

M1D3: Agarose gel electrophoresis

9/23/15

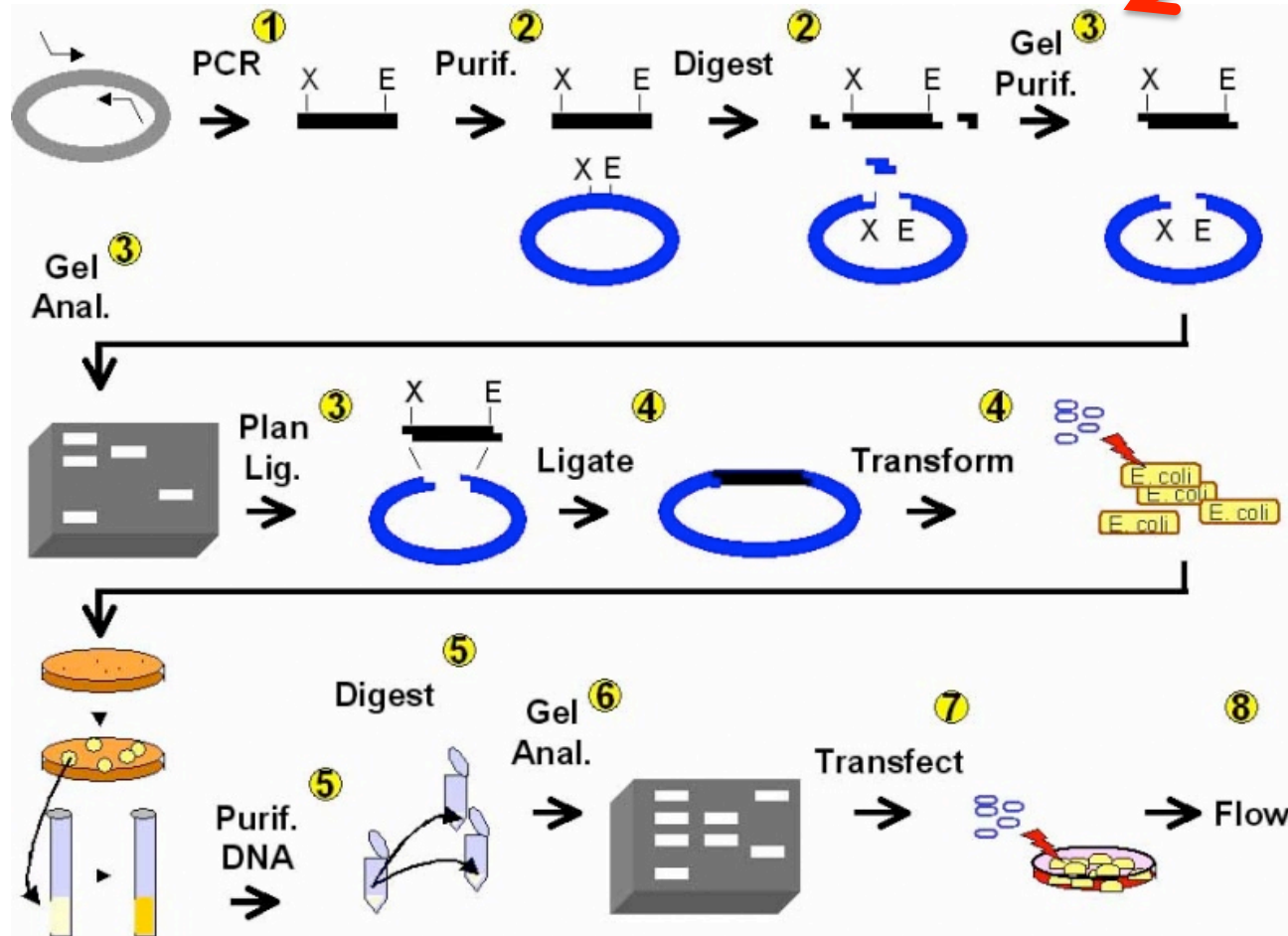
1. Attendance policy addendum

- Lecture absences
- Laboratory absences

2. Next time

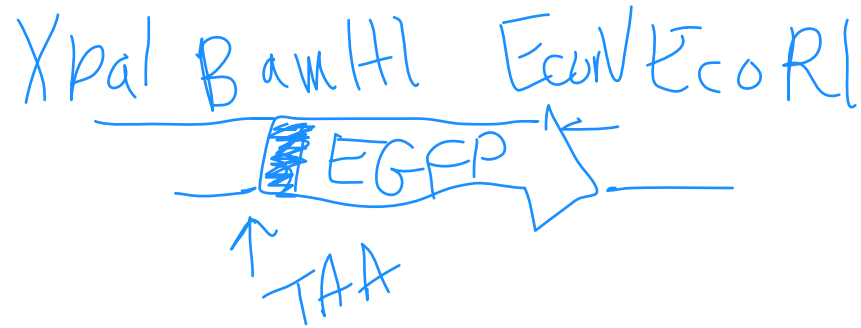
- Lab treat (quiz)
- Homework due M1D4

Mod 1 overview



Complete Part 1 of the protocol, then come together for prelab discussion

From last time...



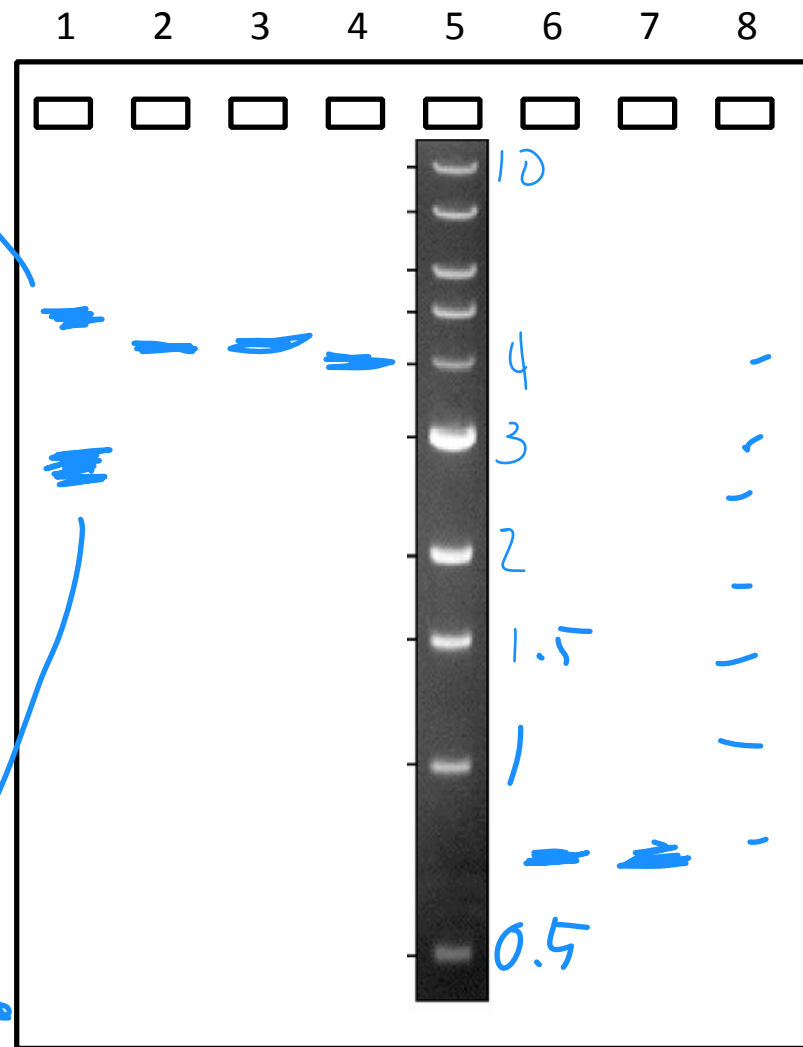
D1: amplify
D2: digest
D3: purify +
verify

How do you know the enzymes cut your DNA?

Evaluating RE digest

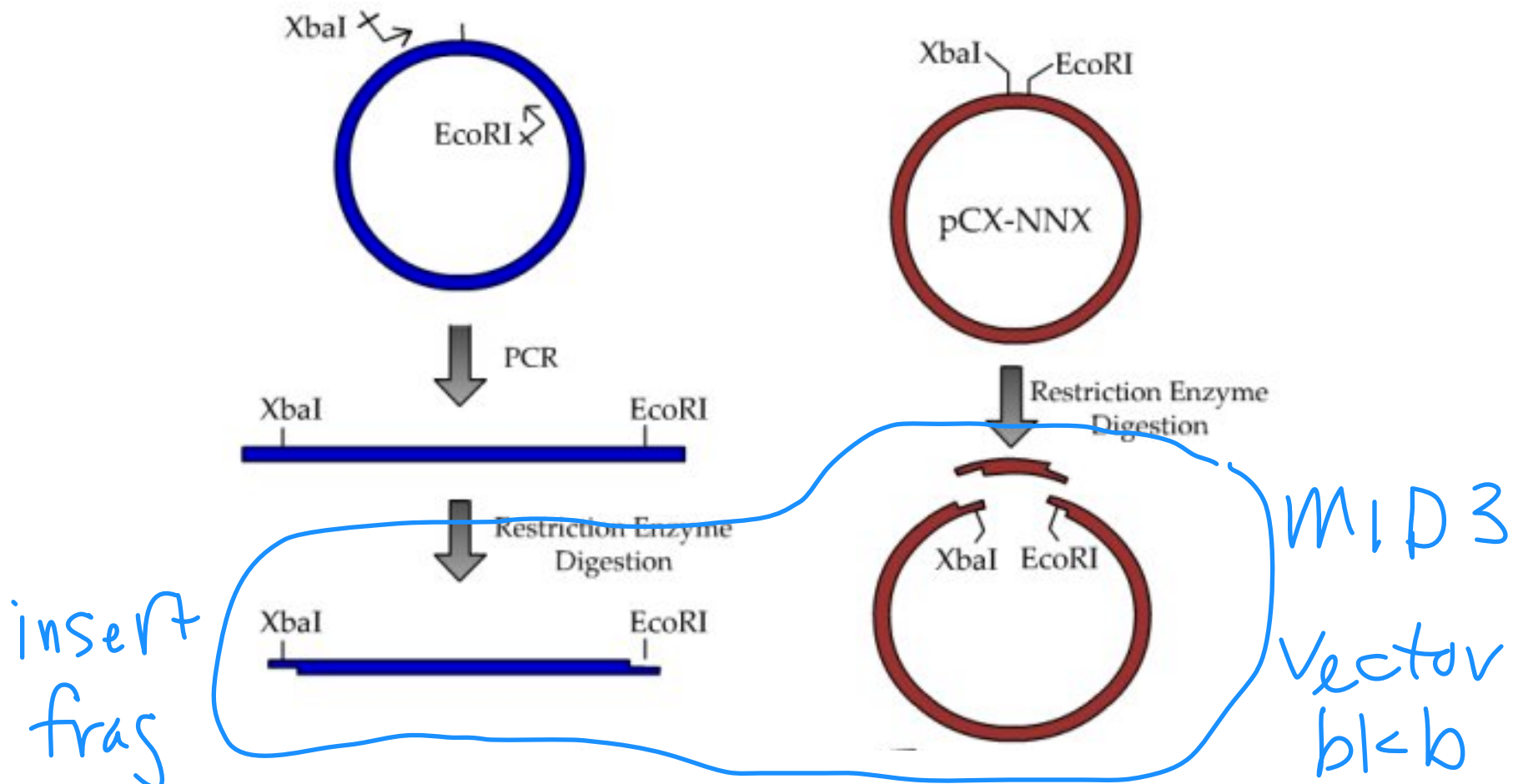
nicked (circular)

Lane	Sample	Volume to load
1 [^]	Uncut pCX-NNX [^]	10 μ L [^]
2	pCX-NNX <i>Xba</i> I	5 μ L
3	pCX-NNX <i>Eco</i> RI	5 μ L
4	pCX-NNX <i>Xba</i> I + <i>Eco</i> RI	25 μ L
5	1kb DNA ladder 🗝	20 μ L
6	PCR product <i>Xba</i> I + <i>Eco</i> RI	25 μ L
7	PCR product uncut	25 μ L
8	PCR no-template-control	25 μ L



Supercoiled

Engineering $\Delta 5$ EGFP construct



How do we recombine the DNA fragments?

LIGASE

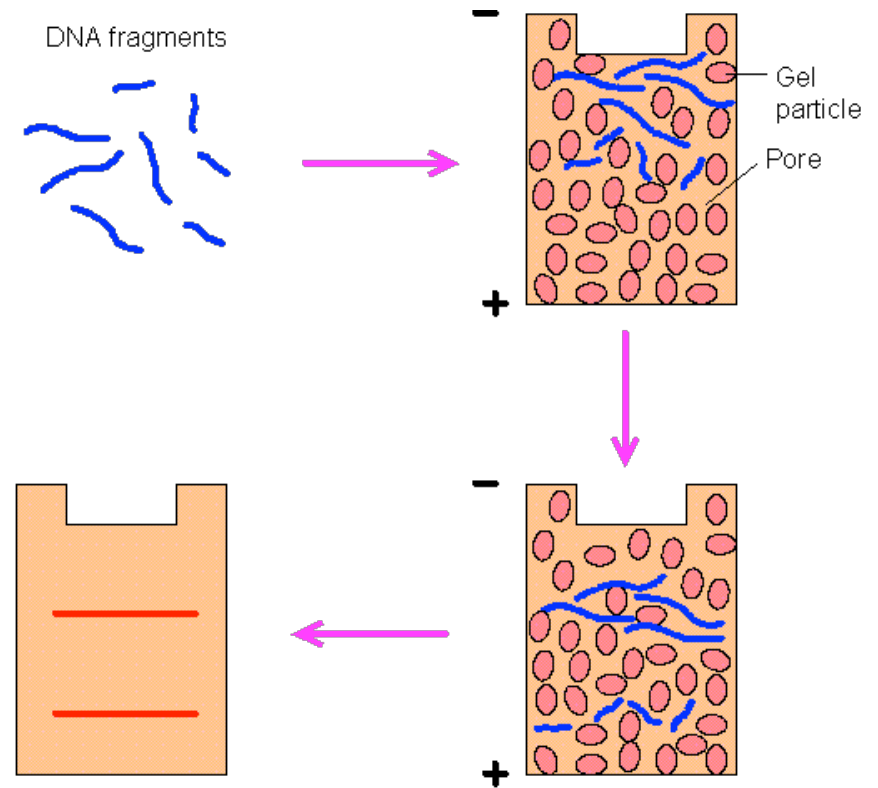
Prep for ligation: visualize DNA

- Gel electrophoresis

Separated by size + charge

driving force? charge
DNA charge? (-)

Separate? size



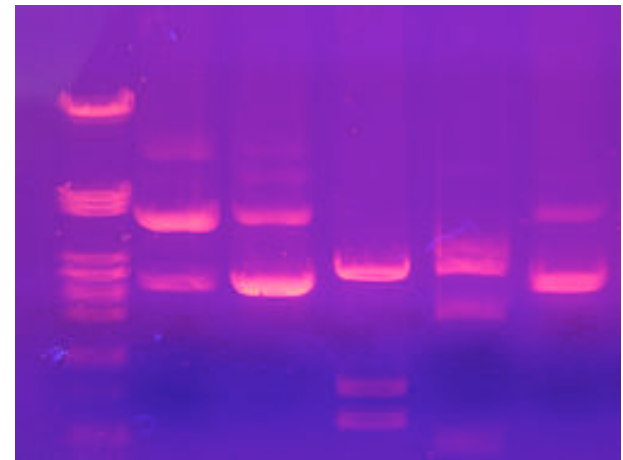
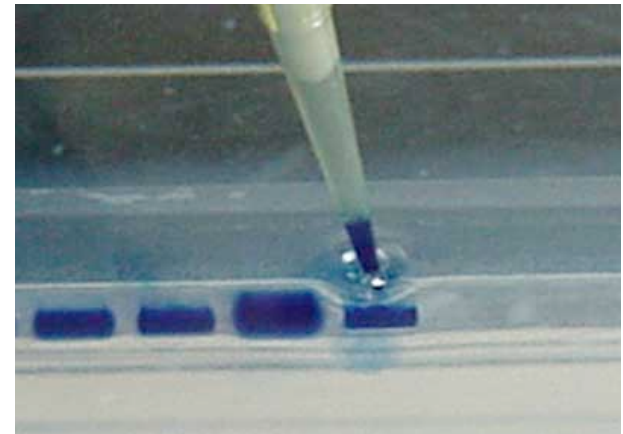
How do we visualize DNA?

- Loading dye

bromophenol blue - ~500bps
glycerol viscosity

- DNA stain

sybr-safe
DNA intercalator
visualize w UV



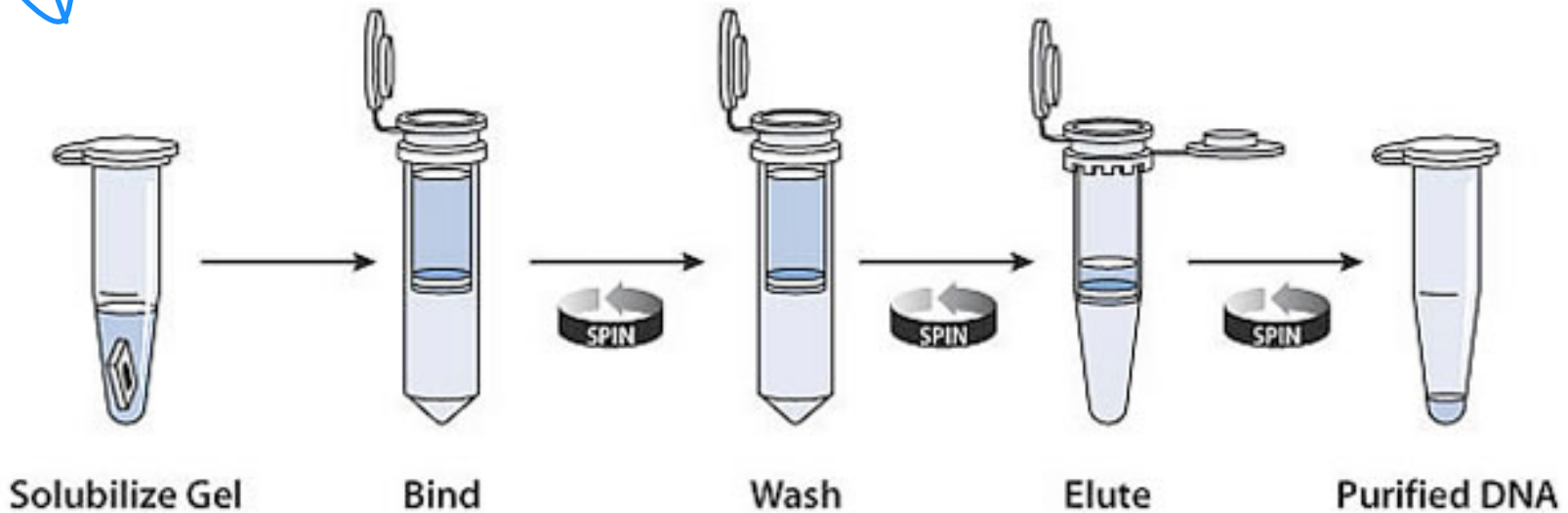
Safety note: wear nitrile gloves and face shield!!

Qiagen

Prep for ligation: purify DNA

- Gel purification

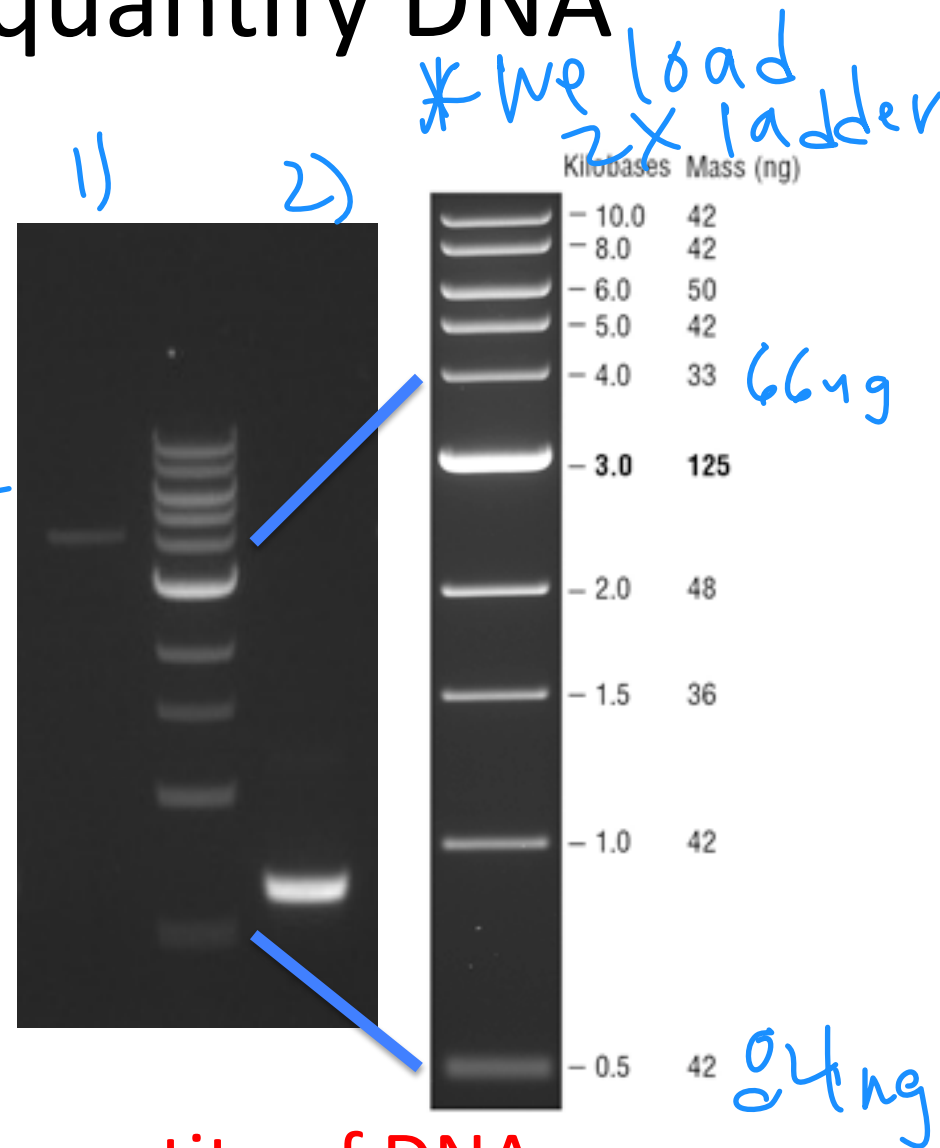
QG: ↑ salt ↓ pH
Wash: EtOH ↑ salt
Elution: ↑ pH (2) ↓ salt



Prep for ligation: quantify DNA

- ① • Amount (ng) of vector
band 1/2 intense
~ 33 ng / X volume
of DNA

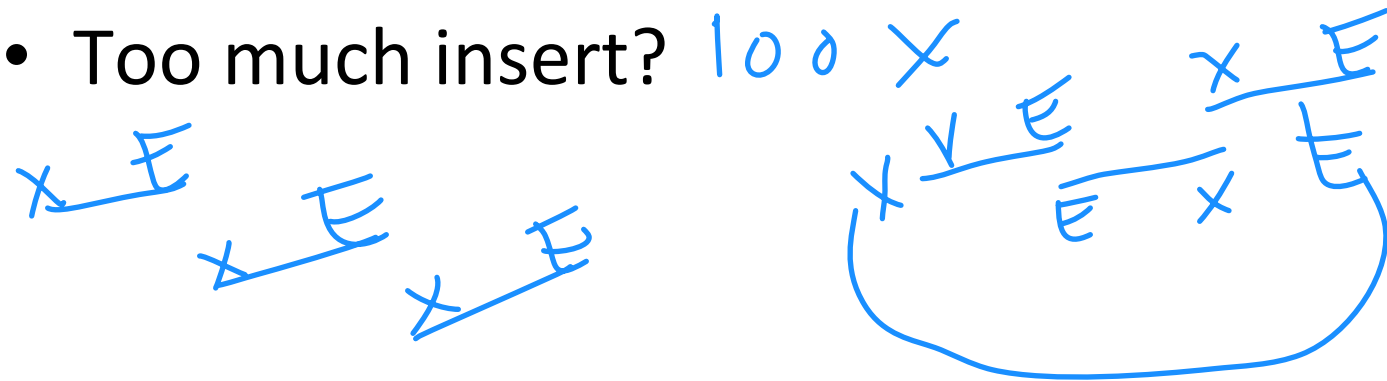
- ② • Amount (ng) of insert
~ 336 / X volume
of DNA



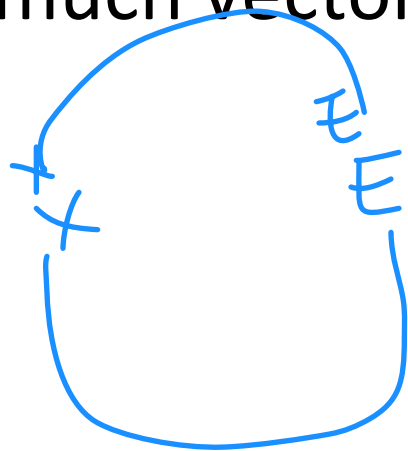
Mass of DNA \neq molar quantity of DNA

Optimal vector-to-insert ratio

- Ideally want 1:4 backbone: frag
– Molar ratio, **not** volume or mass



- Too much vector? 100x



giant vector

linear DNA = no colonies

Prep for ligation: ratio calculation

1. Examine recovery gel
 - will be posted by teaching faculty tonight
2. Estimate mass of vector and insert
3. Determine volume of vector needed (50-100ng)
4. Calculate mol of vector
5. Determine mol of insert needed (2X-4X vector)
6. Calculate volume of insert needed

Today

- Gel electrophoresis (gloves)
- Excise vector and insert from agarose and purify (gloves and eye protection)
- Prepare vector and insert samples for gel electrophoresis
- Pour gel to examine recovery (gloves)
- Networking event
 - September 24 at 5:30p in first floor Koch lobby

