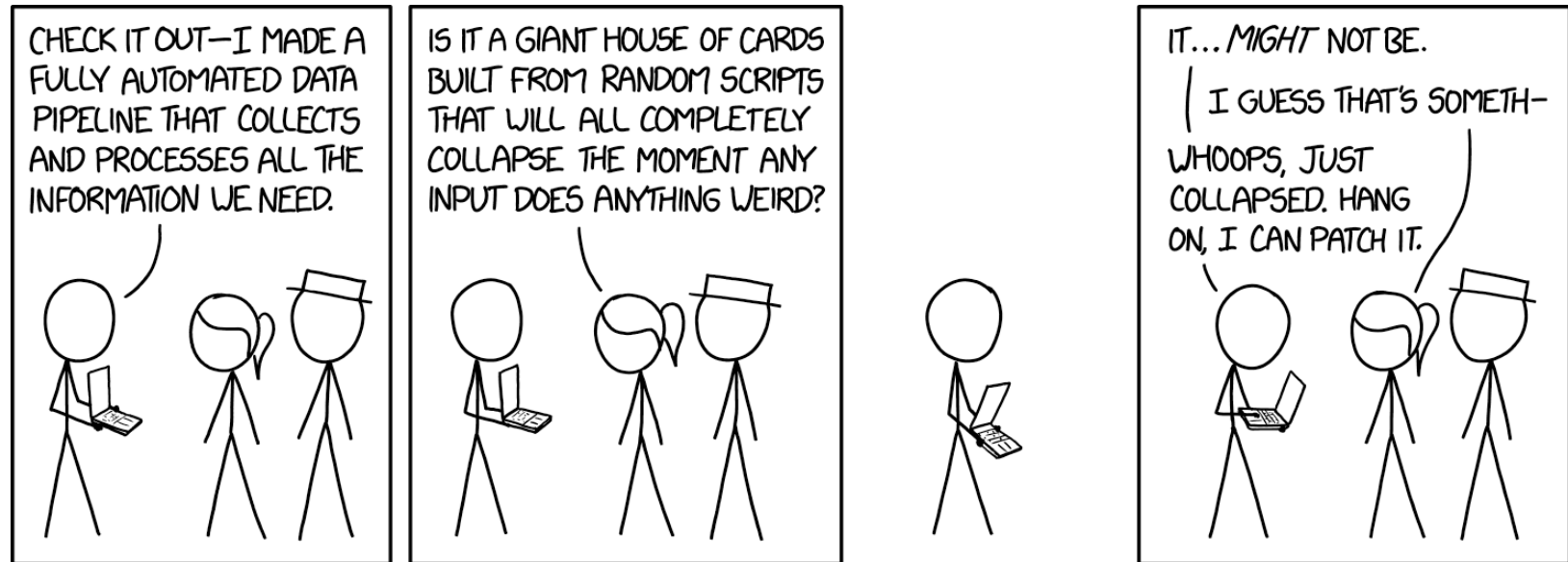


M1D1: Review small molecule microarray (SMM) technology

- Orientation quiz!
- Prelab discussion
- Walk through SMM procedure



xkcd

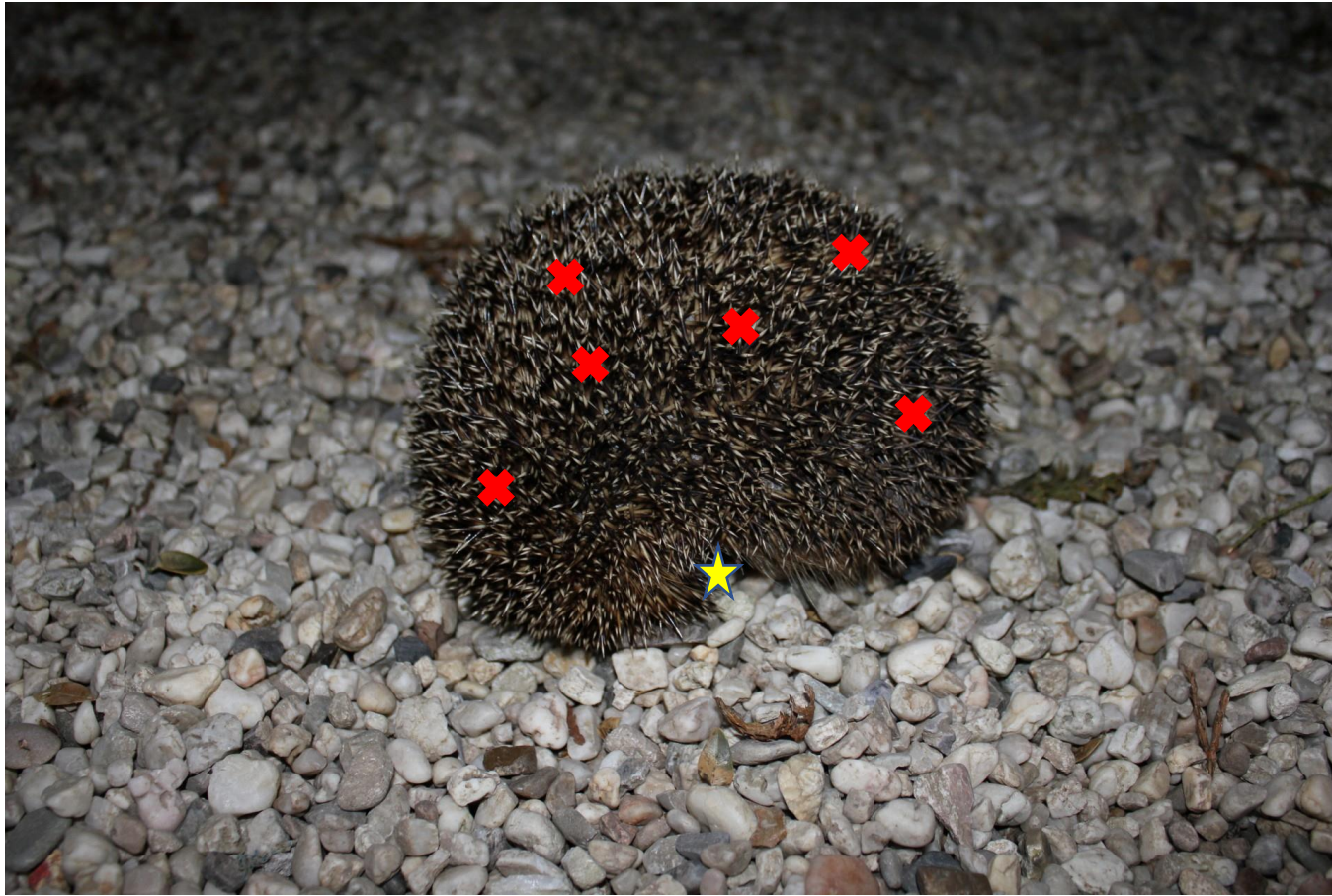
Mod 1: Major Assignments

- **Data summary (15%)**
 - In a team
 - Draft due 3/12, final revision due 3/20
 - Format: Bullet points, .PPTX
- **Research Talk (5%)**
 - Individual, submit video via gmail
 - Due 2/23 by 10pm
- **Lab quizzes (5% collectively)**
 - Individual (orientation quiz is exception)
- **Notebook (5% collectively)**
 - Due 3/4 by 10pm, graded by Christine
- **Blog (part of 5% Participation)**
 - Due 3/14 by 10pm

**I love deadlines.
I like the whooshing
sound they make as
they fly by.**

DOUGLAS ADAMS

What is an “undruggable” target?



Difficult targets may lack nice binding pockets, hence, undruggable

If only there was a way to find molecules appropriately shaped and sized that could squeeze and fit into difficult pockets.....

Mod 1 Background

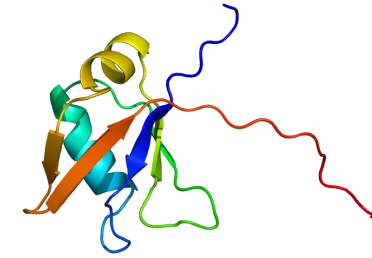
Overarching focus of Mod1: Drug discovery!

- We are studying the effects of small molecules on an “undruggable” target known to play a role in neurodegenerative disease.
 - Can small molecule interactions with our protein provide any biological insight?
 - Do any small molecules provide insight about potential therapeutics for our protein of interest?

Topics we'll cover today:

- What is TDP-43/ why is it an interesting drug target?
- What kind of drugs will be our focus?
- How did we screen for potential drugs in a previous semester?
- How are you going to follow up on that initial screen?

What is our target?



- TAR DNA-binding protein 43 (TDP-43)

Healthy State	Disease State
<ul style="list-style-type: none">- DNA & RNA binding protein- Mainly lives in the nucleus	<ul style="list-style-type: none">- Can mislocalize to the cytoplasm- Can form aggregates- Can be aberrantly modified- Linked to diseases like amyotrophic lateral sclerosis (ALS), frontotemporal dementia (FTD)

- Contains 4 Domains
 - N termini
 - C termini
 - 2 RNA binding domains

TDP-43



Your predecessors expressed a recombinant TDP43...

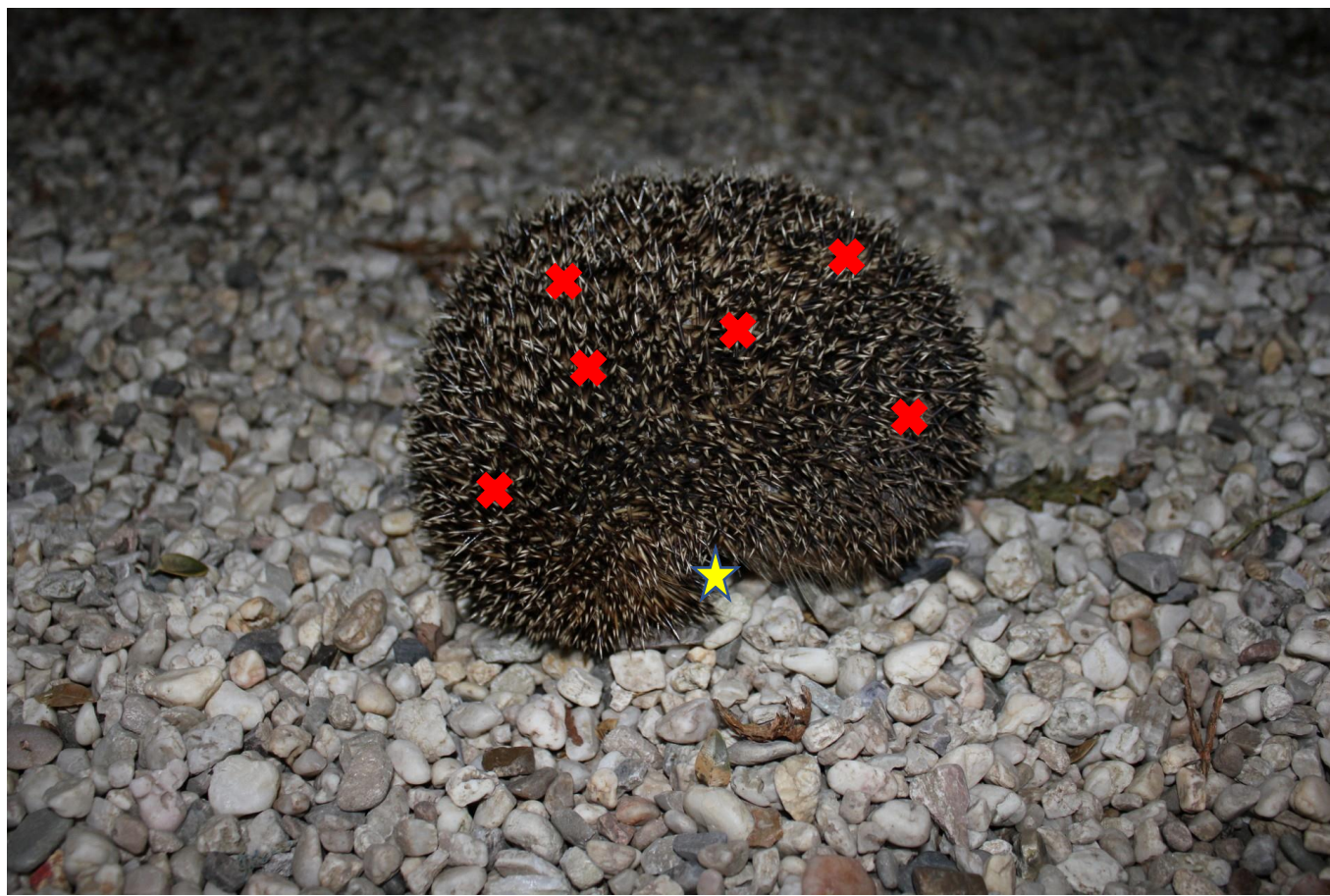


What are the pros and cons of using this construct?

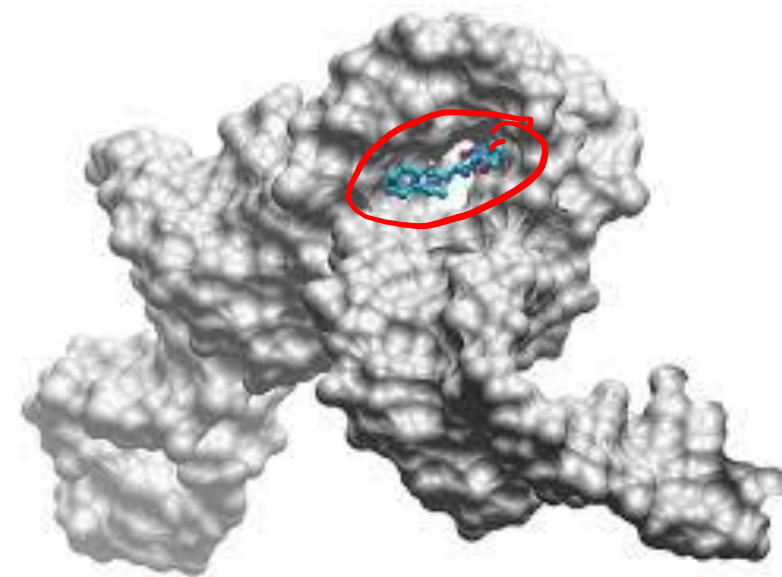
Pro: Can get more specific information about the mechanism of action of our SM

Con: Can cause misfolding, or creation of new binding pockets, less face validity

TDP-43 is an undruggable target



What are small molecules?



- Small molecules
 - $M_w < 1000$ Da
 - Natural or synthetic
 - Frequently comprised of Carbon/Nitrogen/Oxygen
- Why are they interesting probes/therapeutics?
 - Potential to cross membranes and target intracellular molecules
 - Designable/modifiable
 - Numerous possibilities for target interaction

Pro: Can increase the probability of a hit

Can target whole classes of molecules

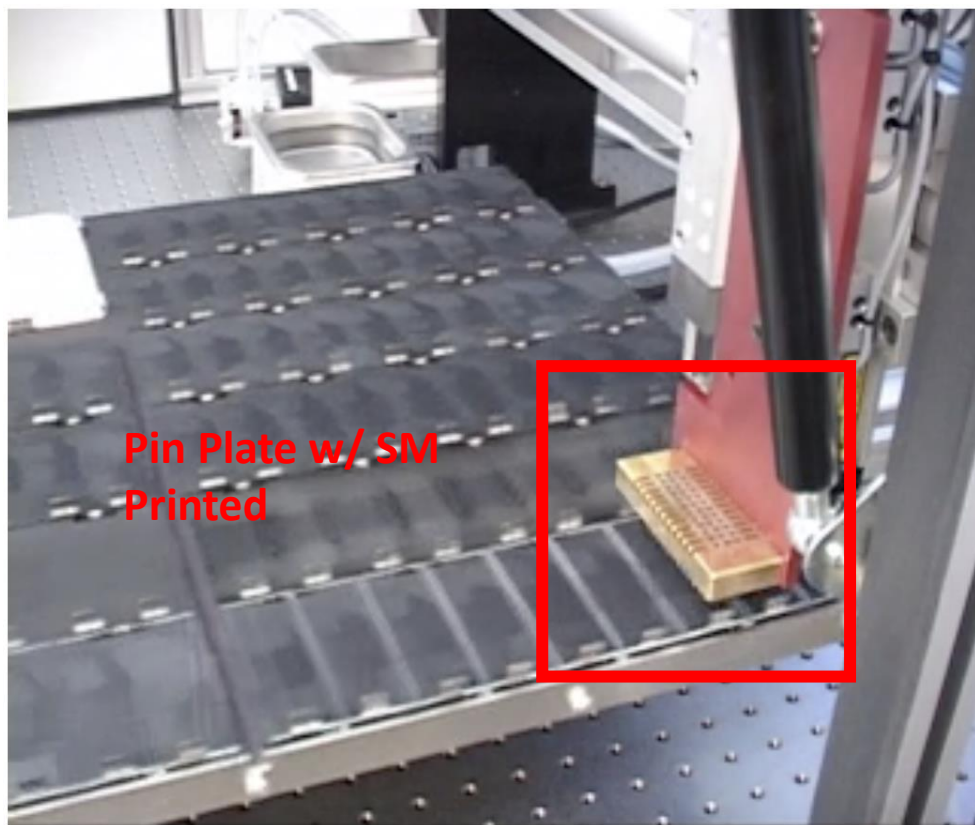
Cons: Potential for off target effects

Can small molecules be useful for understanding “undruggable” targets?

How did previous 20.109 students screen for potential small molecule binders for TDP-43?

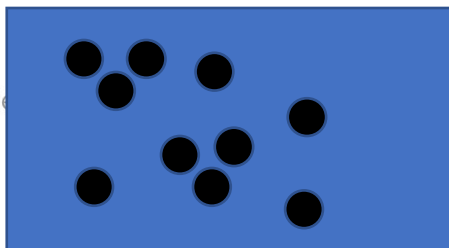
- Used a high throughput assay, the **small molecule microarray (SMM)**
- High throughput assays like the SMM:
 - Allow unbiased exploration of potential therapeutics
 - Allow examination of targets with limited information
 - Allow for the screening of potentially millions of putative binders at a time

Small Molecule Microarray (SMM)

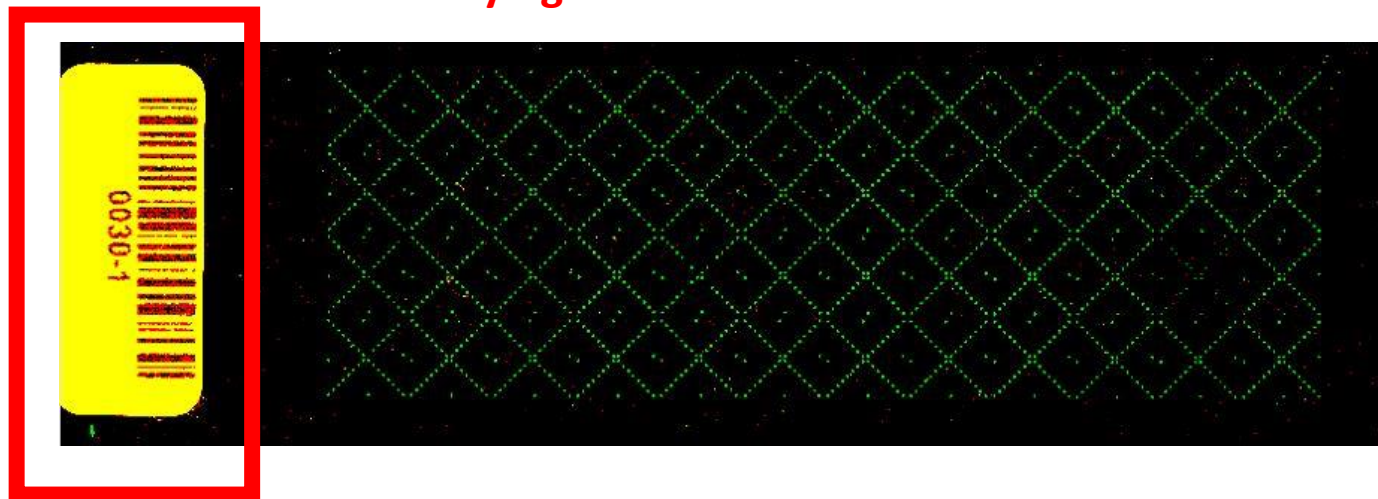


Koehler Lab 2014 - Small-molecule

/imeo.



Identifying Barcode

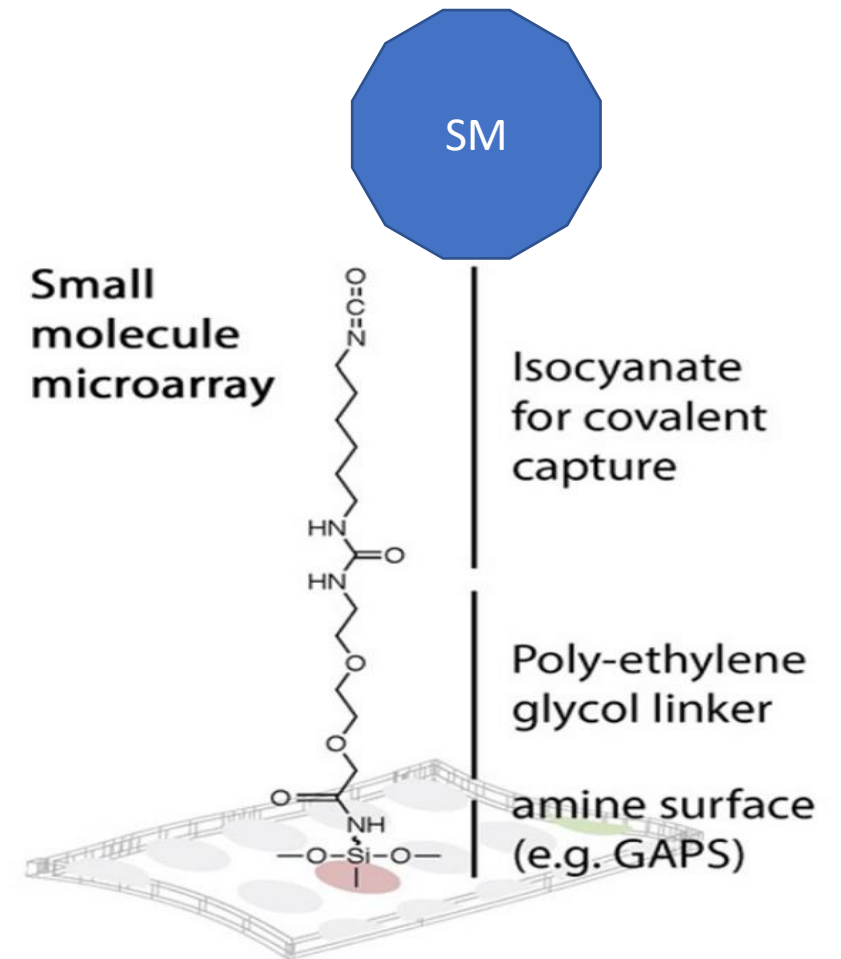


- Each slide contains ~12,000 spots
 - ~4,200 small molecules / ligands (in duplicate = ~8,400)
 - Fluorescein sentinel spots
 - DMSO negative control spots

Why DMSO? – SMs dissolved in DMSO

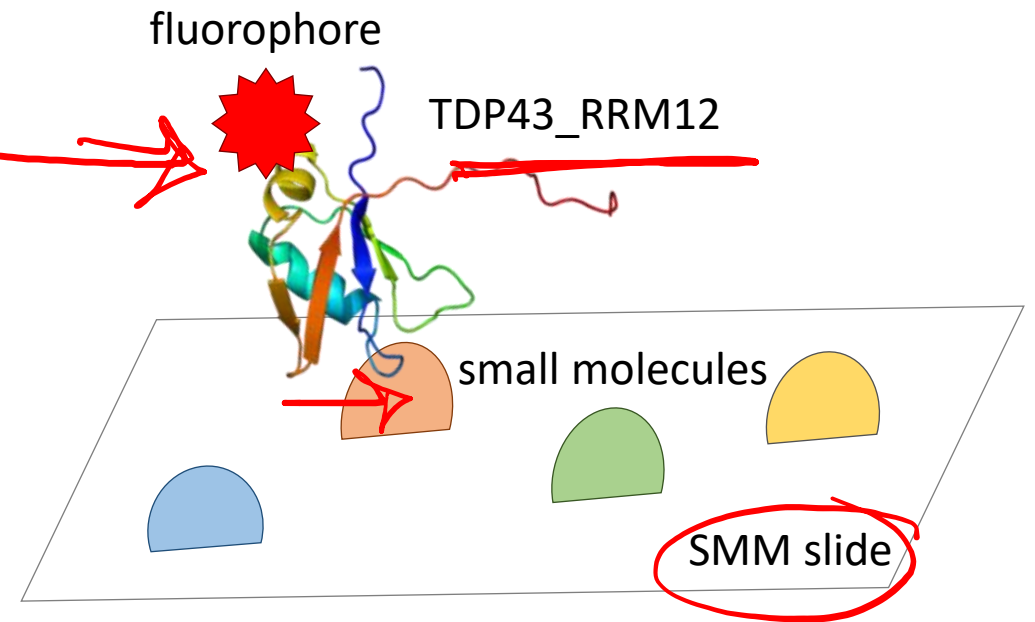
SMM slide preparation

- Gamma-aminopropylsilane (GAPS) coated slide with polyethylene glycol (PEG) spacer
- PEG coupled to 1,6-diisocyanatohexane to generate isocyanate-functionalized slide
- Isocyanate able to react with nucleophilic functional groups (strong, but not specific binding)



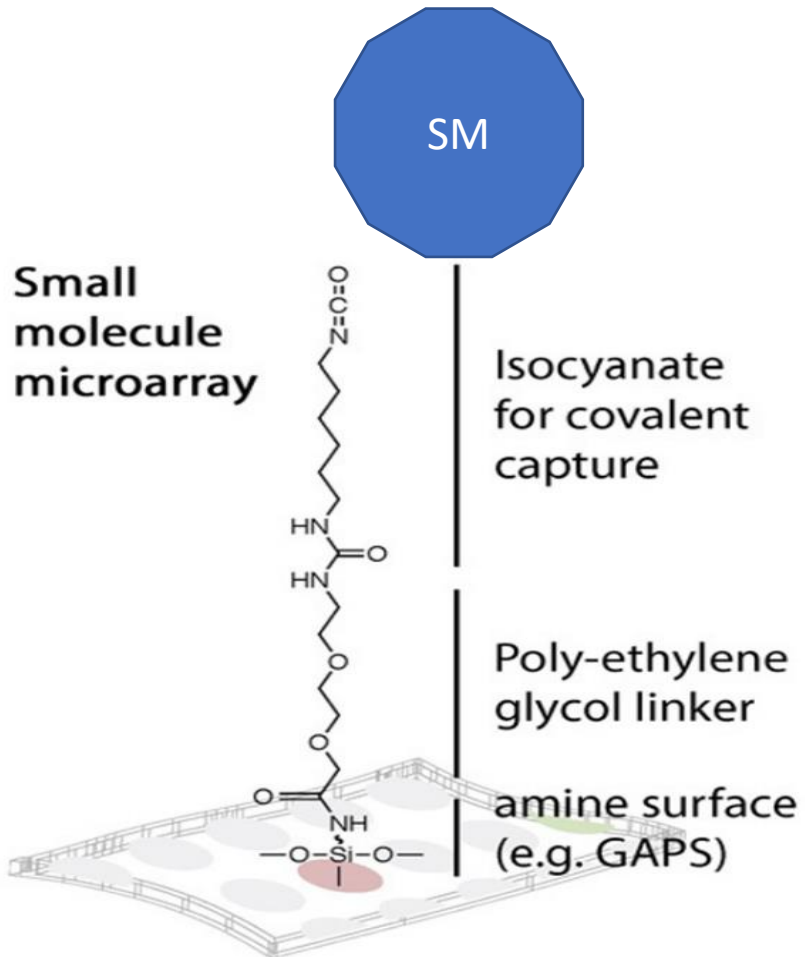
How do we use the SMM to screen for ligands that bind our protein of interest?

- Create a recombinant protein of the TDP43 RNA binding domains (TDP43_RRM12)
 - Label this protein with a Alexa647 fluorophore
- Incubate the SMM slide with our purified and labeled TDP-43_RRM12
- Wash away unbound protein
- Store for scanning

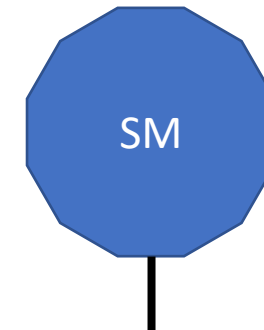


Why might we need a long linker between the SM and the slide?

Elevated SM has more contactable surface area exposed for protein to bind to



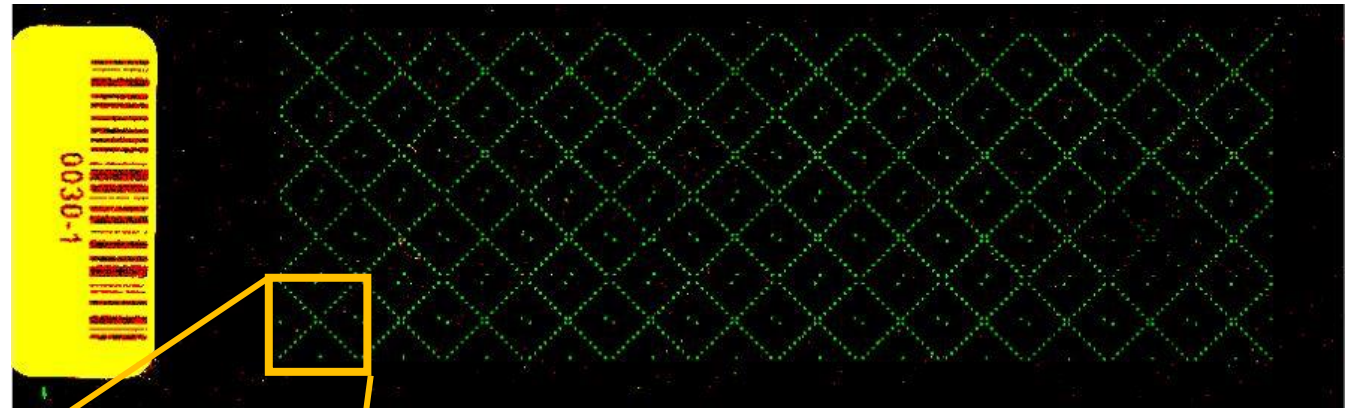
VS



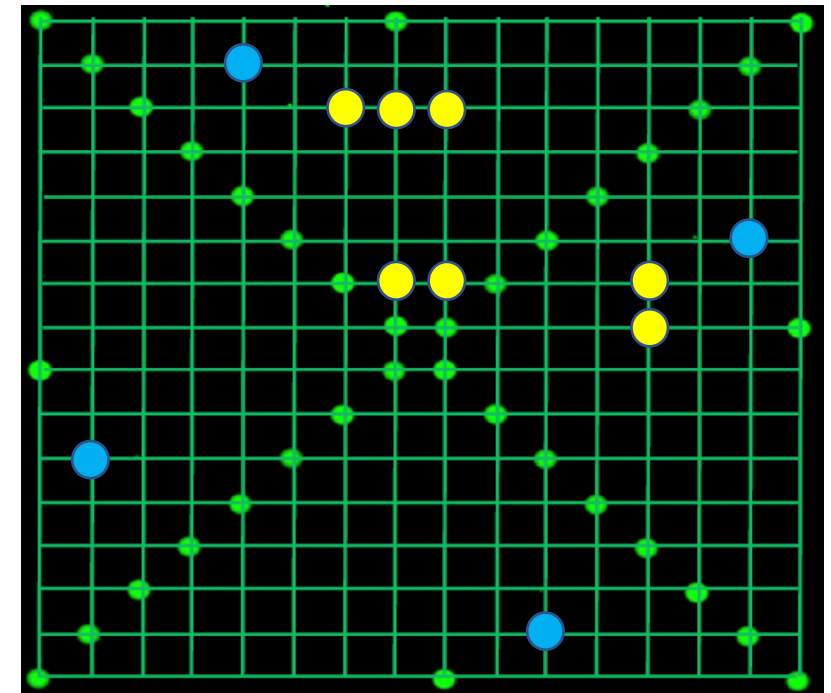
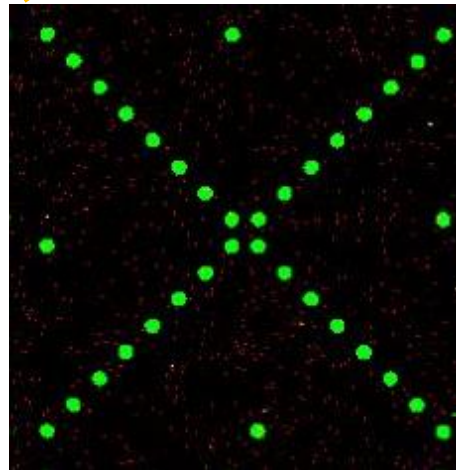
Guide to the SMM slide

- Each slide has several blocks
- Each block has sentinel spots which are landmarks
- Rest of dots are small molecules and controls
- Can overlay a computational map to identify the location of each small molecule

Slide



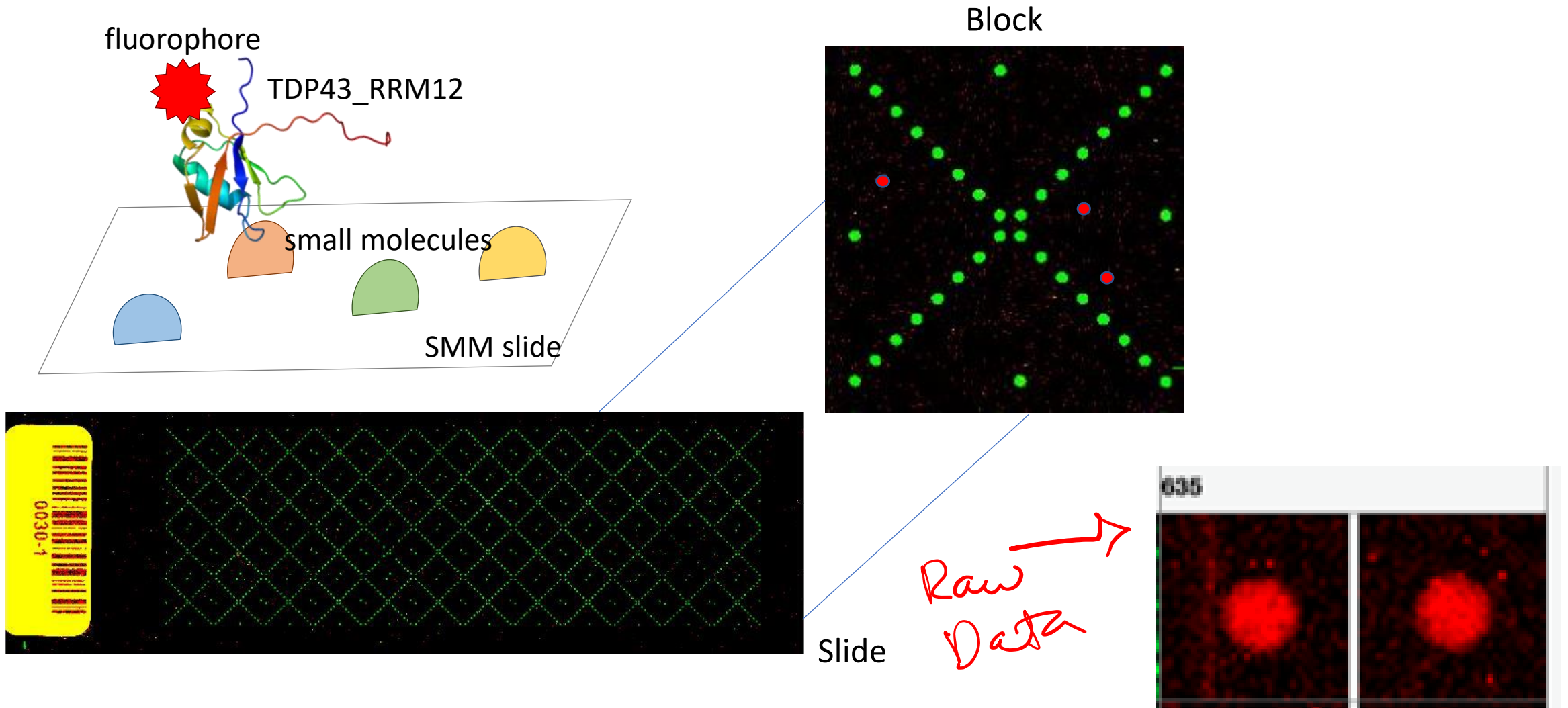
Block



Green= sentinel spots
(fluorescein dye)

Blue= DMSO
Yellow= SM

What do putative binders look like on the SMM slide?



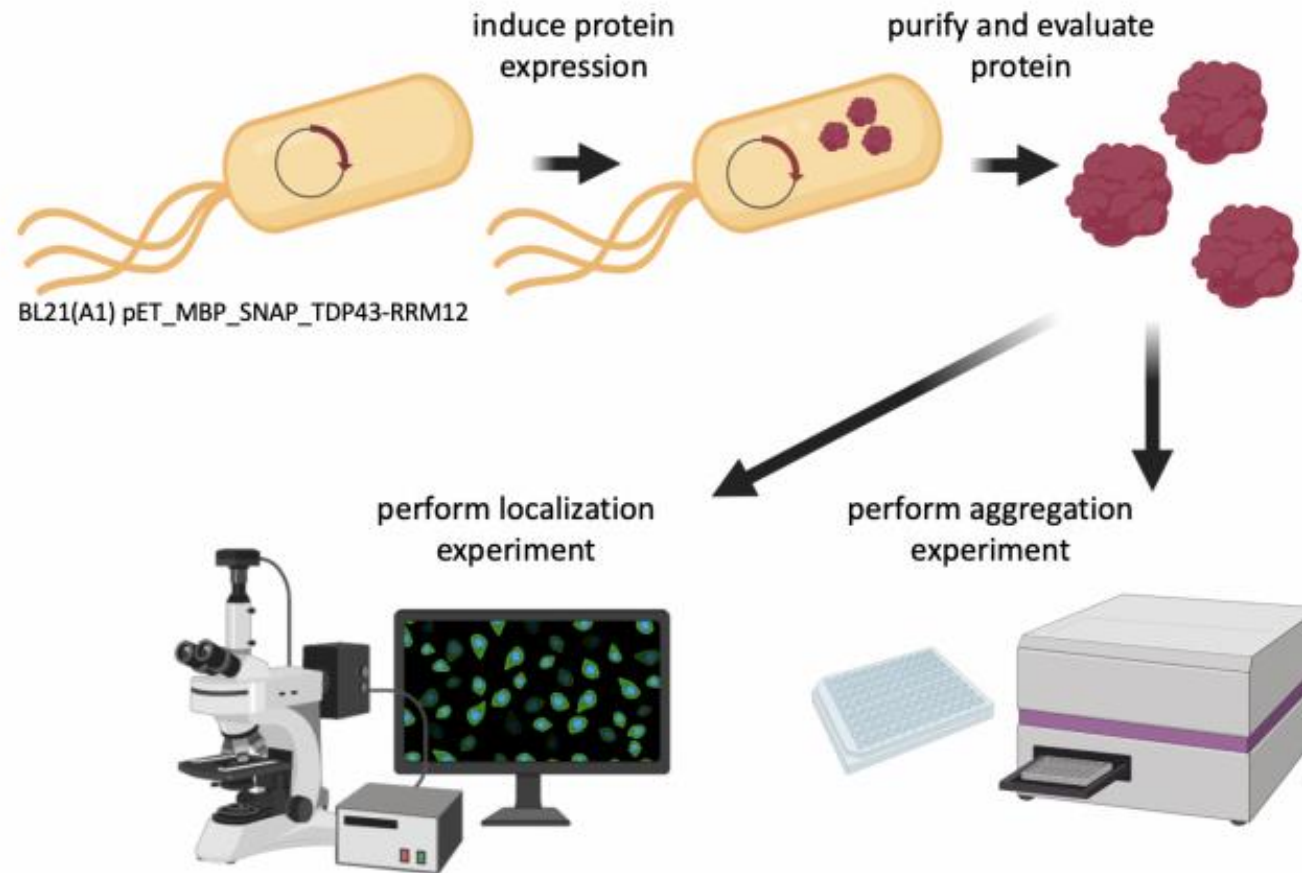
How do you identify small molecules for further study?

- Next class you will learn about the computational workflow used to analyze the SMM data to determine small molecule hits
- A combination of:
 - Identifying potential signal bias inherent to the production of the slides
 - Identifying a threshold for a strong fluorescent signal
 - Visually validating that fluorescent signal conforms to expected shape

Once we have a group of small molecules that are putative binders to the TDP-43 protein, we will perform follow up assays to assess potential biological impact of association

Overview of Mod1 experiments

Research goal: Use functional assays to characterize ligands identified as binders to TDP43 from SMM technology



For today...

- Work through SMM on wiki
- Take notes in your Benchling notebook using the template you created
 - Show today's entry to Tyler before you leave to receive participation points

For M1D2

- Read the article and guidelines linked on the M1D2 wiki page