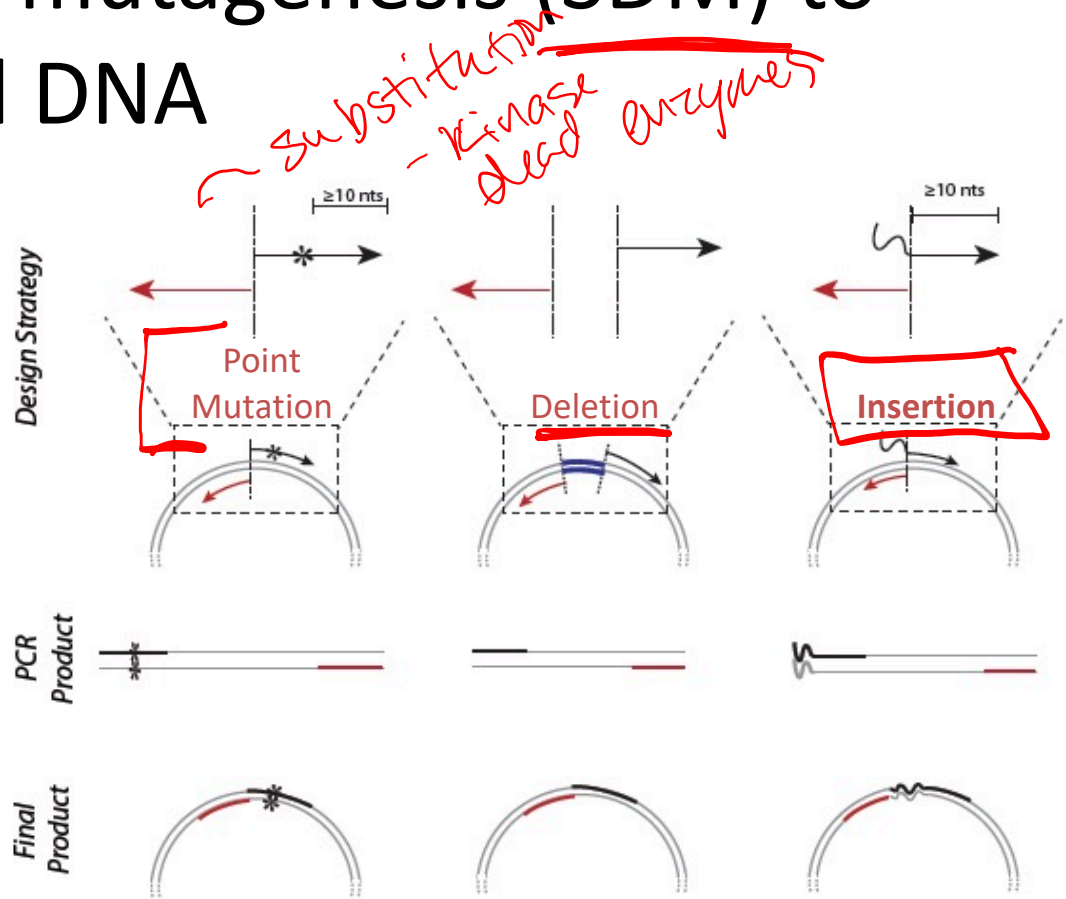
## Next Lab:

Confirm correct sgRNA cloning and do preliminary CRISPRi system preparations

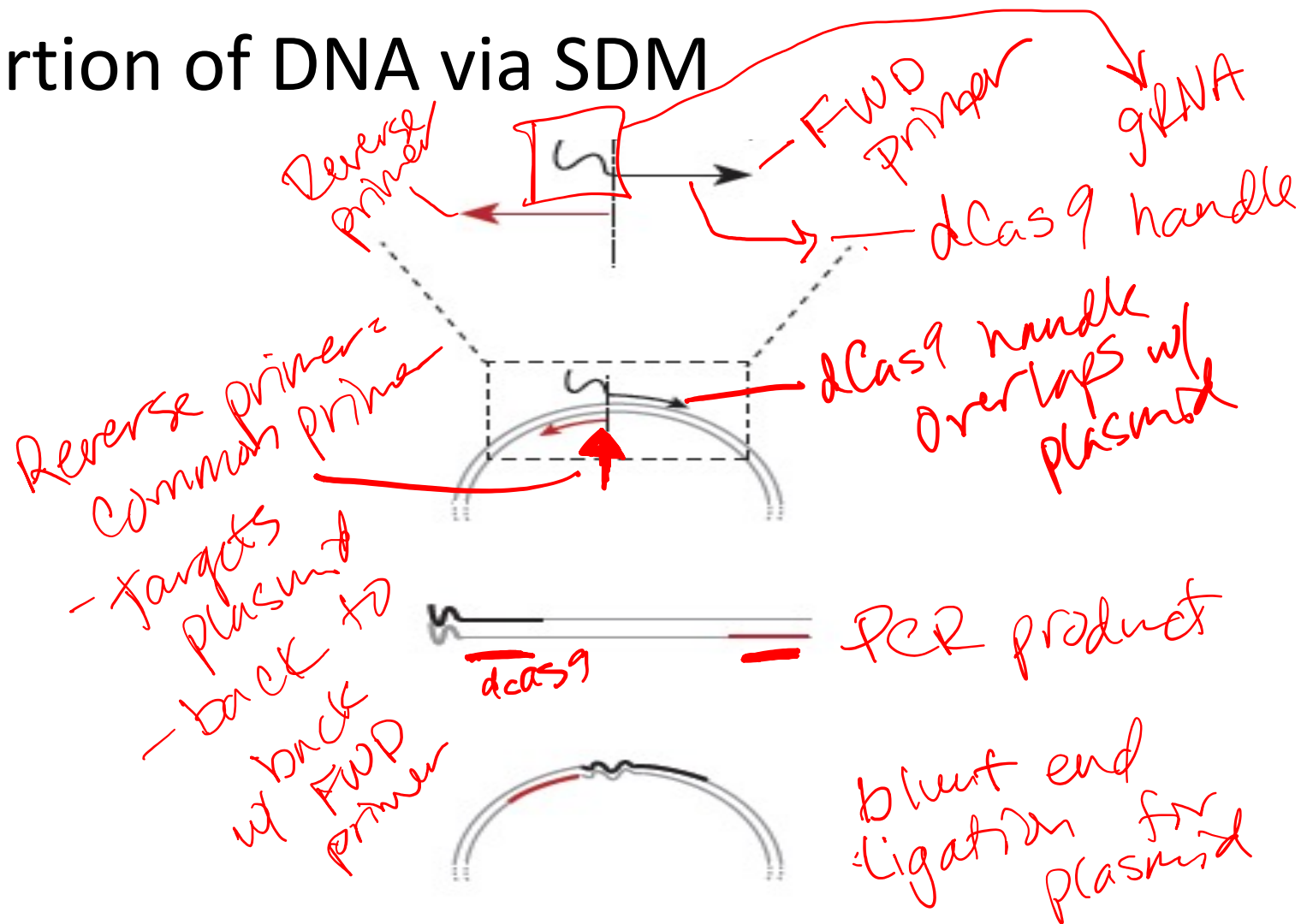


# Use site-directed mutagenesis (SDM) to engineer plasmid DNA

- NEB Q5 SDM kit  
*polymerase*
- Create specific, targeted changes in double-stranded plasmid DNA

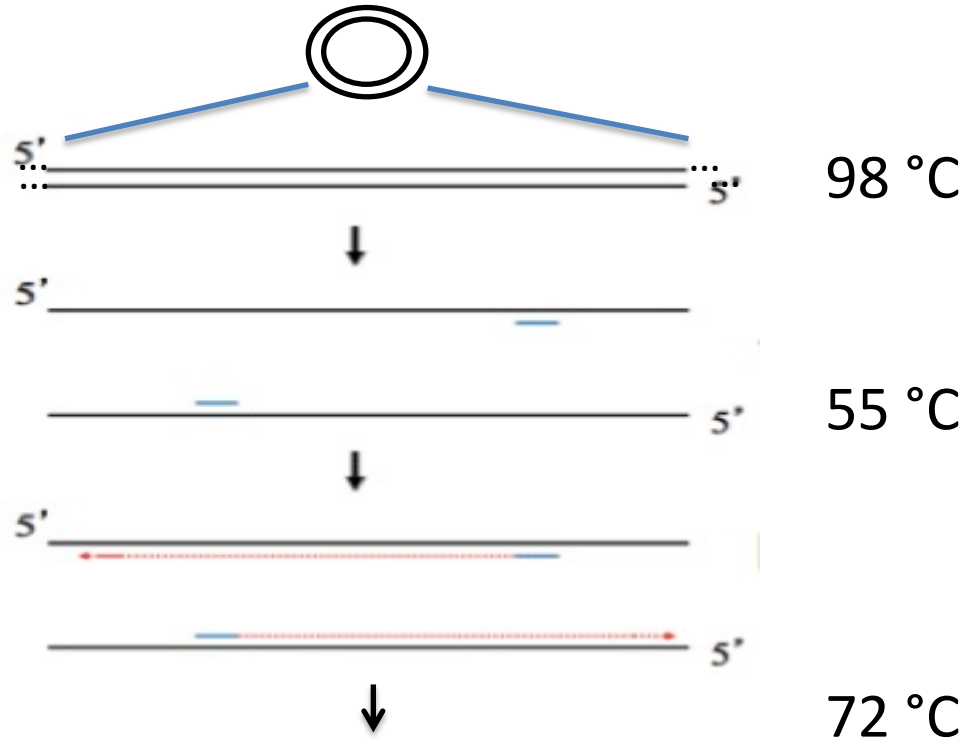


# Insertion of DNA via SDM



# SDM Part 1: PCR amplification of DNA

Ingredients	
F primer	
R primer	
template → pSGRNA	
dNTPs	
Q5 Taq	→ Polymerase
	High fidelity
Buffer (pH, cofactors like $Mg^{2+}$ )	
$H_2O$	



# SDM Part 2: Recover circular plasmid product using Kinase-Ligase-Dpn1 (KLD) enzyme mix

1. Exponential Amplification (PCR)

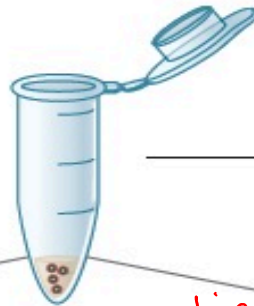
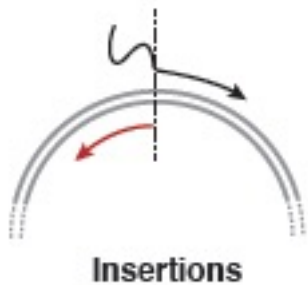
- Q5 Hot Start 2x Master Mix

2. Treatment & Enrichment: Kinase, Ligase and Dpn1

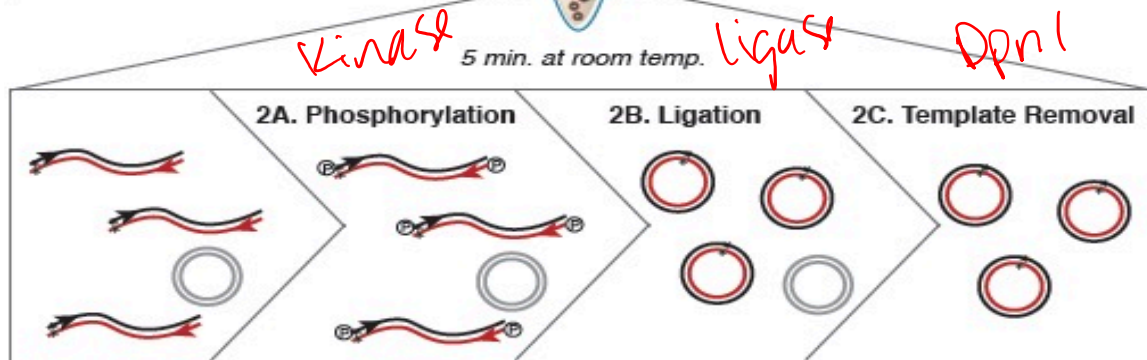
- 10x KLD enzyme mix

3. High-efficiency transformation

- NEB 5-alpha competent cells

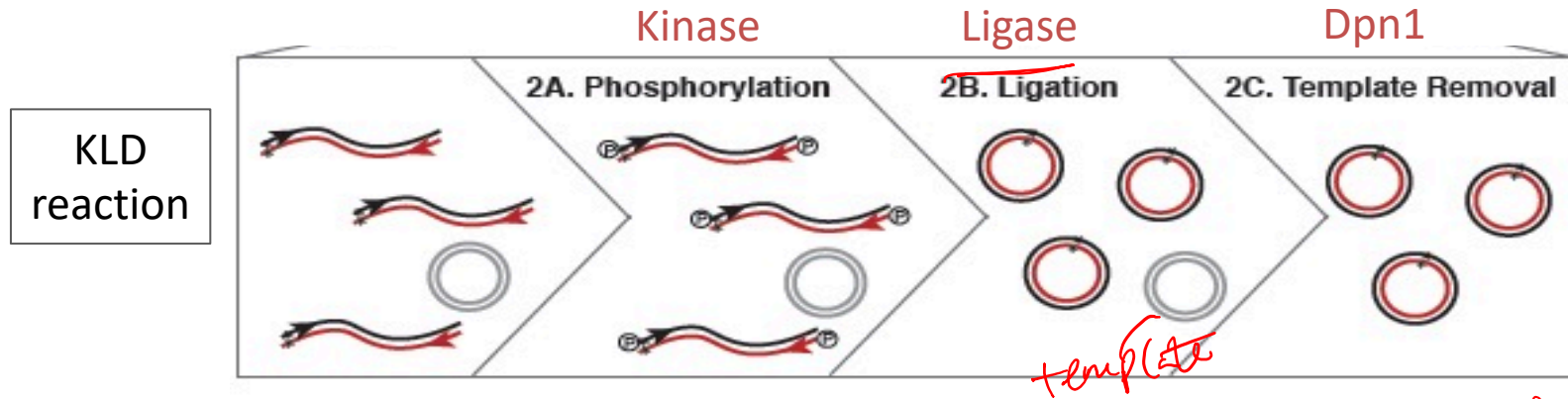


After PCR:  
1) linear product w/ insert  
2) template psgRNA - no insert



## Phosphorylation video:

<https://www.neb.com/tools-and-resources/video-library/the-mechanism-of-dna-phosphorylation>



**K:** phosphorylates PCR product ends - promotes ligation

**L:** ~~repair~~ repair plasmid backbone - now circular

**D:** removes methylated DNA (template)  
↳ cleave  
↳ produced in bacteria



# For today

1. Continue production of sgRNA plasmid
2. Work on journal club in down time

## For M2D4...

1. Create a journal club slide and script for 1 figure from your paper
  - Use the homework checklist on the wiki for guidance

# For homework, make a Journal Club presentation slide

Craft 1 slide using your JC article to present the data from a figure.

- Your slide should show the data and highlight the key finding(s).
- The information should be clear and large enough to read.
- Keep text to a minimum. (NO figure captions on slide!)

➔ The title should state the take-home message of the data that are shown.

no more  
than 1/2 of  
slide is text

# EXAMPLE SLIDE: Football coaches are the highest paid academic employees at doctoral-granting universities

- Data represent expression of Y using method A
- Possibly something about the control(s), if applicable
- Perhaps an important note about the data that is not already stated in the title
- Transition to next slide...

→ in text or verbally

