# M2D1: Examine SMM data collected using TDP43 protein

- 1. Prelab
- 2. Walk through SMM analysis
- 3. Examine chemical structure of hits
- 4. Discuss journal article

#### **Office Hours:**

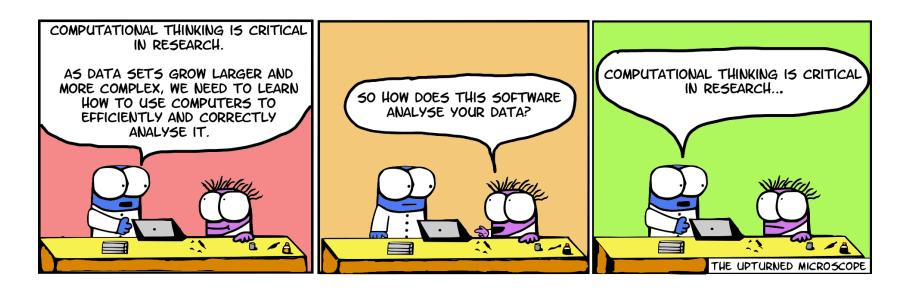
Monday: 1-2pm @ 16-319 and Zoom Becky

3-5pm @ 16-317 and Zoom Noreen

Tuesday: 10-11am @ 1-390 Becky & Jamie
Thursday:10-11am @ 1-390 Noreen & Jamie

\*After lecture by request

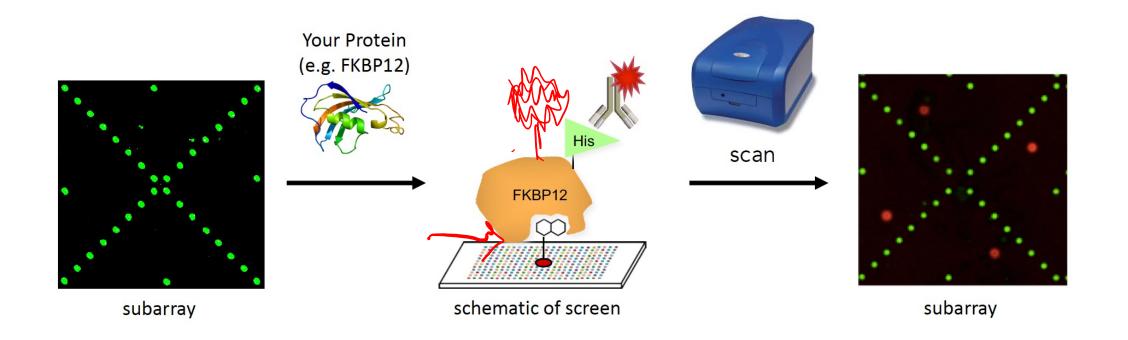
\*Also available by appointment



## SMM workflow

#### **SMM Screen**

#### Data Acquisition



## How do we use the SMM to screen for ligands that bind our protein of interest?

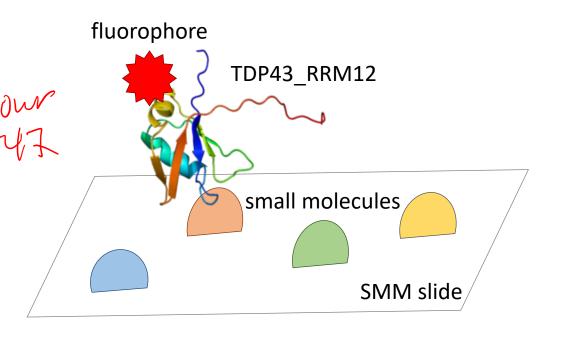
 Create a recombinant protein of the TDP43 RNA binding domains (TDP43\_RM12)

 Label this protein with a Alexa647 fluorophore

 Incubate the SMM slide with our purified and labeled TDP-43 RRM12

Wash away unbound protein

Store for scanning



### Workflow for SMM data analysis

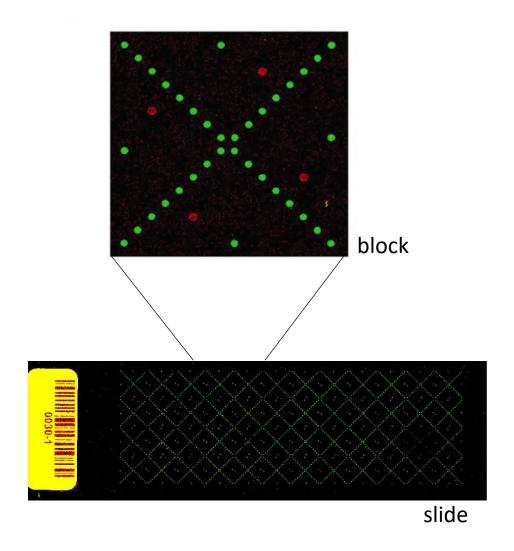
1. Align spots using fluorescence on 532 nm channel (sentinel spots)

2. Quantify fluorescence on 635 nm channel

3. Identify 'hits' with improbably high fluorescence

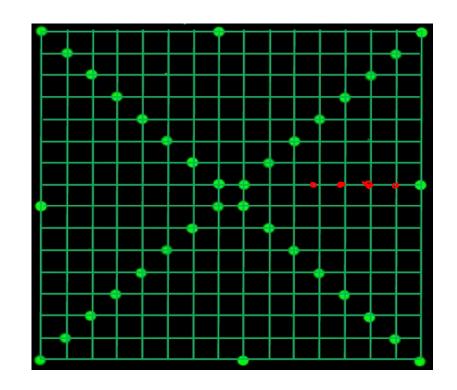
1

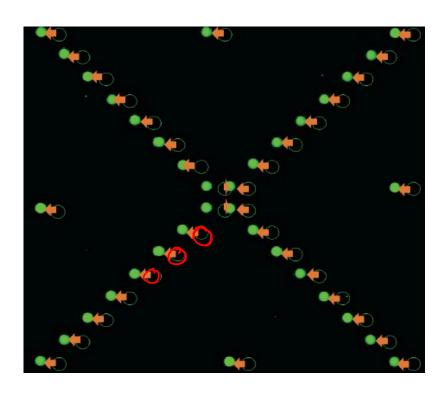
4. Complete 'by eye' analysis of putative hits to manually remove false positives

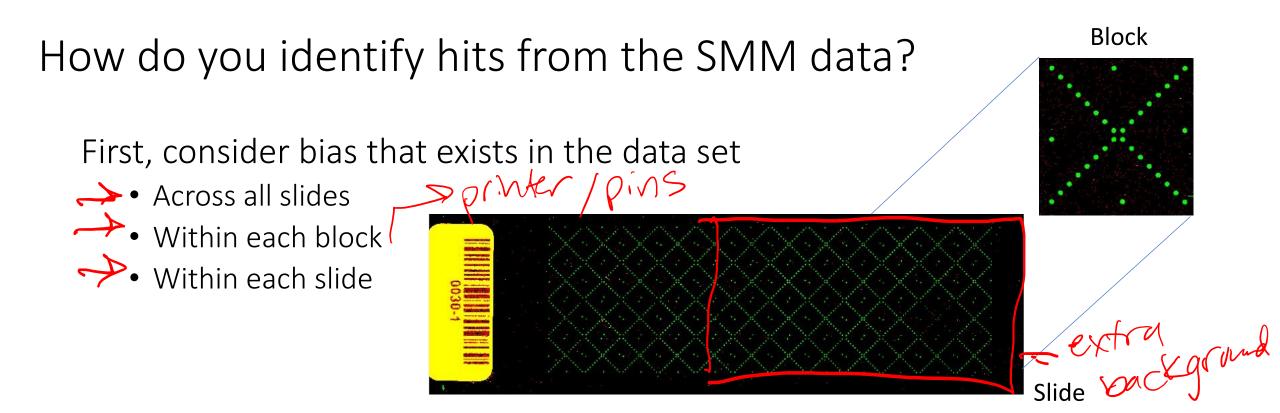


#### Align SMM using sentinel spots

- Slides are printed in block patterns (16 rows x 16 columns)
- Each ligand spot is identifiable via intersecting lines between sentinels







Then, identify hits with significantly higher fluorescence over background

Lastly, manually confirm hits to eliminate false positives

#### Spots are represented by an array of numerical values

 Each pixel is represented by a number that indicates intensity of the signal

 Computational analysis used to define 'hits'

```
8 173 625 816 823 856 815 831 568 136
       8 273 839 814 835 873 890 836 857 818 771 801
  7 175 780 805 877 941 936 920 973 921 842 819 714 125
  29 568 868 867 905 909 936 994 954 931 963 875 813
5 131 754 85 2 906 958 920 963 923 917 904 951 930 851 7 6
5 229 796 879 924 934 923 962 961 993 993 945 989 867 780 162
7 254 827 879 965 949 960 982 926 918 955 927 984 872 765 204
5 175 808 888 996 951 998 935 976 971 940 922 961 872 804 132
 57 666 859 968 999 947 977 985 916 928 960 974 841 678
 11 406 839 897 915 930 946 993 914 911 977 900 880 859
     60 624 830 890 973 903 921 912 930 881 850 623 54
       7 92 602 873 856 882 913 887 885 842 589 82
```

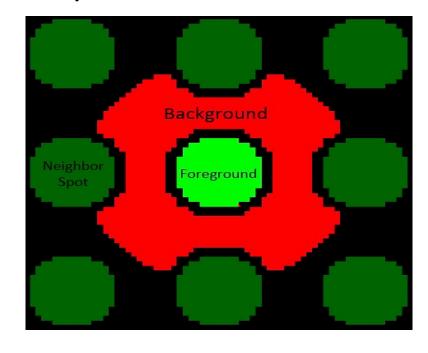
#### Fluorescence is quantified to identify hits

• Foreground:

SM printed

• Background:

noise



Signal-to-noise ratio (SNR) = 
$$\mu_{\text{foreground}}$$
 -  $\mu_{\text{background}}$   $\sigma_{\text{background}}$ 

#### SNR is then used to calculate the **robust z score**

- How different is the foreground signal from the background?
- Able to plot the distribution of the z scores to give an overview of whole data set

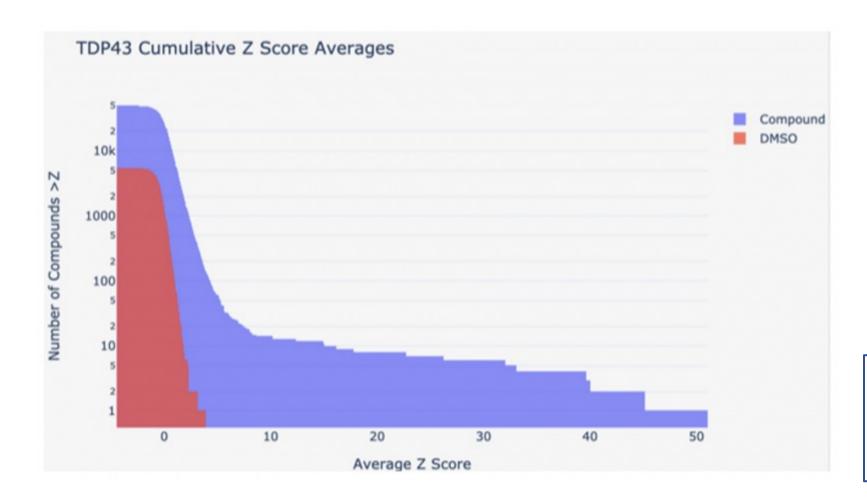
#### Average Z-score calculated for all compounds



Replicate spots averaged

Each count = unique compound

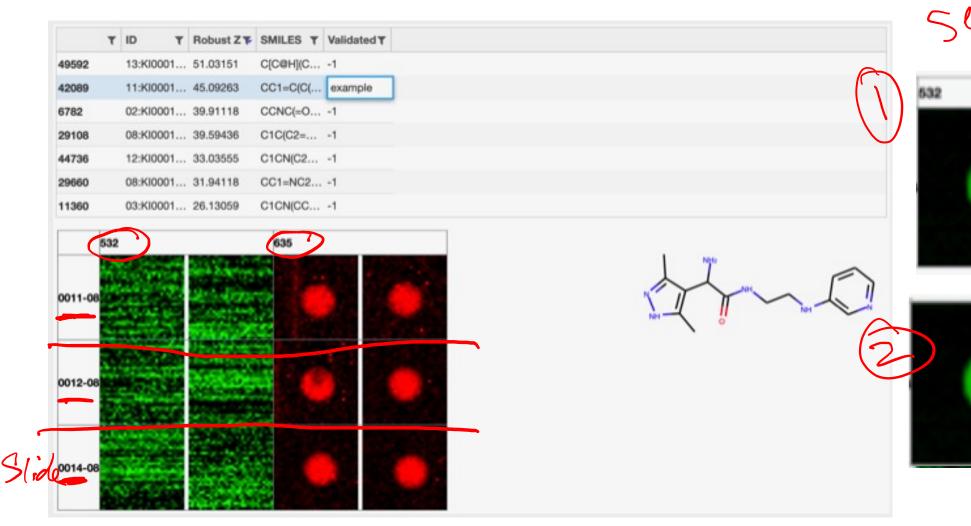
#### How many compounds have a particular z score?

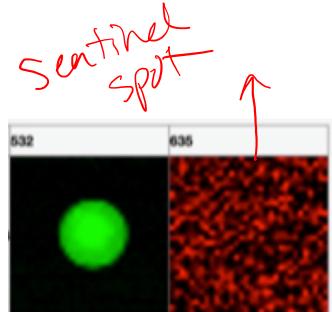


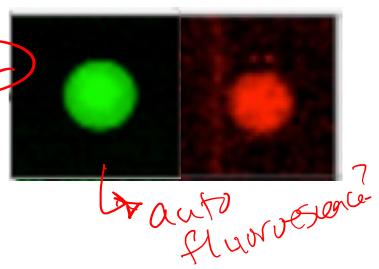
Height at x (average z score)
= number of compounds (y)
with that z-score or higher

• Useful for setting a threshold to exclude likely non-binders

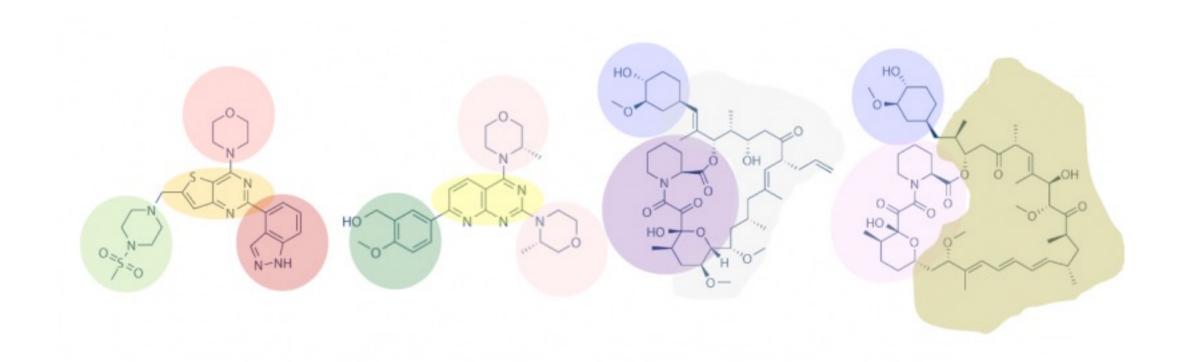
How do you validate hits manually?





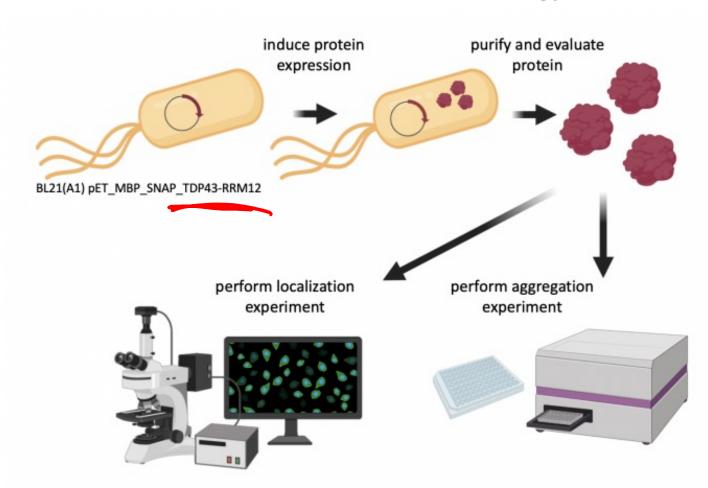


## How will you identify common structures?



#### Overview of Mod1 experiments

Research goal: Use functional assays to characterize ligands identified as binders to TDP43 from SMM technology



## For Today

- Work through SMM analysis procedure
- Evaluate chemical structures of identified hits
- Discuss reading of scientific papers with Noreen
  - Group 1: Teal, White, Gray, Purple, Pink
  - Group 2: Blue, Green, Yellow, Orange, Red

#### For M1D3

- Begin thinking about Background and Motivation for Data Summary
  - Submit document answering questions on the Homework section of wiki
  - Due Thursday, Feb. 10 at 1:05pm on Stellar
- Visit Comm Lab by M1D5
  - Can visit to discuss an assignment from any class, a personal statement for an internship application, etc...