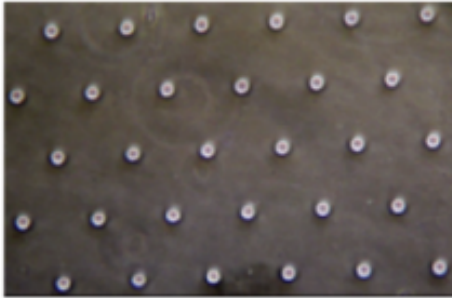


M1D5:

Complete biochemical experiment and apply chemical treatments for sub-nuclear foci assay

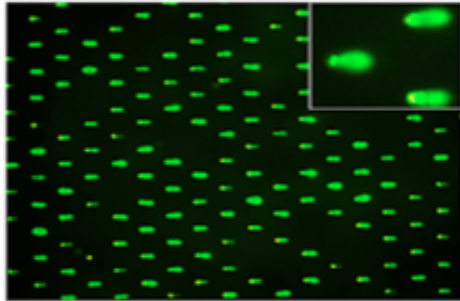
1. Communication lab workshop
2. M1Q1
3. Prelab discussion
4. Benchwork
 - Complete CometChip enzyme treatment
 - Electrophorese CometChips
 - Induce DNA damage for H2AX assay

Overview of Mod1 experiments



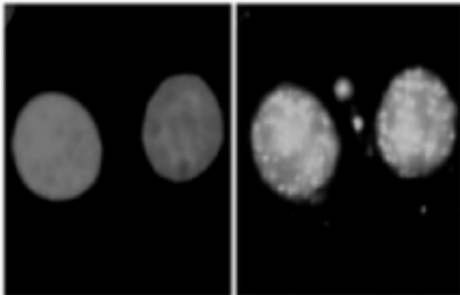
1. Optimize comet chip assay

- Test loading variables



2. Use comet chip assay to measure DNA repair

- Measure effects of MMS and H₂O₂ on BER

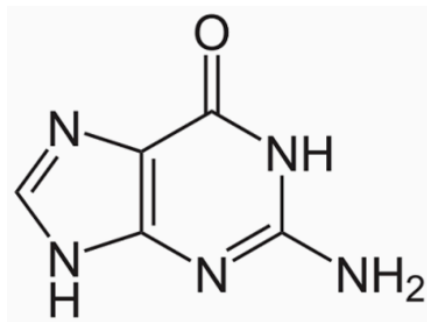


3. Use immuno-fluorescence assay to measure DNA repair

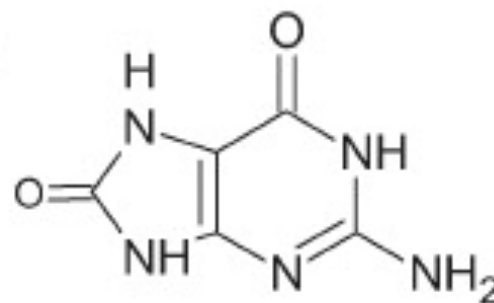
- Examine effect of MMS and H₂O₂ on DSB abundance

Damaging agents 'decorate' bases

Oxidative damage: H_2O_2

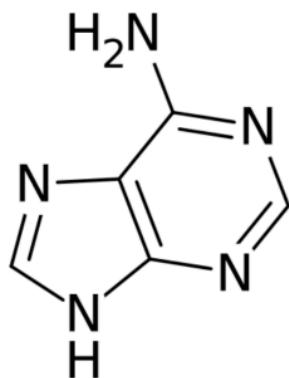


guanine

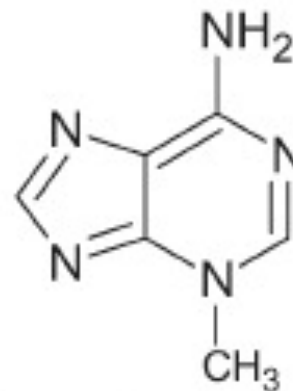


8oxoG

Alkylating damage: MMS



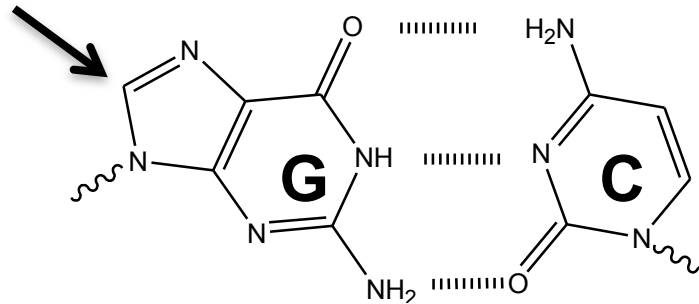
adenine



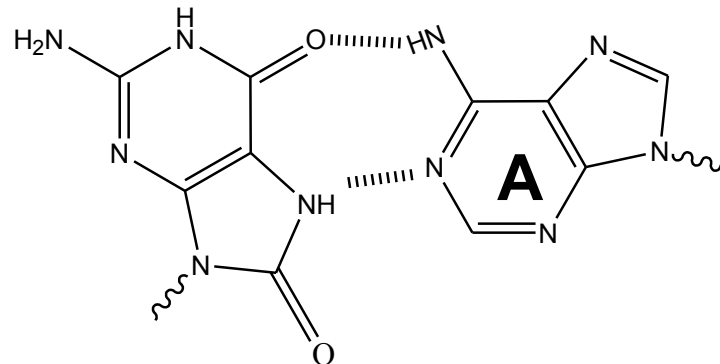
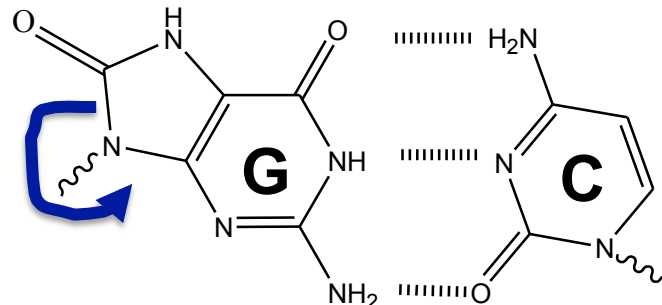
3meA

Oxidative damage induced by H_2O_2

•OH

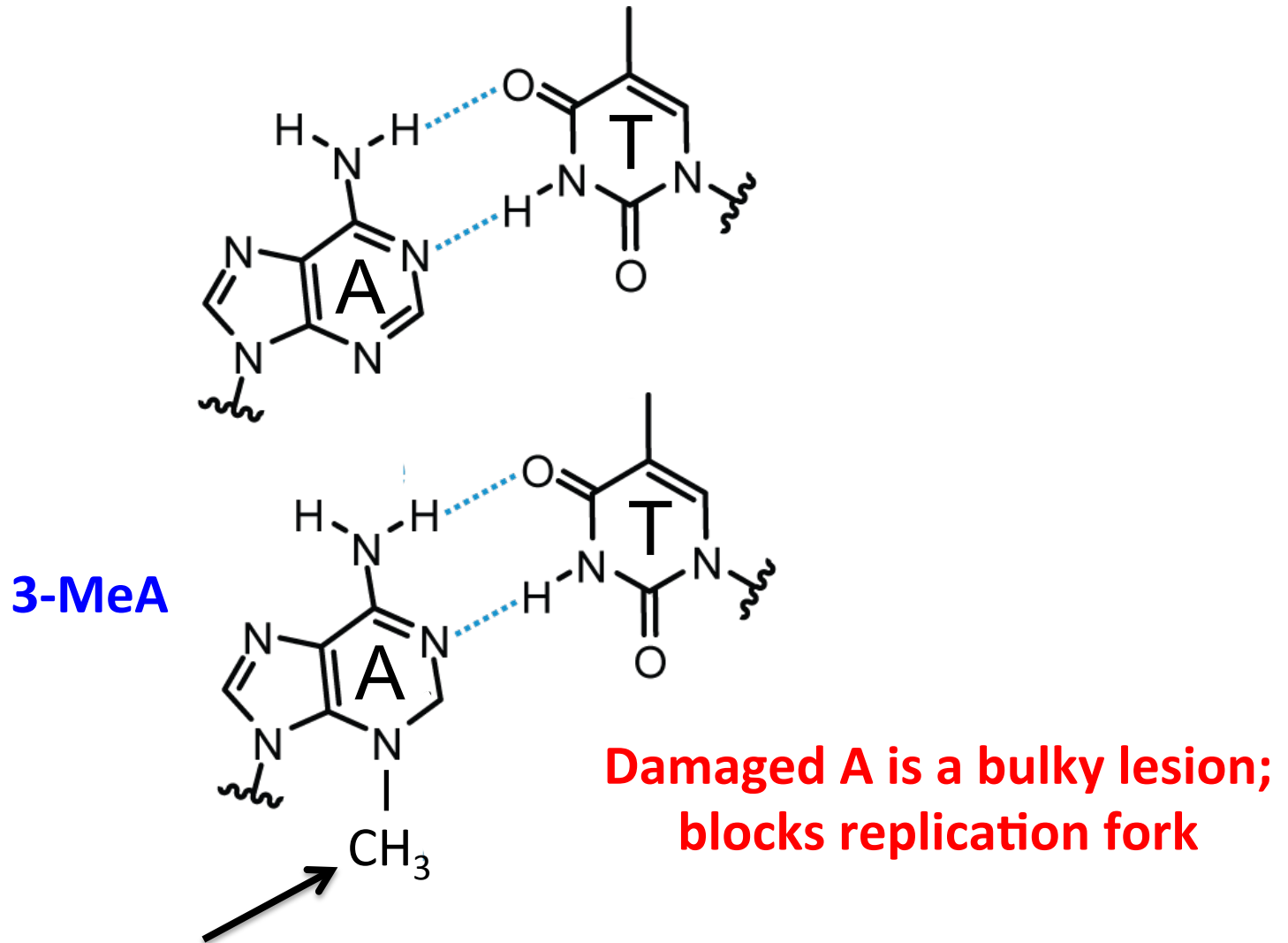


8oxoG



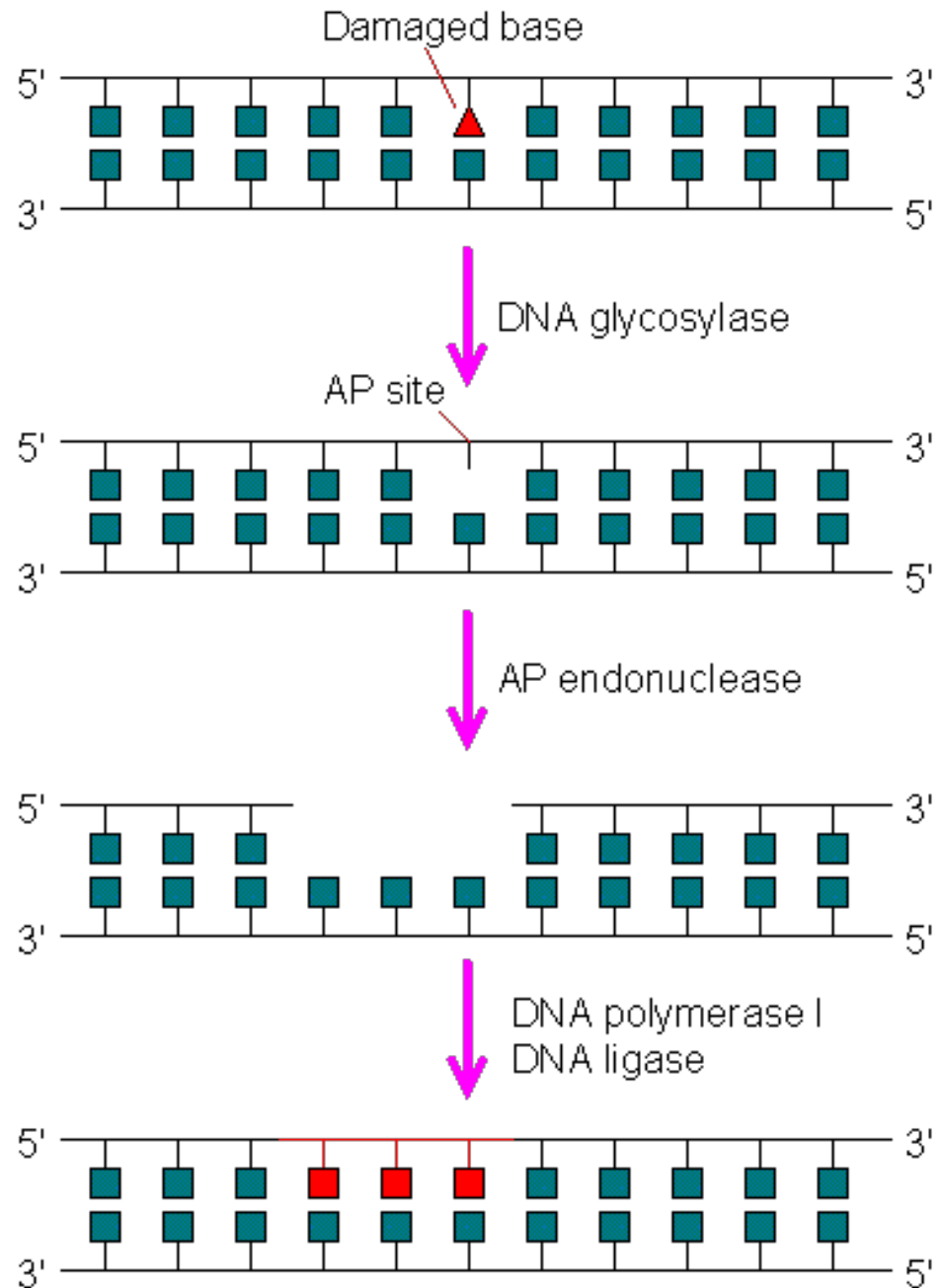
Damaged G read as T;
mutation GC→AT

Alkylating damage induced by MMS

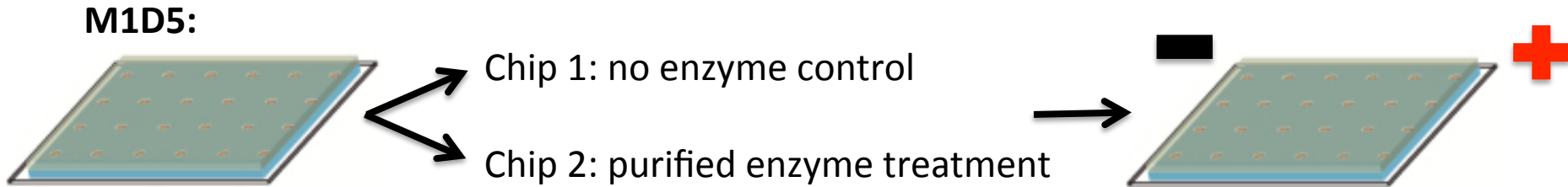


BER pathway corrects damaged bases

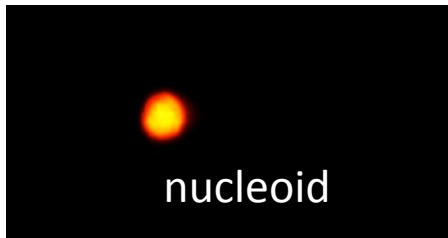
- Glycosylases are specific to type of damage
 - Oxoguanine DNA glycosylase (Ogg)
 - Alkyladenine DNA glycosylase (Aag)



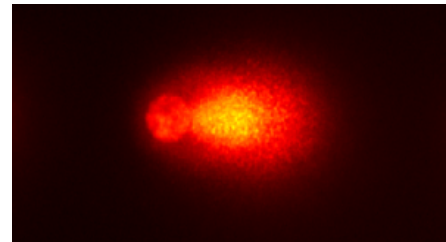
Exp2: Biochemical testing using CometChip



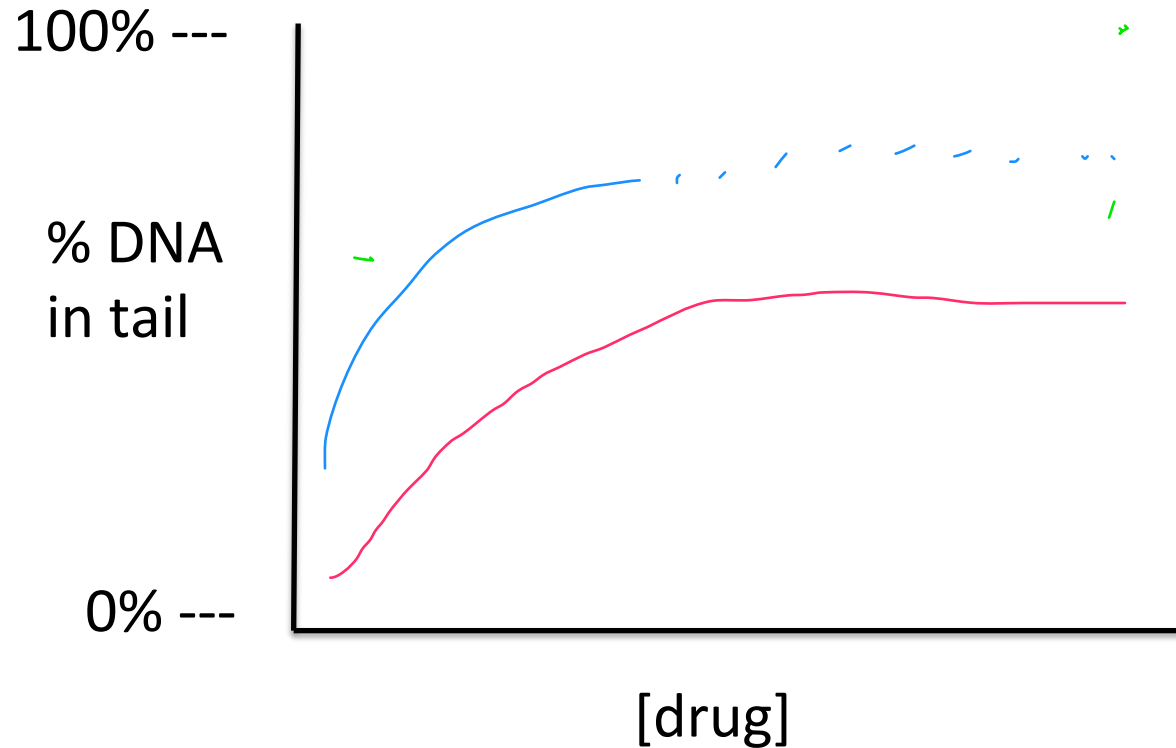
M1D6 and D7:



No damage:
supercoiled



What result(s) do we expect...



WT

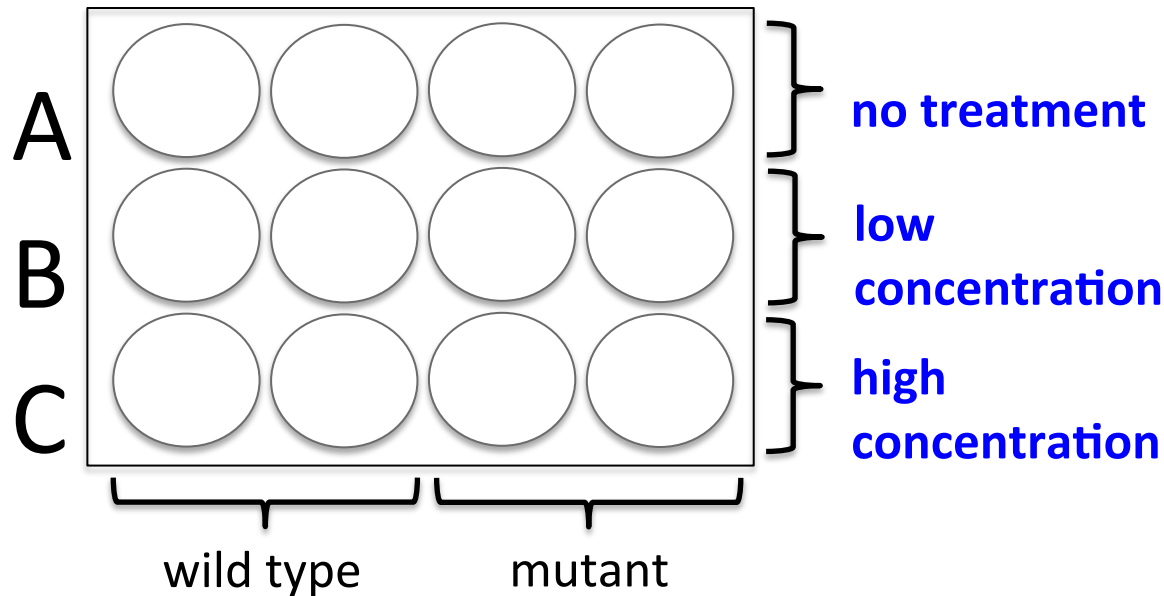
mutant

Keep track of the players!

	Oxidative damage	Alkylating damage
Chemical treatment:	H ₂ O ₂	MMS
Mutant cell line:	Ogg1-/-	Aag-/-
Enzyme:	Fpg (<i>E. coli</i> Ogg1)	hAAG (human Aag)

Exp3: DSB abundance using H2AX

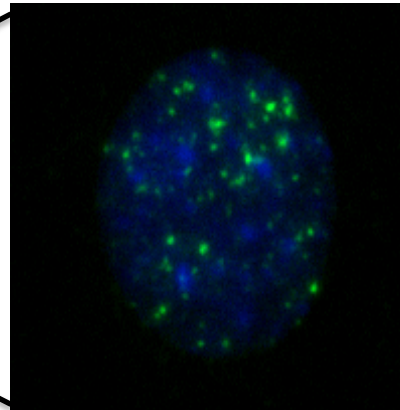
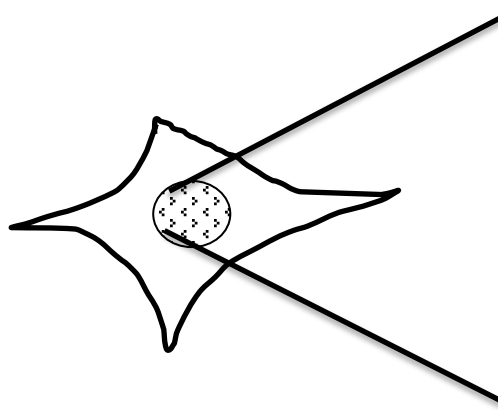
M1D5:



→
fix cells

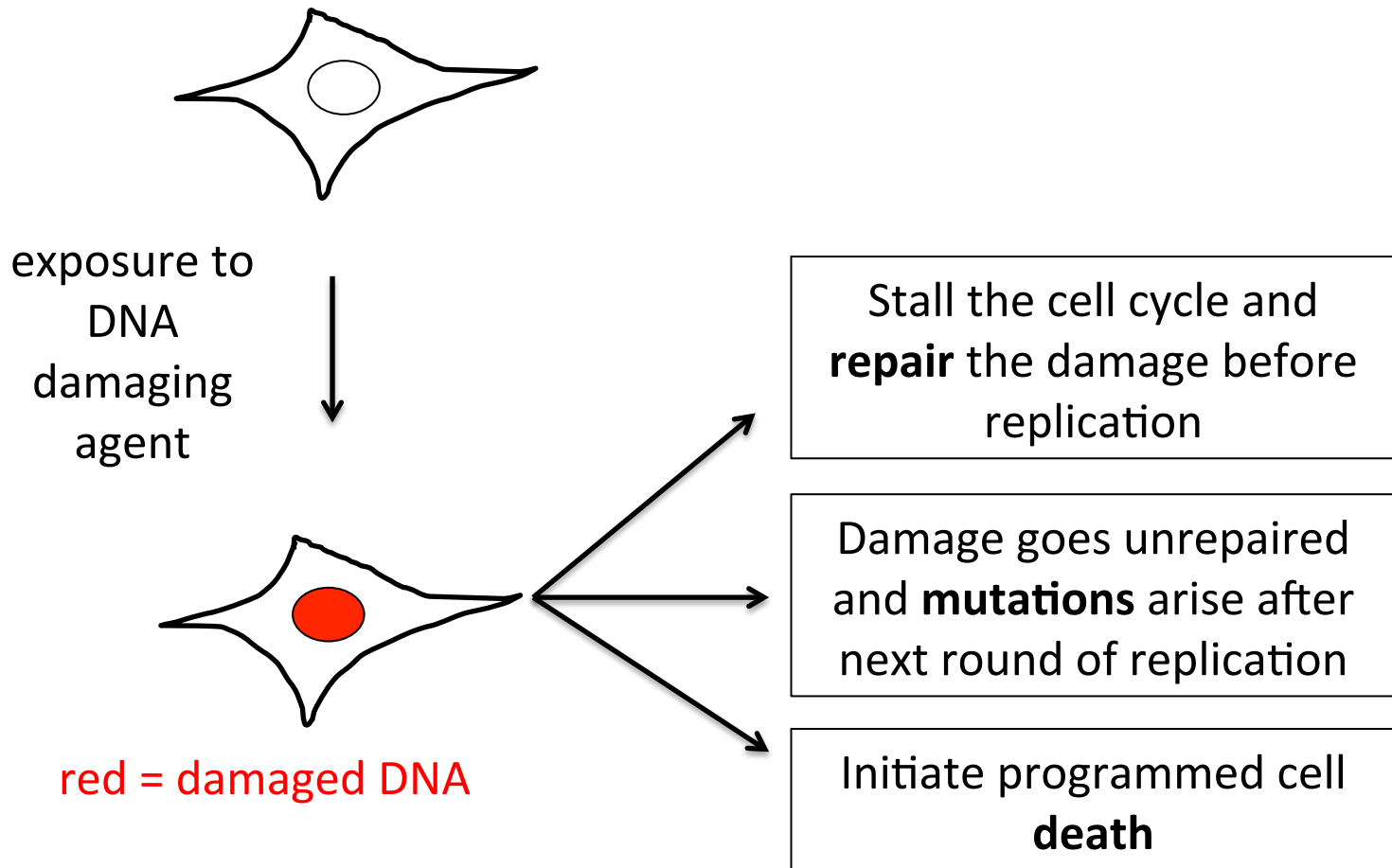
M1D6 and D7:

→
stain DNA



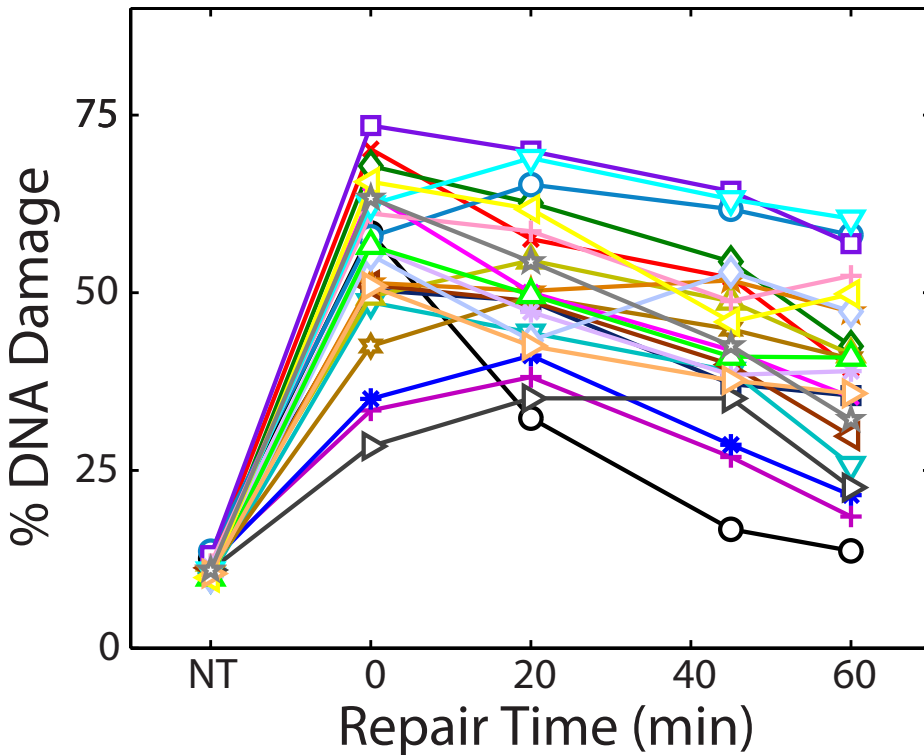
Blue: DNA
Green: γ H2AX foci (DSB)

How does a cell respond to DNA damage?

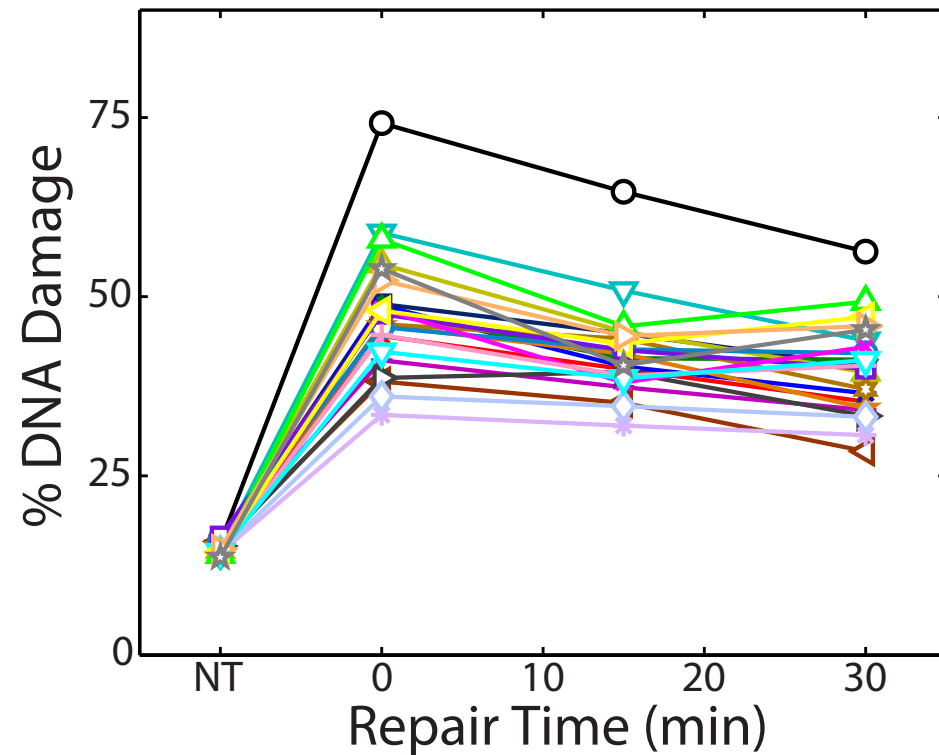


DNA repair capacity varies

Oxidative Damage



Alkylation Damage

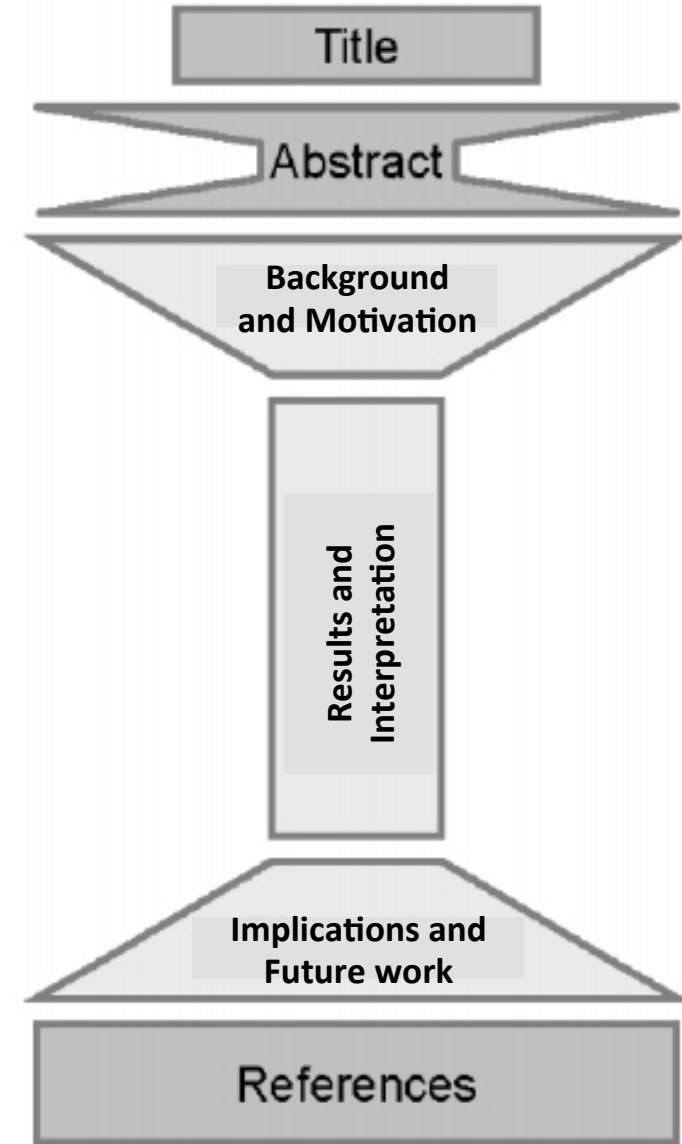


Options to consider: concentrations and recovery time

1. Which concentrations will you use to induce DNA damage?
 - H_2O_2 : 5, 10, 15, 20, 25 μM
 - MMS: 0.1, 0.2, 0.5, 6, and 8 mM
2. How long will you allow for repair of damaged DNA?
 - 30 min, 60 min, or 24 hr

Data summary due Wed., Oct. 11 at 10p!

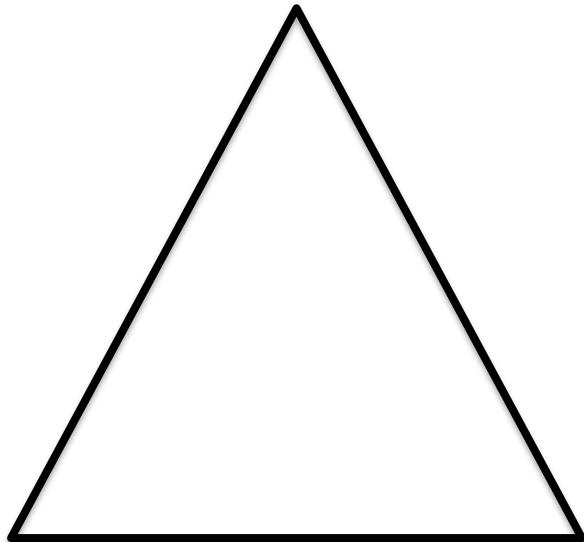
- Remember to ‘funnel’ the information
- Be sure the pieces ‘match’ throughout
- ONLY the abstract is written as a paragraph, ALL OTHER sections should be in bullets



Writing future works & implications

Implications and Future Work: potential topics [\[edit\]](#)

- **Topic:** Did your results match your expectations?
 - If no, provide a putative explanation. If yes, how can you further test if your hypothesis is correct?
- **Topic:** Based on the results, whether they matched your expectations or not, what experiments might you recommend next?
 - Follow-up experiments could distinguish between competing explanations of a given outcome or broaden the sample set for a question you already asked, to give just two examples.
- **Topic:** How might this assay be improved?
- **Topic:** How might this assay be used as a research tool? in the clinic? in industry?



Starts with summary of your findings – DO NOT just repeat all of the data



Finishes with tie back to the impact statement (big picture topic)

Other notes on homework due M1D6

- Draft mini-presentation outline
 - Abide by content and time guidelines (see wiki!)
 - Introduce yourself and your research
 - State hypothesis clearly
 - Be quantitative (use placeholders in this assignment)
- Read journal article and prepare for discussion

Cytometry

PART A
Journal of the
International Society for
Advancement of Cytometry

Standard Fluorescent Imaging of Live Cells is Highly Genotoxic

Jing Ge,¹ David K. Wood,² David M. Weingeist,¹ Somsak Prasongtanakij,³ Panida Navasumrit,³ Mathuros Ruchirawat,³ Bevin P. Engelward^{1*}

Today in lab...

- Complete enzyme treatments with CometChips in main laboratory
- Treat cells with DNA damaging agent for H2AX assay in tissue culture
 - Depending on timing, pairs may have to split between rooms
 - If time permits, fix cells
- Electrophorese CometChips