

Start to think about these questions

Why should we care about scientific communication? Rewards, scenarios?

What makes you feel that communication has been successful?

As a receiver?

As a sender?

What makes you feel that communication has been successful?

As a receiver?

Clear message, logic flows, you can find your way around, actionable, visual appeal

As a sender?

Reward (citation, grade, funding)  
Feedback: questions or criticism

When we struggle to understand talks or papers, we often blame ourselves.

“I got stuck.”

“huge logical leap”

“way too much going on”

“what am I supposed to look at?”

but poor communication is often the barrier, not your scientific understanding.

# Our workshops cover

WHEN scientific communication is confusing

WHY it's confusing

HOW to fix the problems

...and apply these fixes to 20.109 assignments

# How the workshops will go

1. Discuss real examples
2. Derive principles and strategies
3. Practice
4. Go home with a checklist/rubric

Practice with a fellow at the  | Communication Lab !

The MIT BE logo consists of the text 'MIT BE' in a bold, sans-serif font, with 'MIT' in black and 'BE' in green. Below 'MIT BE' is the text 'BIOLOGICAL ENGINEERING' in a smaller, black, sans-serif font. A vertical line is positioned to the right of the logo.

# Designing Effective Figures

20.109 Spring 2018 Communication Workshop 1

Dr. Sean Clarke and Dr. Perna Bhargava

MIT **BE**  
BIOLOGICAL ENGINEERING

Communication Lab

[be.mit.edu/communicationlab](https://be.mit.edu/communicationlab)

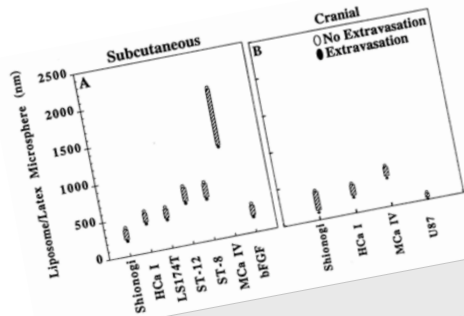
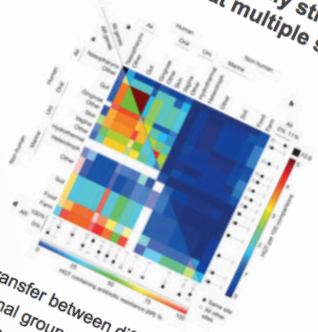


Figure 1

The vascular pore cutoff size for six different types of tumors grown in the dorsal of tumors grown in the cranial window (B) was evaluated. The solid circles represent extravasation at the indicated long-circulating liposome or latex bead size. The size range below the indicated liposome/latex bead size indicates the size range below which extravasated and the first particle that did not extravasate indicates the vascular pore cutoff size range. The majority of tumors have a vascular pore cutoff size range of 100-200 nm (hatched bar). The interaction of the tumor with the vessels (bFGF) when grown subcutaneously in the dorsal chamber. The interaction of the tumor with the vessels (bFGF) when grown in the cranial window (A). Comparison of bFGF-induced vessels (bFGF) with the subcutaneous microenvironment (A). Comparison of bFGF-induced vessels (bFGF) with the cranial microenvironment (B) leads to a smaller pore size than the interaction of bFGF with the subcutaneous microenvironment (A). Comparison of bFGF-induced vessels (bFGF) with the cranial microenvironment (B) demonstrates that the presence of bFGF alone can lead to pores of 100-200 nm.

Figure 3: HGT is ecologically structured by functional class and at multiple spatial scales



The frequency of transfer between different environments is shown for all functional groups (a, b) and for antibiotic resistance (AR) genes only (c, d). Box widths indicate the number of genomes from each environment. a, When all genes...

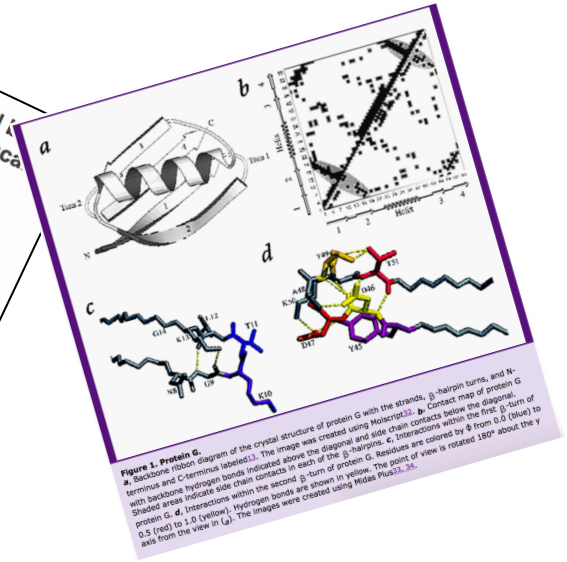
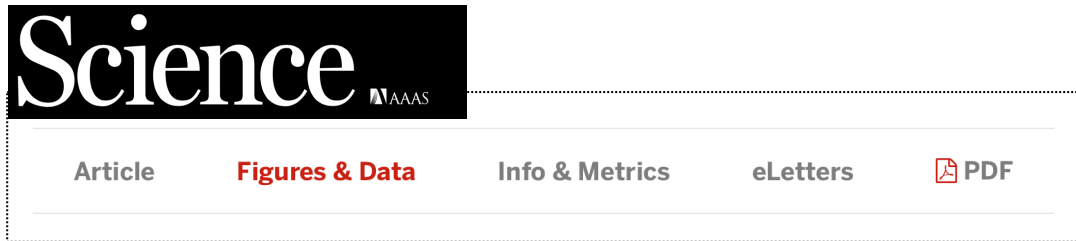


Figure 1. Protein G. a, Backbone ribbon diagram of the crystal structure of protein G with the strands,  $\beta$ -hairpin turns, and N-terminus and C-terminus labeled. The image was created using Molscript(3). b, Contact map of protein G with backbone hydrogen bonds indicated above the diagonal and side chain contacts below the diagonal. Shaded areas indicate side chain contacts in each of the  $\beta$ -hairpins. c, Interactions within the first  $\beta$ -turn of protein G. d, Interactions within the second  $\beta$ -turn of protein G. Residues are colored by  $\phi$  from 0.0 (blue) to 0.5 (red) to 1.0 (yellow). Hydrogen bonds are shown in yellow. The point of view is rotated 180 degrees about the y-axis from the view in (a). The images were created using Molscript(3) and PyMol(4).

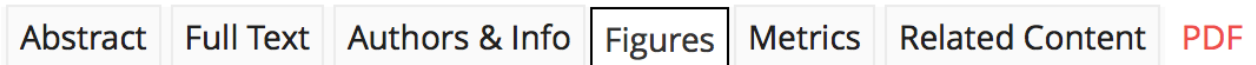
# Figures (and captions)

Why start here?

Experts and many audiences may ONLY READ your title, abstract, and FIGURES.



PNAS



So let's make them

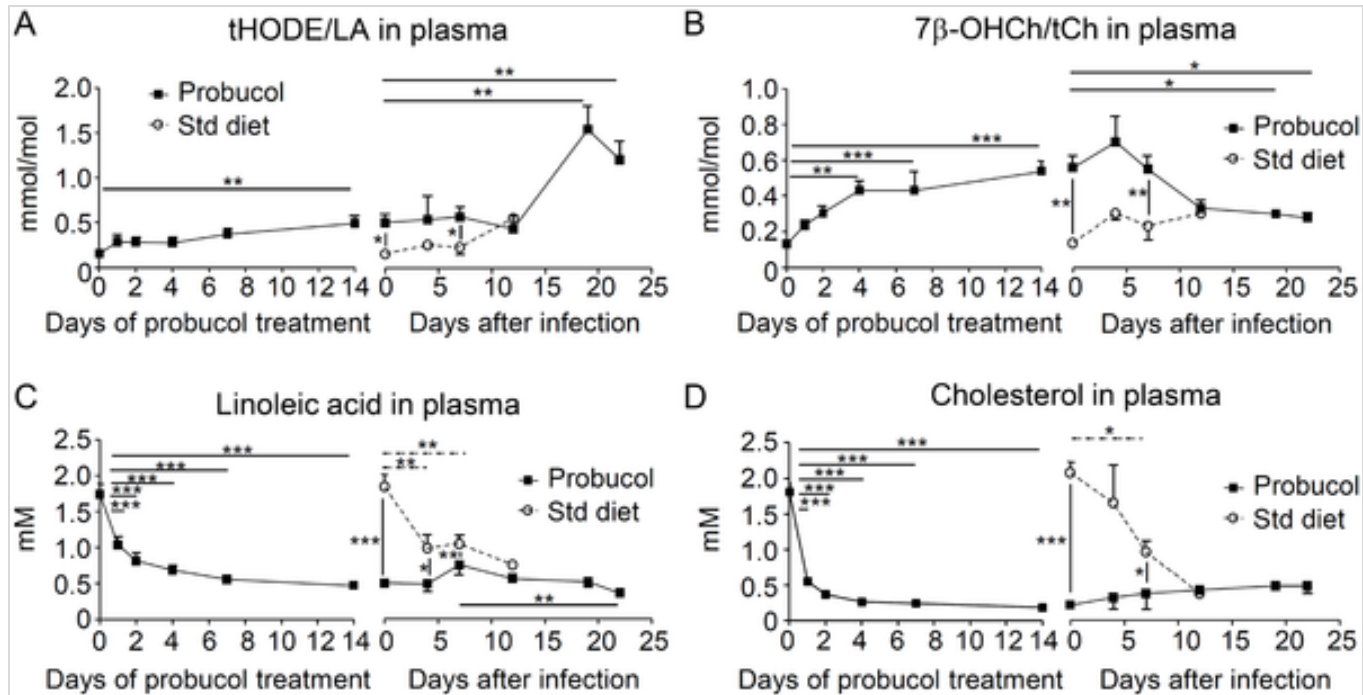
compelling

honest

supporting your story



# Activity: Identify the basic figure components



**Fig 4. The ratios of lipid peroxidation products to parent lipids in plasma increased after probucol pre-treatment.**

Six-week-old C57BL/6J mice were treated with 1% w/w probucol in the diet for 2 weeks and then infected with 0.2 mL of  $1 \times 10^5$  erythrocytes/mL infected with *Plasmodium yoelii* XL-17. Plasma samples were obtained at day 0, 1, 2, 4, 7, and 14 after starting the probucol diet (n = 5 per group) and at day 0, 4, 7, 12, 19, and 22 post-infection (n = 2 to 7). The ratio of total hydroxyoctadecadienoic acid (HODE), a peroxidation product of linoleic acid (LA), to linoleic acid (tHODE/LA) in plasma (A) and the ratio of 7β-hydroxycholesterol (7β-OHCh), a peroxidation product of cholesterol, to total cholesterol (7β-OHCh/tCh) in plasma (B) were measured. The concentration of LA (C) and tCh (D) were measured by using gas chromatography-mass spectrometry (GC-MS). All data are expressed as mean  $\pm$  SE. Statistical analysis was carried out by analysis of variance (ANOVA). \* $p < 0.05$ , \*\* $p < 0.025$ , and \*\*\* $p < 0.001$ . The solid bars indicate the significant changes in probucol-treated groups and the dotted bars indicate the significant changes in standard (Std) diet-fed mice.

# Basic figure components

Figure = message + data

## Choice of data

- Only data that are critical to the conclusion

## Presentation choices

- Type of graph or display, legends & labeling, design choices
- Uncluttered
- Allow quick evaluation of conclusions, without referring to legend or caption.

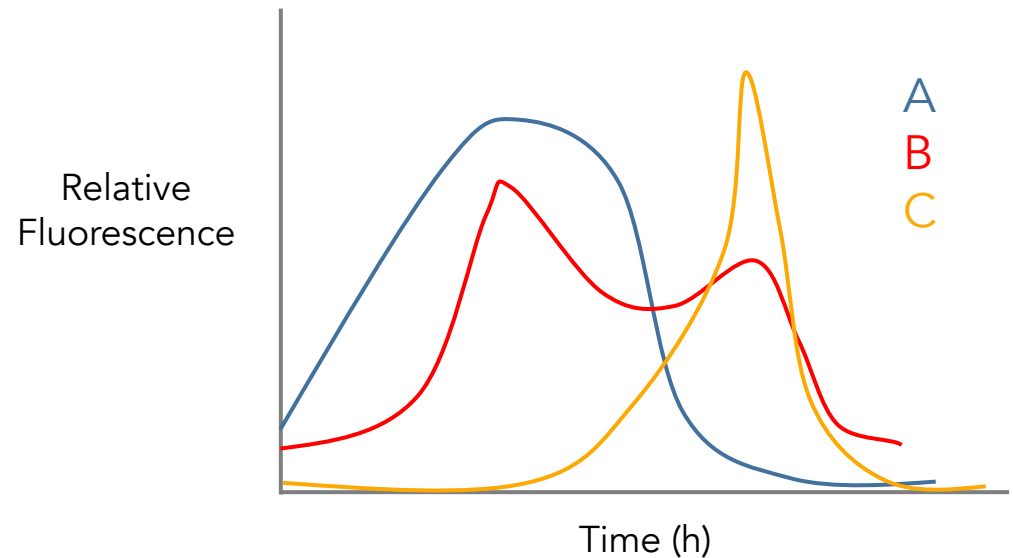


Fig. 1: A, B, and C have different dynamics under Condition X. A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control.

# Basic figure components

Figure = message + data

## Title

- Take-home *message* of the figure
- What conclusion should the reader evaluate when looking at the figure?

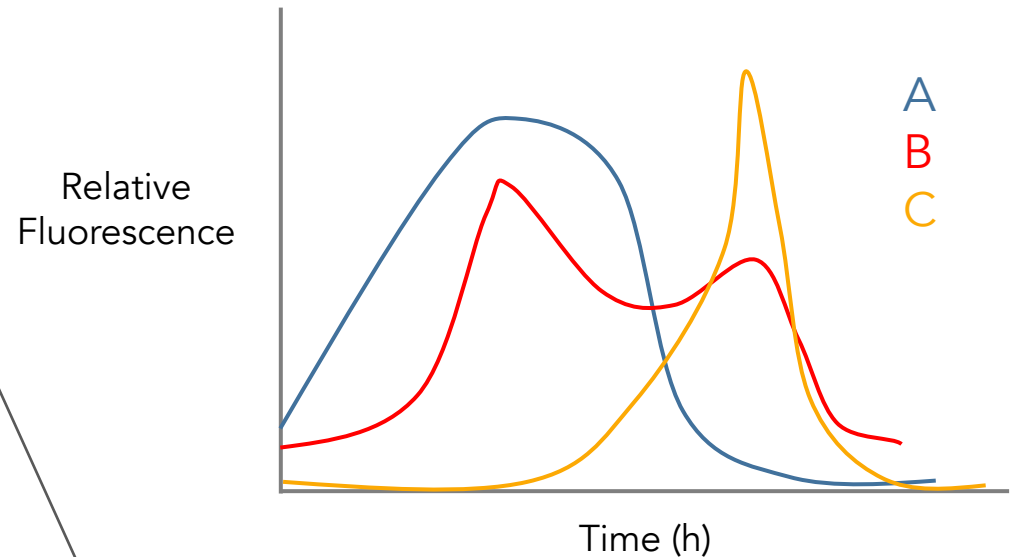
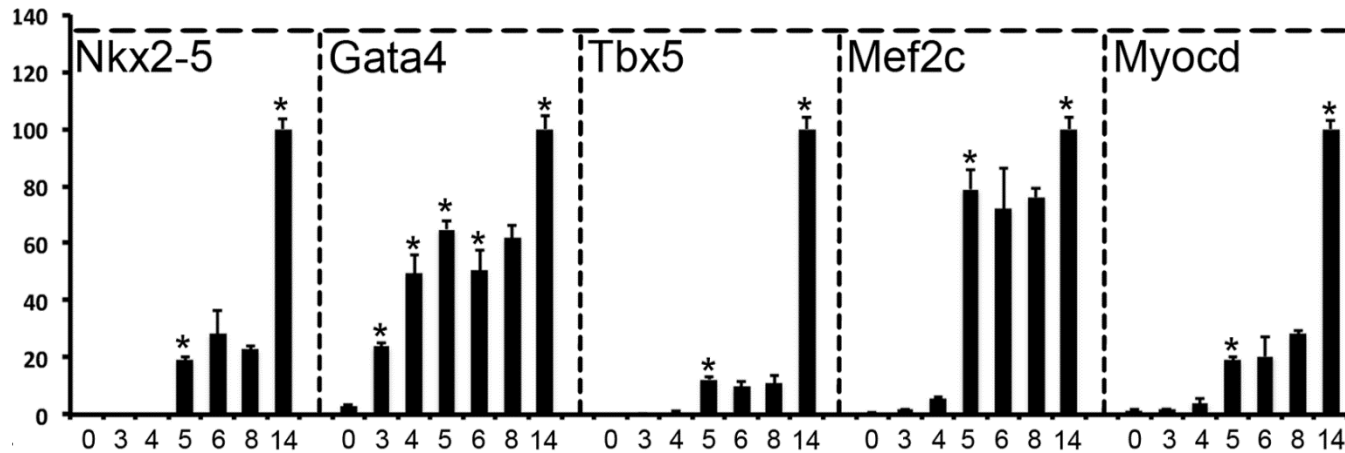


Fig. 1: A, B, and C have different dynamics under Condition X. A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control.

# Message: Use titles to state a figure's message, not the method

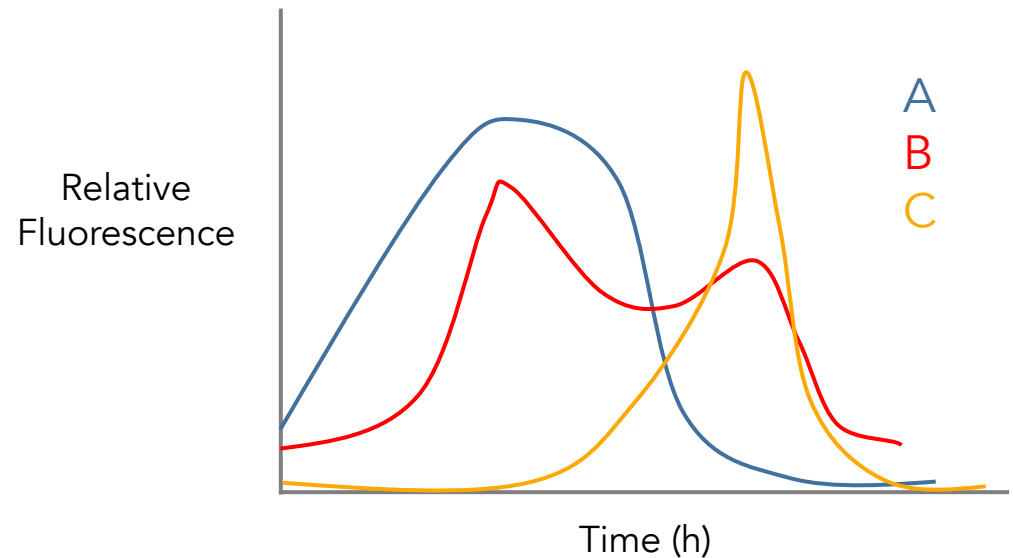


Gene expression analysis performed on differentiating mouse iPS cells

Expression of early cardiac transcription factors increases over time in differentiating mouse iPS cells

# Basic figure components

Figure = message + data



## Caption

- Descriptive, not explanatory/interpretive
- Only enough methodological detail to make it clear how results were obtained.

Fig. 1: A, B, and C have different dynamics under Condition X. A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control.

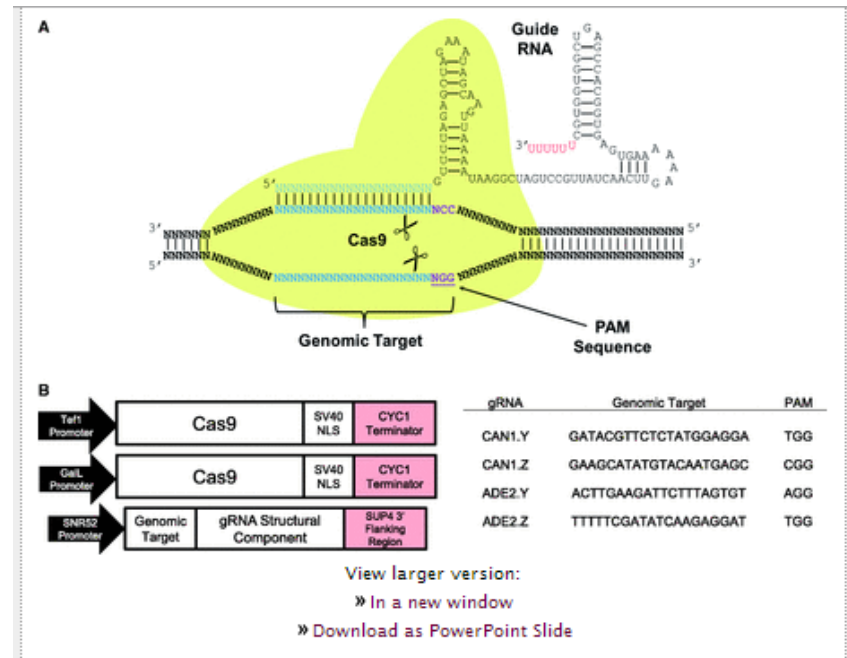
# All the figures you make need all of these components

Schematics

Diagrams

Photos

...count as figures too.

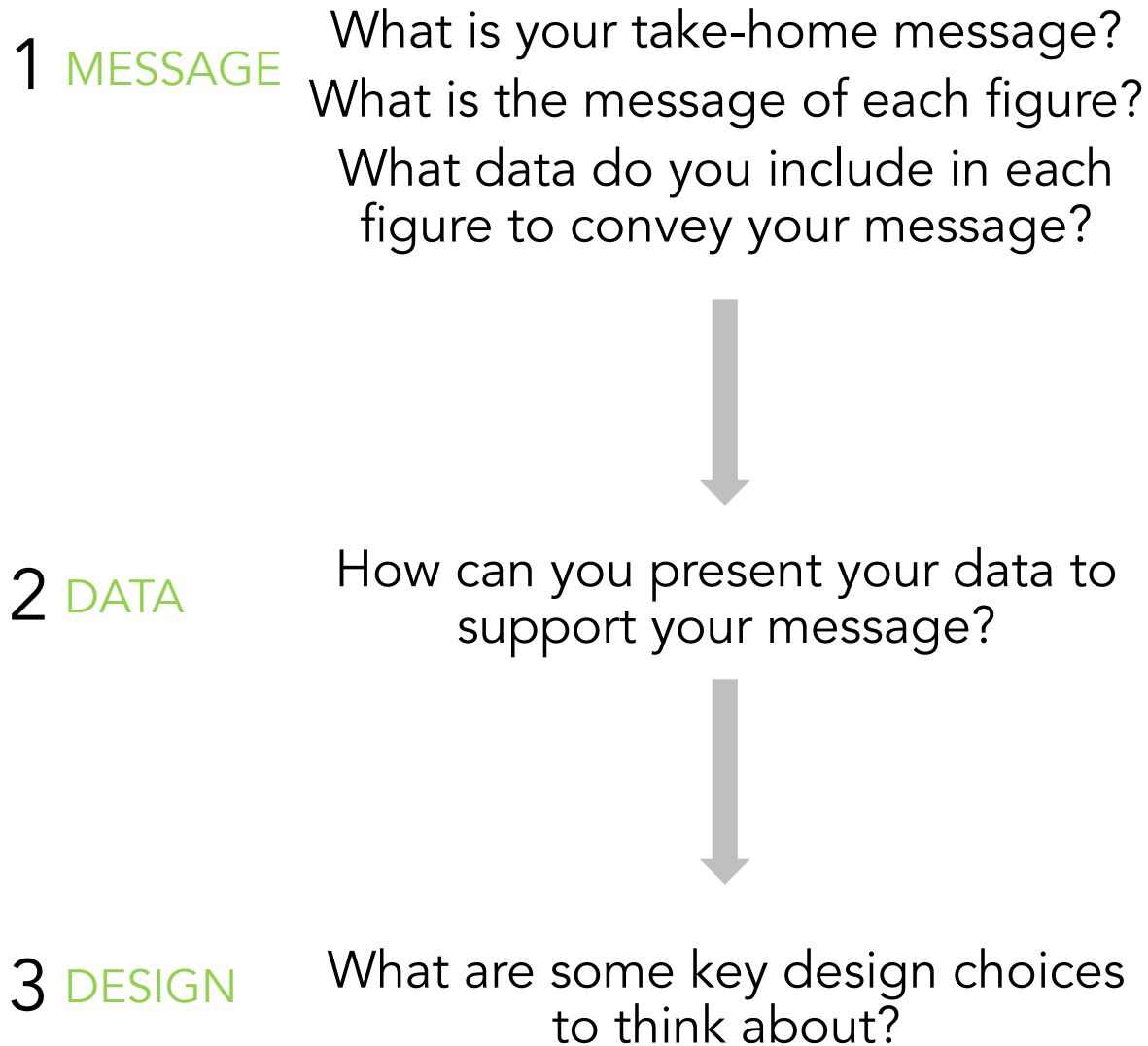


**Figure 1.**

Diagram of Cas9 complex and schematic of genetic constructs. (A) Illustration of Cas9 protein interacting with CRISPR gRNA to direct endonuclease activity proximal to the PAM sequence. (B) Design of the Cas9 and gRNA constructs. Cas9 gene contained a SV40 nuclear localization signal and was expressed under the Gal-L inducible promoter in CAN1 experiments and the TEF1 constitutive promoter in ADE2 experiments. The gRNA was

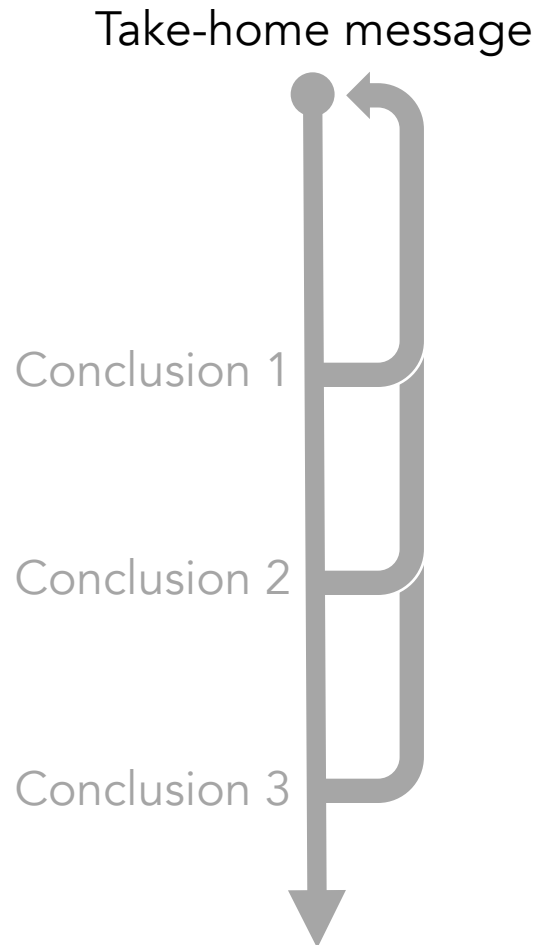
*DiCarlo et al., 2013 Nuc. Acids. Res.*

# Steps to turn your pile of data into figures...

- 1 MESSAGE**  
What is your take-home message?  
What is the message of each figure?  
What data do you include in each figure to convey your message?
  - 2 DATA**  
How can you present your data to support your message?
  - 3 DESIGN**  
What are some key design choices to think about?
- 
- ```
graph TD; A["1 MESSAGE  
What is your take-home message?  
What is the message of each figure?  
What data do you include in each figure to convey your message?"] --> B["2 DATA  
How can you present your data to support your message?"]; B --> C["3 DESIGN  
What are some key design choices to think about?"];
```

# Message: Create a single storyline.

Identify your take-home message; everything else leads to it.





# Message: To find your story, organize your messages and figures.

Rearrange until you've created a logical series of conclusions.

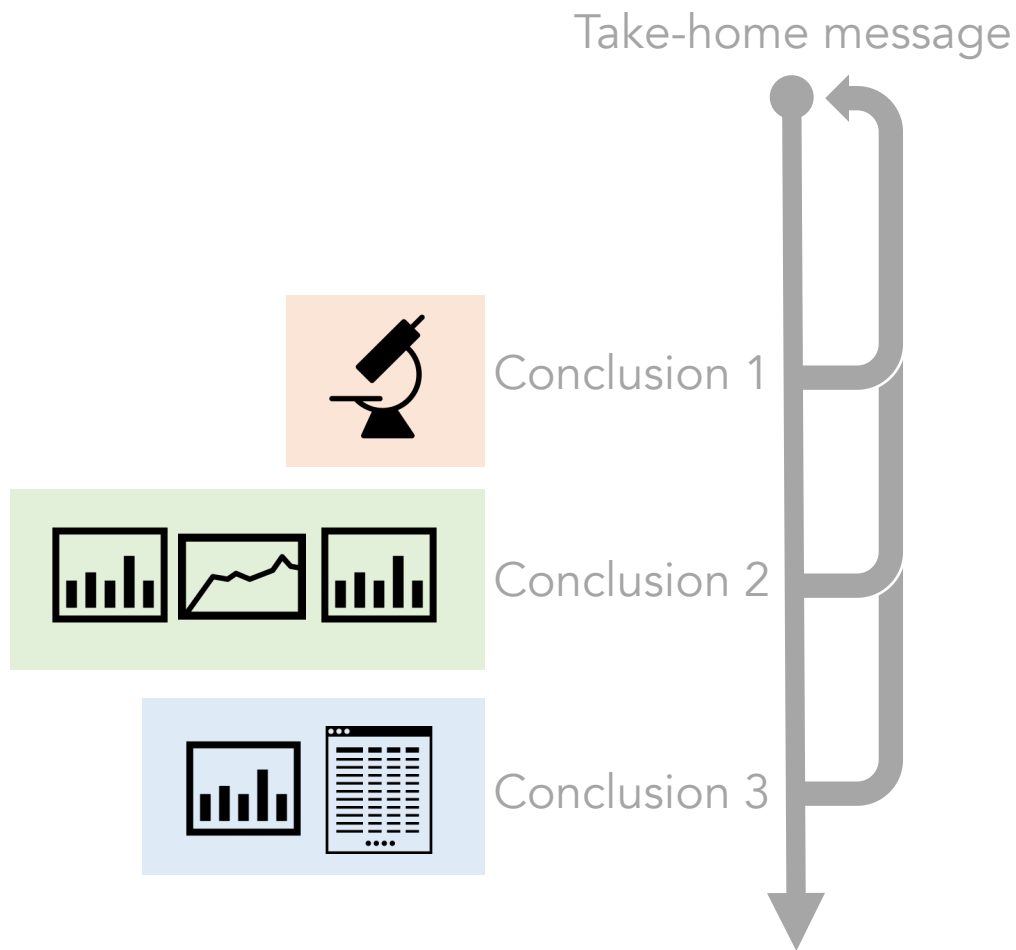


# Message: Organize your Figures into modules.

Identify modules that correspond to conclusions.



# Message: Modules correspond to conclusions



**Message:** All the data in a figure should support one clear message.

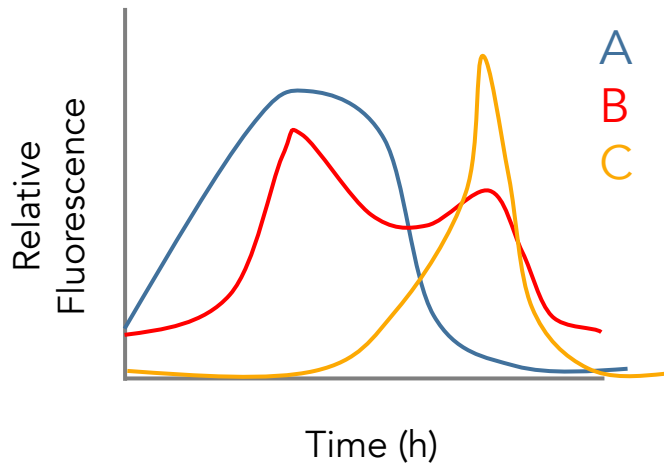


Fig. 1: A, B, and C have different dynamics under Condition X. A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control.

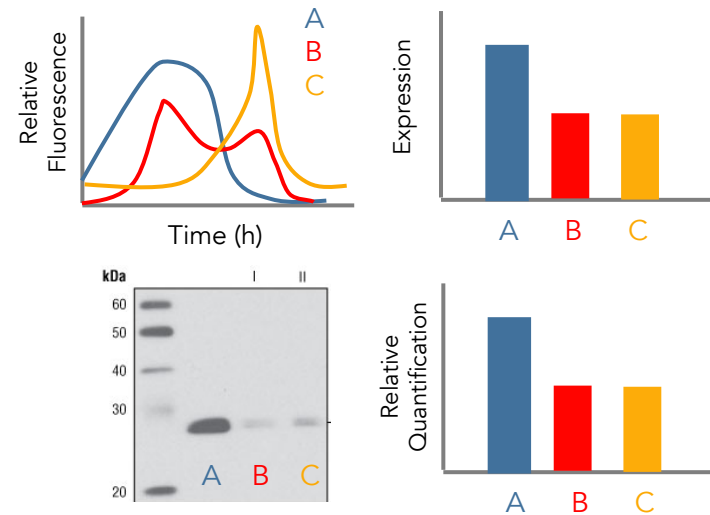
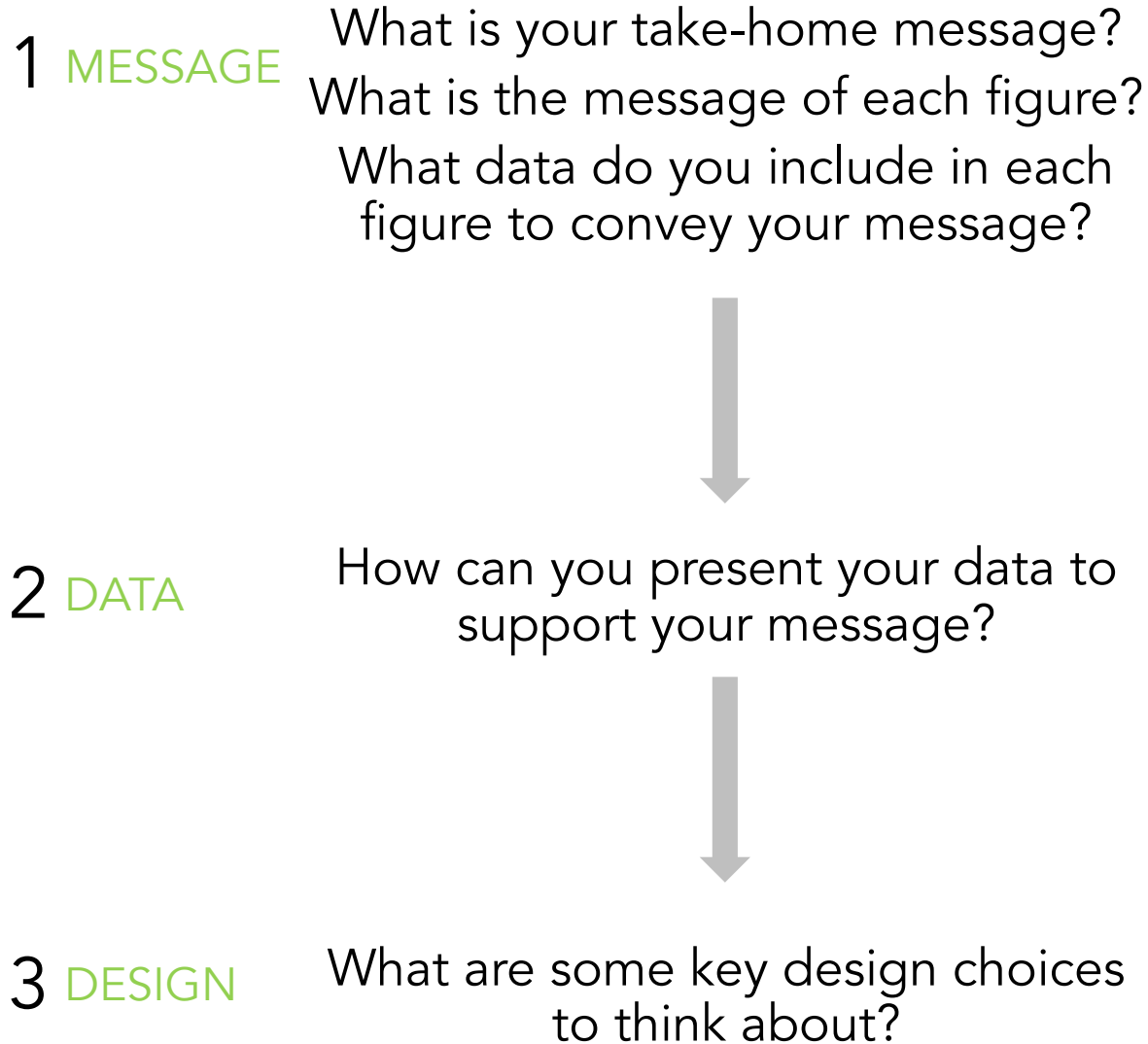


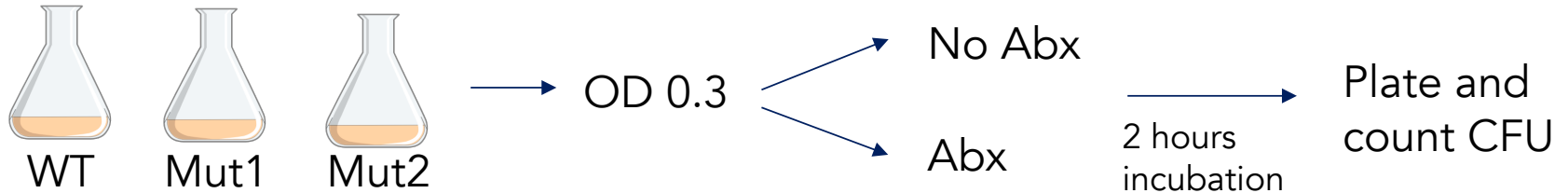
Fig. 1: A, B, and C have different dynamics under Condition X. A) A, B, and C were sampled using Method 1 and their fluorescence quantified with Method 2. Fluorescence data normalized to negative control. B) Gene expression data of samples A, B, and C, under condition X. Samples were collected at time T. C) Western blot analysis of samples A, B, and C, under condition X. D) Quantification of Western Blot.

# Steps to turn your pile of data into figures...

- 1 MESSAGE**  
What is your take-home message?  
What is the message of each figure?  
What data do you include in each figure to convey your message?
  - 2 DATA**  
How can you present your data to support your message?
  - 3 DESIGN**  
What are some key design choices to think about?
- 
- ```
graph TD; A["1 MESSAGE  
What is your take-home message?  
What is the message of each figure?  
What data do you include in each figure to convey your message?"] --> B["2 DATA  
How can you present your data to support your message?"]; B --> C["3 DESIGN  
What are some key design choices to think about?"];
```

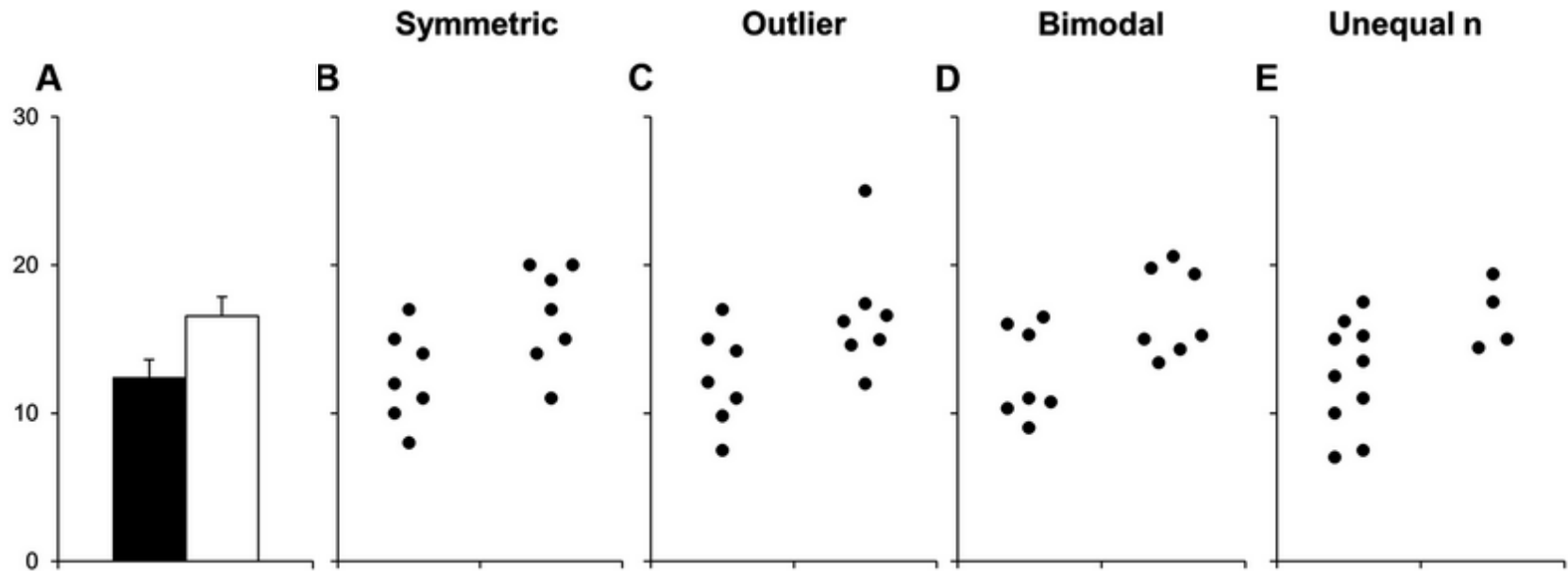
# Activity:

What stories can we tell with this data?  
How do they affect figure choices?



Strain	Dosage	Replicate 1	Replicate 2	Replicate 3	Average (CFU)
WT	Baseline	1.8e5	3.2e5	7.8e5	4.3e5
WT	No Abx	1.0e9	1.3e9	8.0e8	1.0e9
WT	Abx	2.3e2	2.8e2	5.5e2	3.5e2
Mut1	Baseline	2.5e5	8.3e5	4.6e5	5.1e5
Mut1	No Abx	5.5e7	2.3e7	1.1e7	3.0e7
Mut1	Abx	4.3e5	7.5e5	2.2e5	4.7e5
Mut2	Baseline	5.3e5	3.2e5	1.3e5	3.3e5
Mut2	No Abx	3.4e9	2.2e9	3.0e9	2.9e9
Mut2	Abx	2.2e9	5.3e9	1.2e9	2.9e9

**Data:** Consider which plot type best allows the reader to evaluate your conclusion.

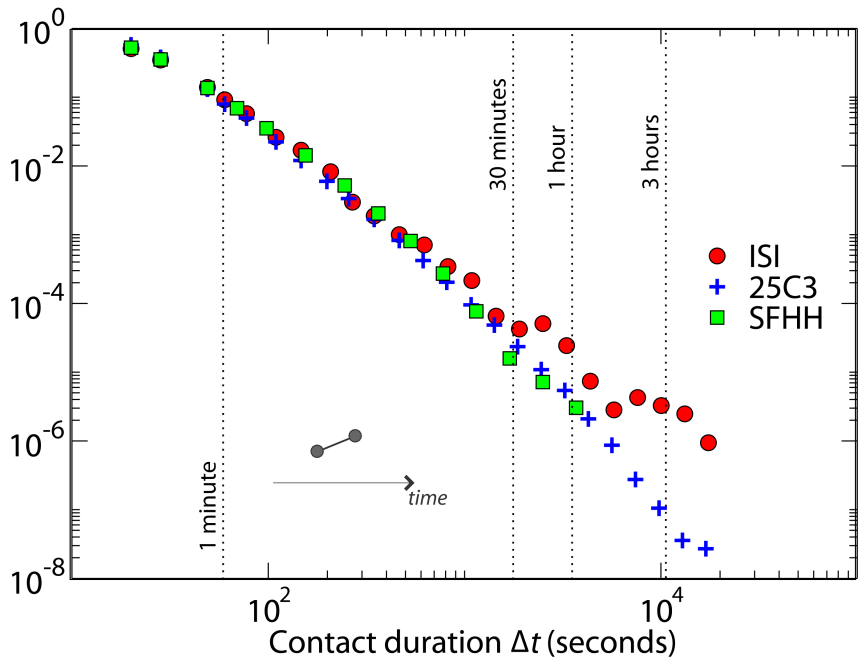


Bar charts  
assume a  
Gaussian  
distribution.

Scatter plots reveal the true distribution.

# Data: Only show as much data as you need to convey your message.

*Probability distribution of human interactions at 3 conferences*



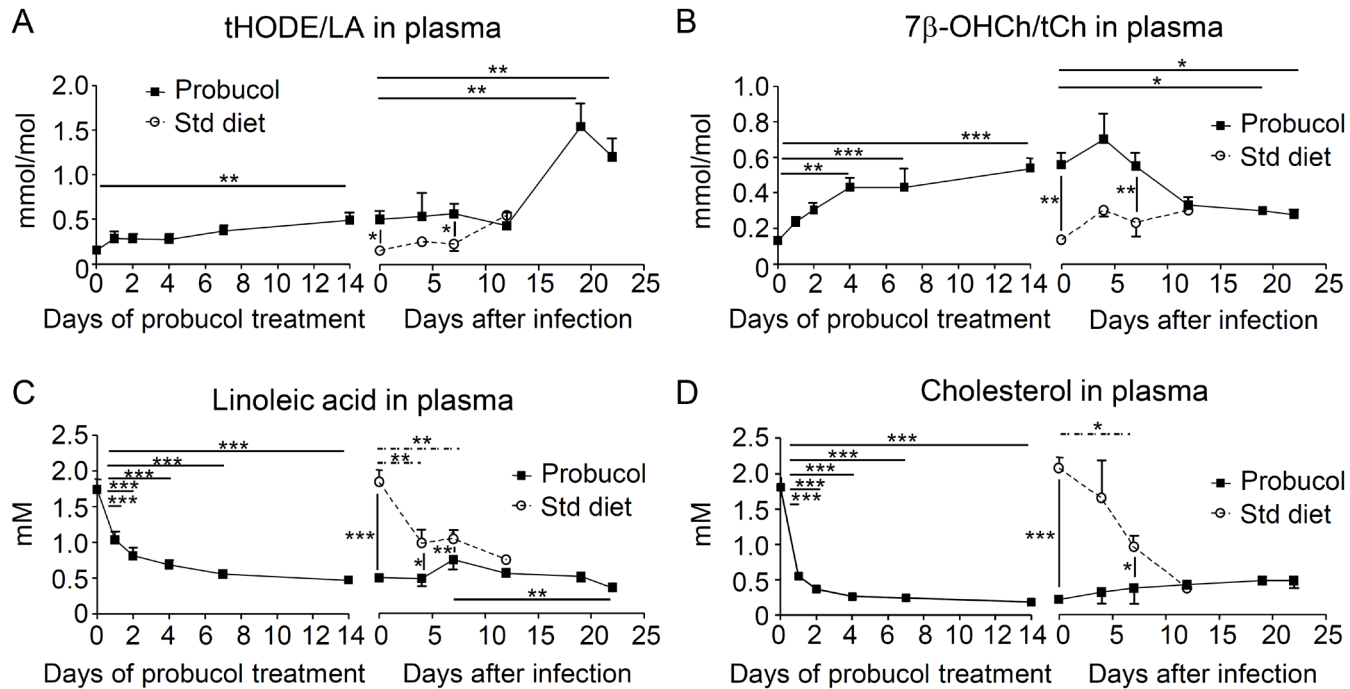
*At a poster session, 50% of your audience walks away after 20 seconds*



Time talking	Probability listening
0 sec	100%
20 sec	50%
1 min	10%
2 min	5%
5 min	<1%



# Activity: Improve this published figure

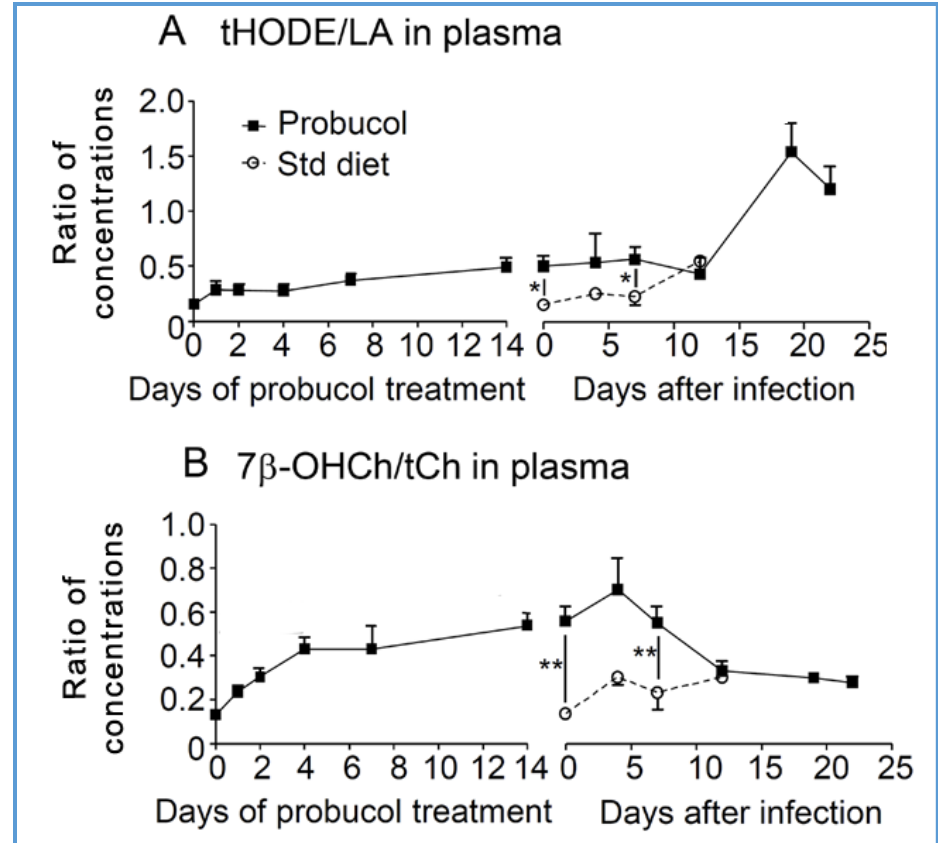
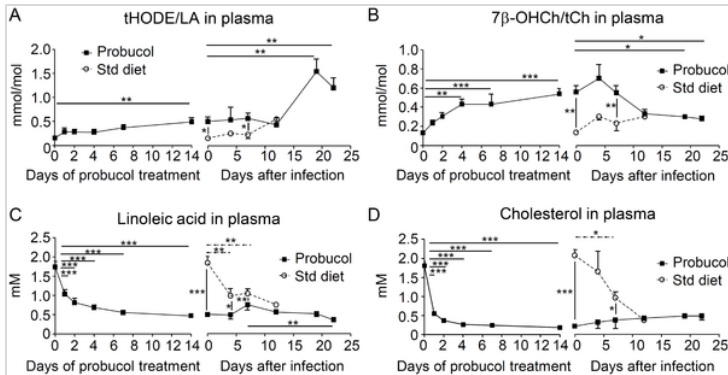


**Fig 4. The ratios of lipid peroxidation products to parent lipids in plasma increased after probucol pre-treatment.** Six-week-old C57BL/6J mice were treated with 1% w/w probucol in the diet for 2 weeks and then infected with 0.2 mL of  $1 \times 10^5$  erythrocytes/mL infected with *Plasmodium yoelii* XL-17. Plasma samples were obtained at day 0, 1, 2, 4, 7, and 14 after starting the probucol diet (n = 5 per group) and at day 0, 4, 7, 12, 19, and 22 post-infection (n = 2 to 7). The ratio of total hydroxyoctadecadienoic acid (HODE), a peroxidation product of linoleic acid (LA), to linoleic acid (tHODE/LA) in plasma (A) and the ratio of 7β-hydroxycholesterol (7β-OHCh), a peroxidation product of cholesterol, to total cholesterol (7β-OHCh/tCh) in plasma (B) were measured. The concentration of LA (C) and tCh (D) were measured by using gas chromatography-mass spectrometry (GC-MS). All data are expressed as mean ± SE. Statistical analysis was carried out by analysis of variance (ANOVA). \*p < 0.05, \*\*p < 0.025, and \*\*\*p < 0.001. The solid bars indicate the significant changes in probucol-treated groups and the dotted bars indicate the significant changes in standard (Std) diet-fed mice.

# Evaluate these figure choices

- Is the central message validated by the data shown?
- Which data are irrelevant?
- Are there any data/labels missing?
- What could be done to better highlight the most important data?
- Is there a better way to present the data?
- Do the statistics actually add anything here?

Only include the minimum information necessary to express the message honestly.

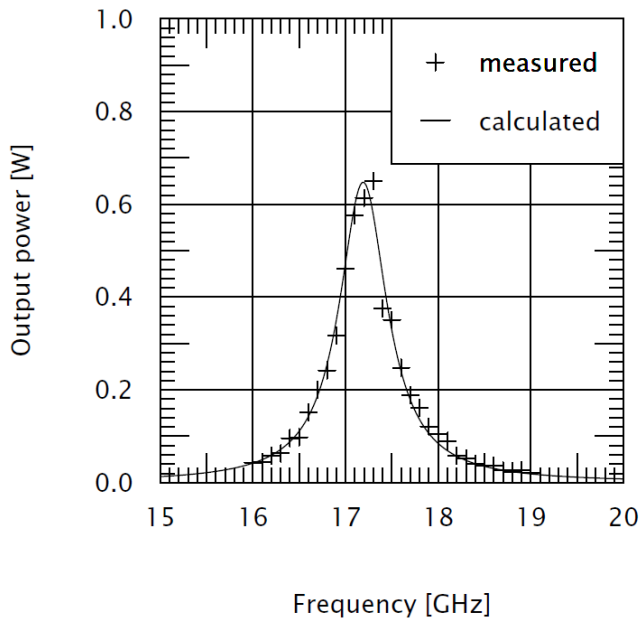


# Steps to turn your pile of data into figures...

- 1 MESSAGE**  
What is your take-home message?  
What is the message of each figure?  
What data do you include in each figure to convey your message?
- 2 DATA**  
How can you present your data to support your message?
- 3 DESIGN**  
What are some key design choices to think about?

# Design: Maximize signal-to-noise

Reward yourself for cutting things

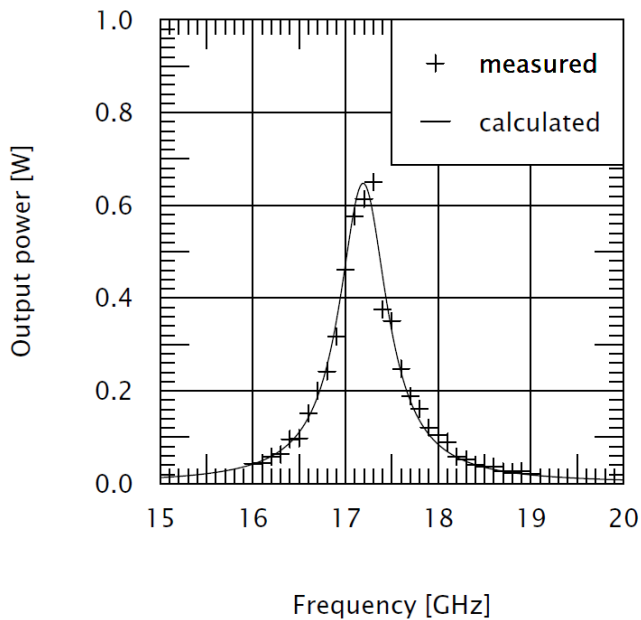


# Design: Maximize signal-to-noise

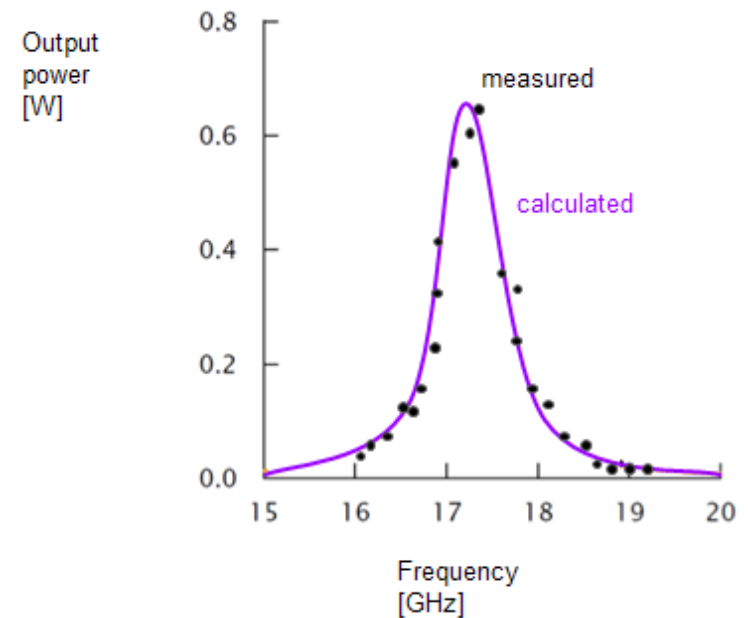
State your message.

Eliminate anything that distracts from it.

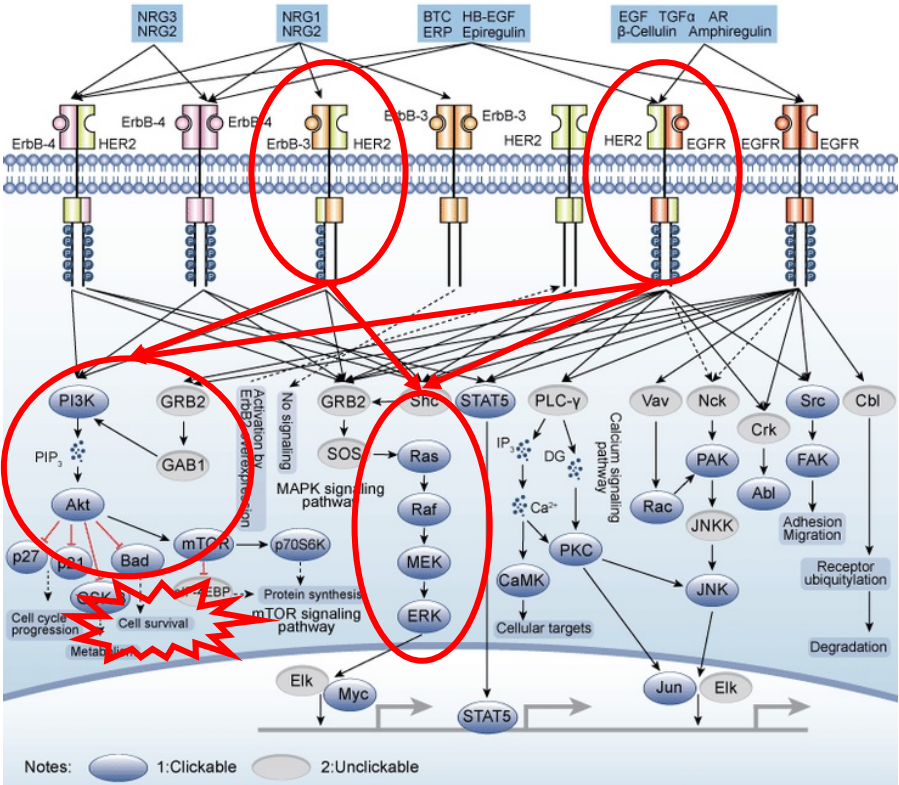
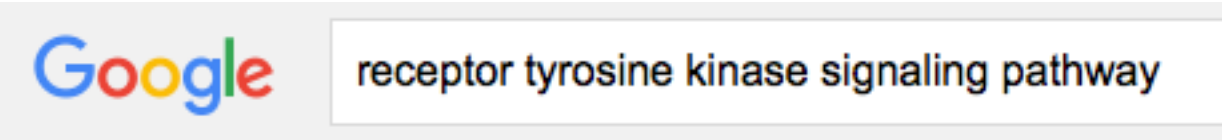
Power vs. frequency



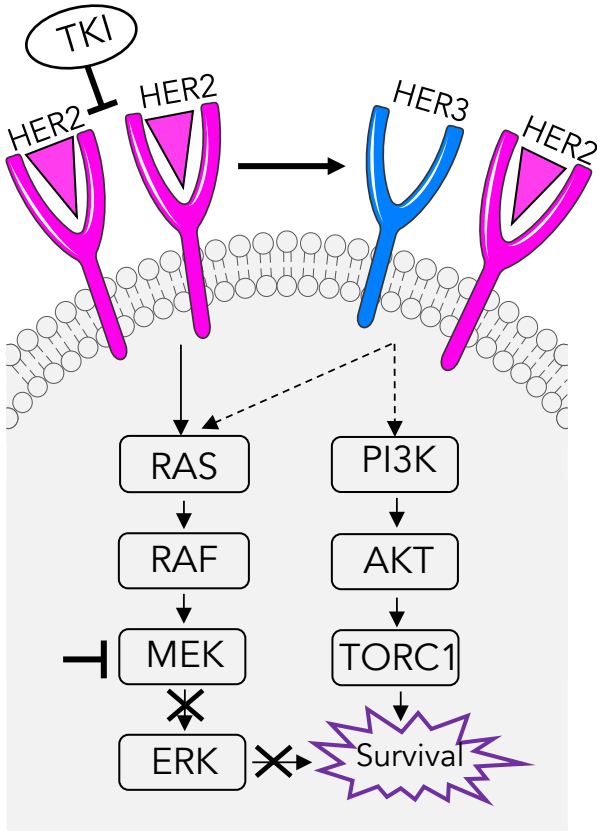
True power spectrum agrees with prediction



# Design: Make schematics with a message



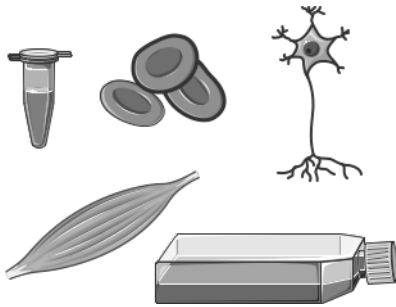
# Design: Schematics have messages too



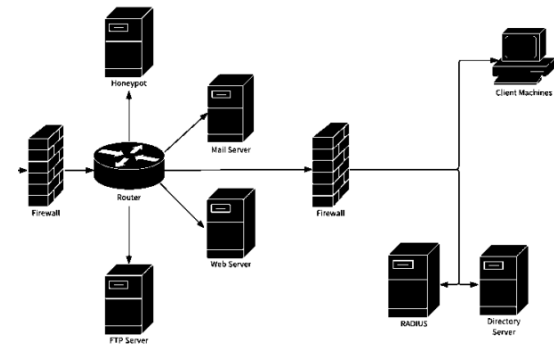


# Design: You don't need to reinvent the wheel

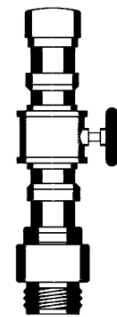
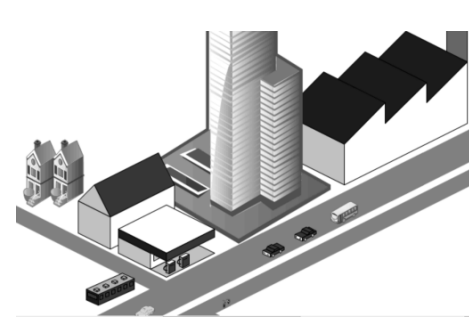
Servier Powerpoint Image Bank  
Biology, lab equipment  
(free)



MS Visio & Lucidchart  
Networks, engineering, circuits, charts  
(\$\$) & (free)



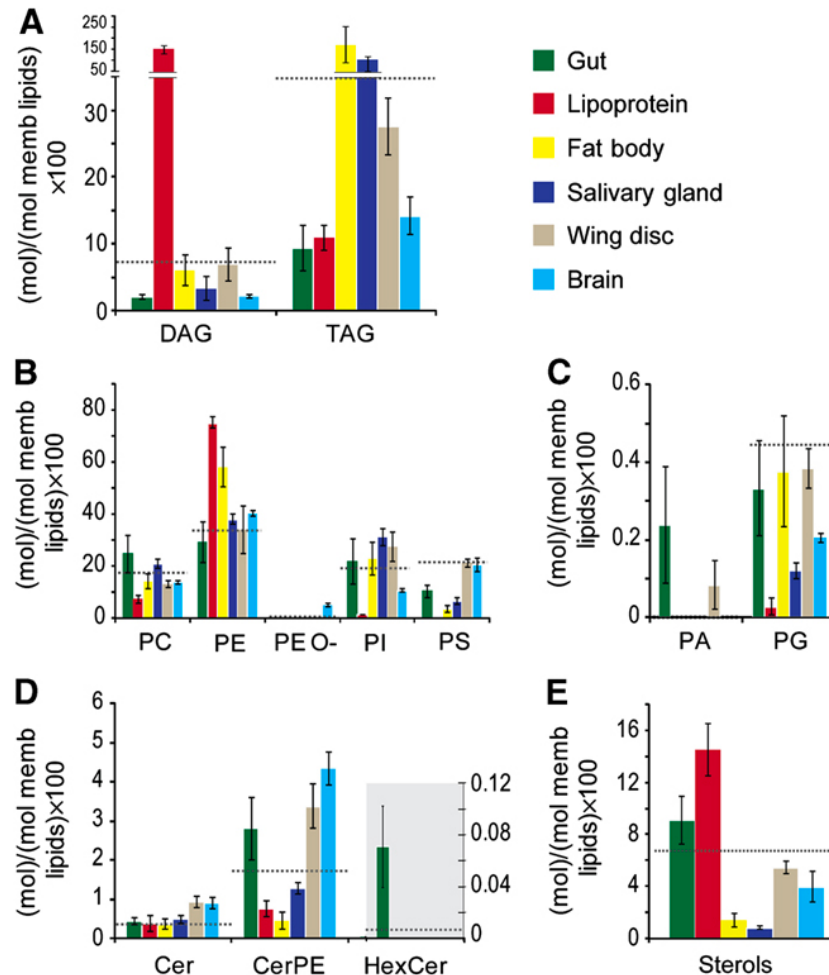
Noun Project  
Everything  
(free)



# Design: Maximize signal-to-noise

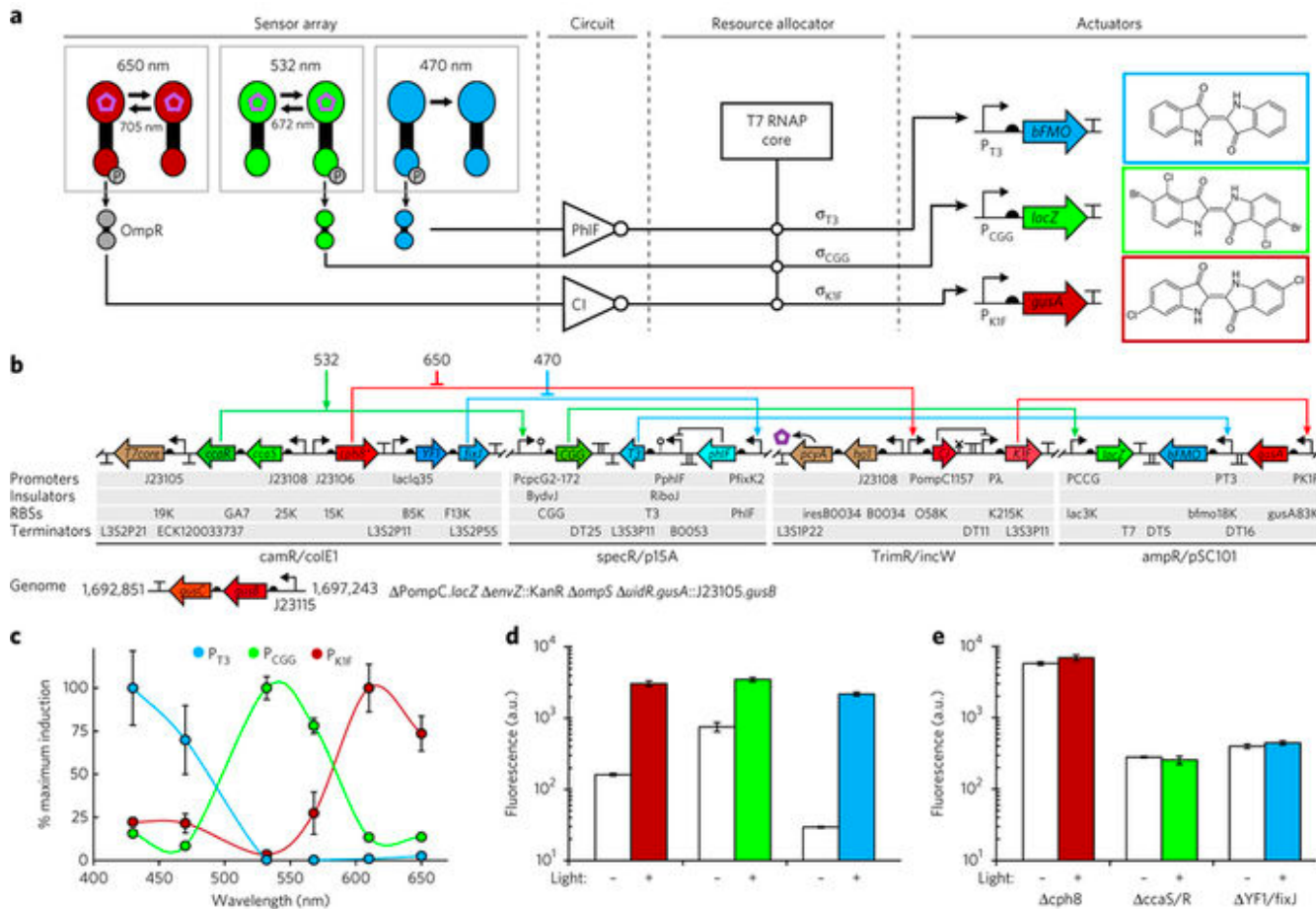
Minimize # style choices and be consistent

- Grayscale with a consistency of colors



# Design: Maximize signal-to-noise

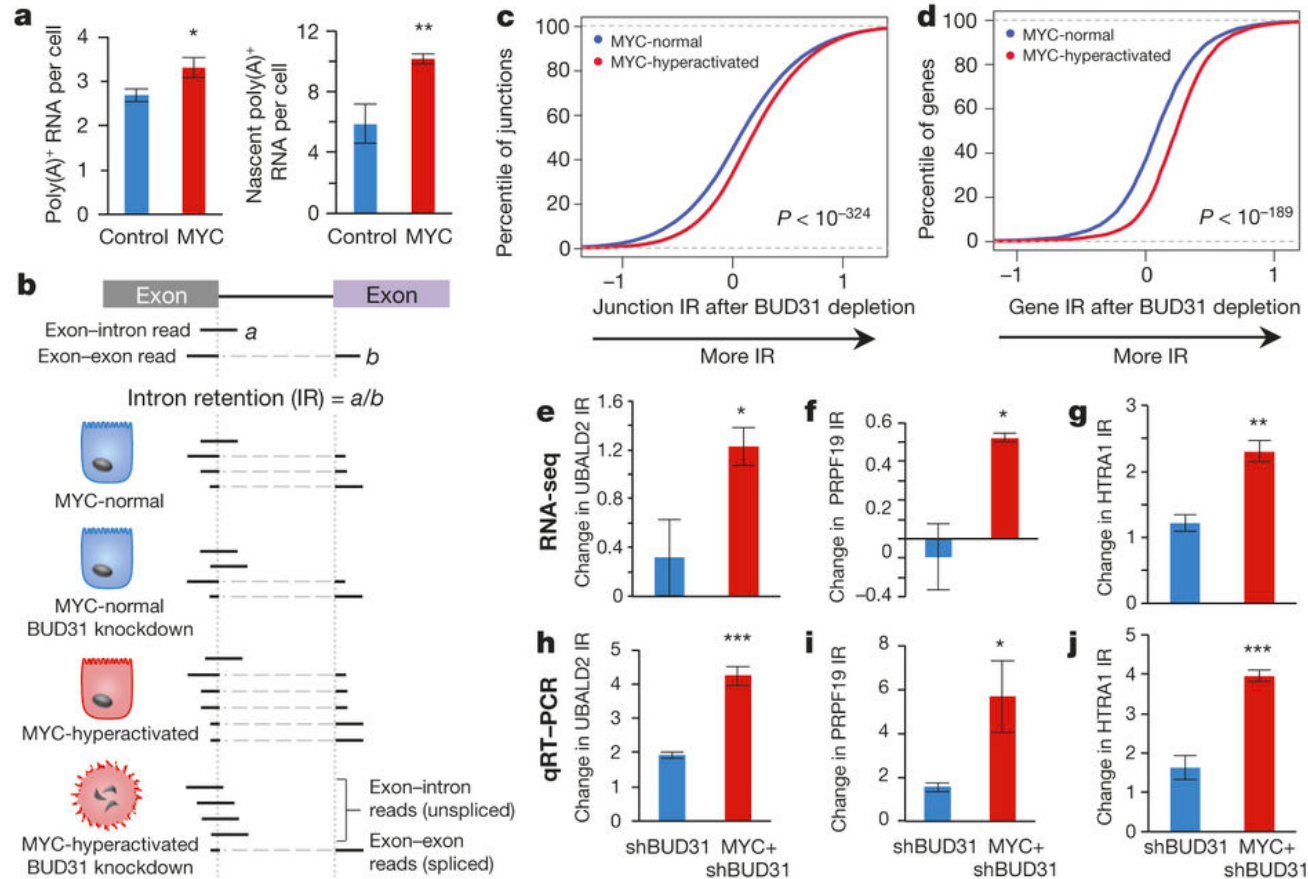
- Try Grayscale or consistent colors





# Design: Maximize signal-to-noise

Consider

Font and size  
Line thickness  
Alignment



# Steps to turn your pile of data into figures...

- 1 MESSAGE**
  - What is your take-home message?
  - What is the message of each figure?
  - What data do you include in each figure to convey your message?
- 2 DATA**
  - How can you present your data to support your message?
- 3 DESIGN**
  - What are some key design choices to think about?

# Go forth and figure

Wiki has this and more

Put these tips to work

Prerna and Sean in lab to answer questions

Next week we'll get concrete about Abstracts!

# Optimize your figures with these reminders

## High-level questions

- *Strategic purpose:*
  - What do you want to convey?
  - How will you and/or your audience use this figure?
- *Organizational structure:*
  - Where does this figure fit into the communication?
  - Why?

## Checklist

- Choice of data
- Type of plot for the data
- Title/Caption
  - Can figure stand alone?
- Consistent features
  - Fonts, spacing, colors
- Text amount and placement
- Scale, axes, tick marks
- Error analysis
- Ink-to-whitespace ratio

