

# M2D7: Analyze RNA-Seq data

04/07/2017

1. Prelab Discussion
2. Intro to R Exercise
3. Start Mod2 Report early!

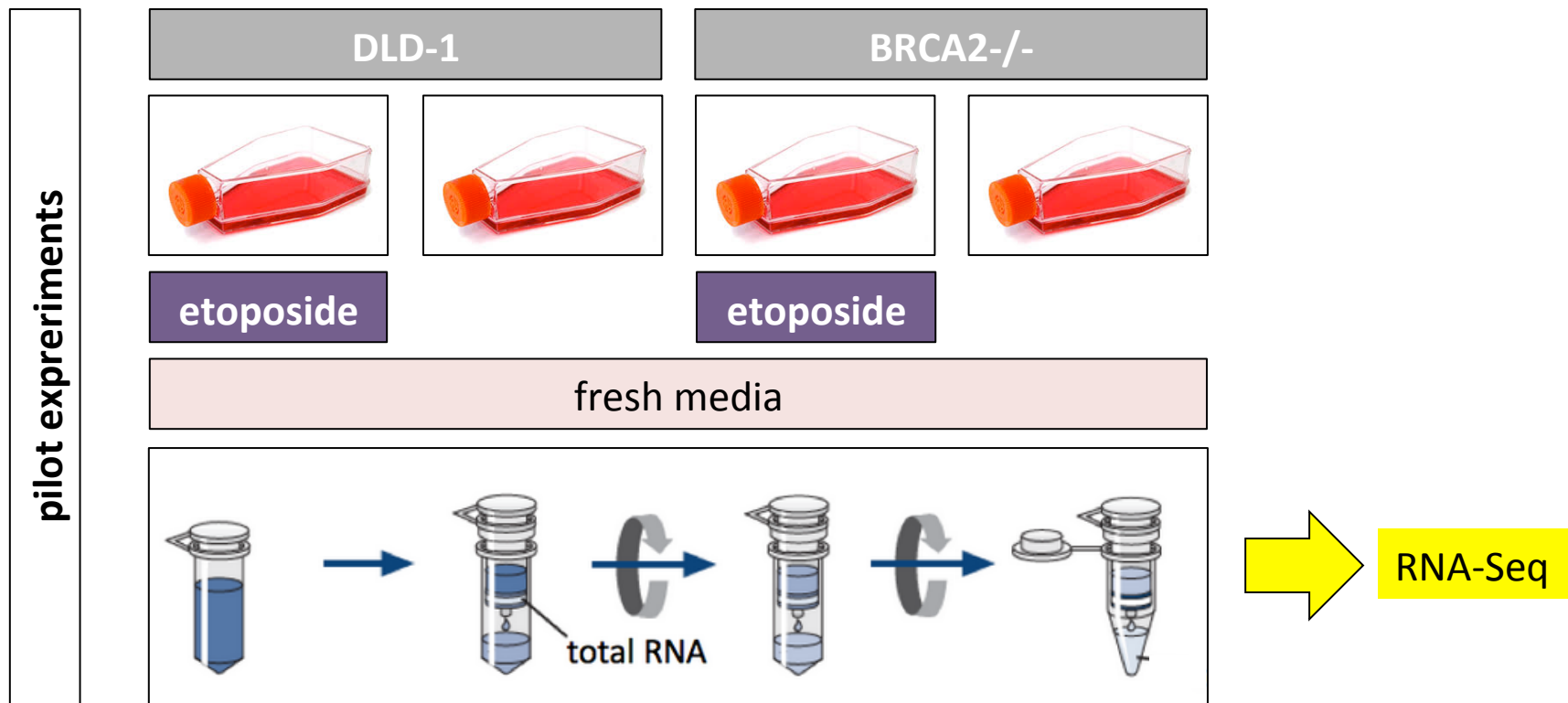




# Let's analyze RNA-Seq data

## DNA damage response

- With qPCR, you looked at p21 transcript levels (norm. to GAPDH)
- With RNA-Seq, we'll peek into the entire transcriptome

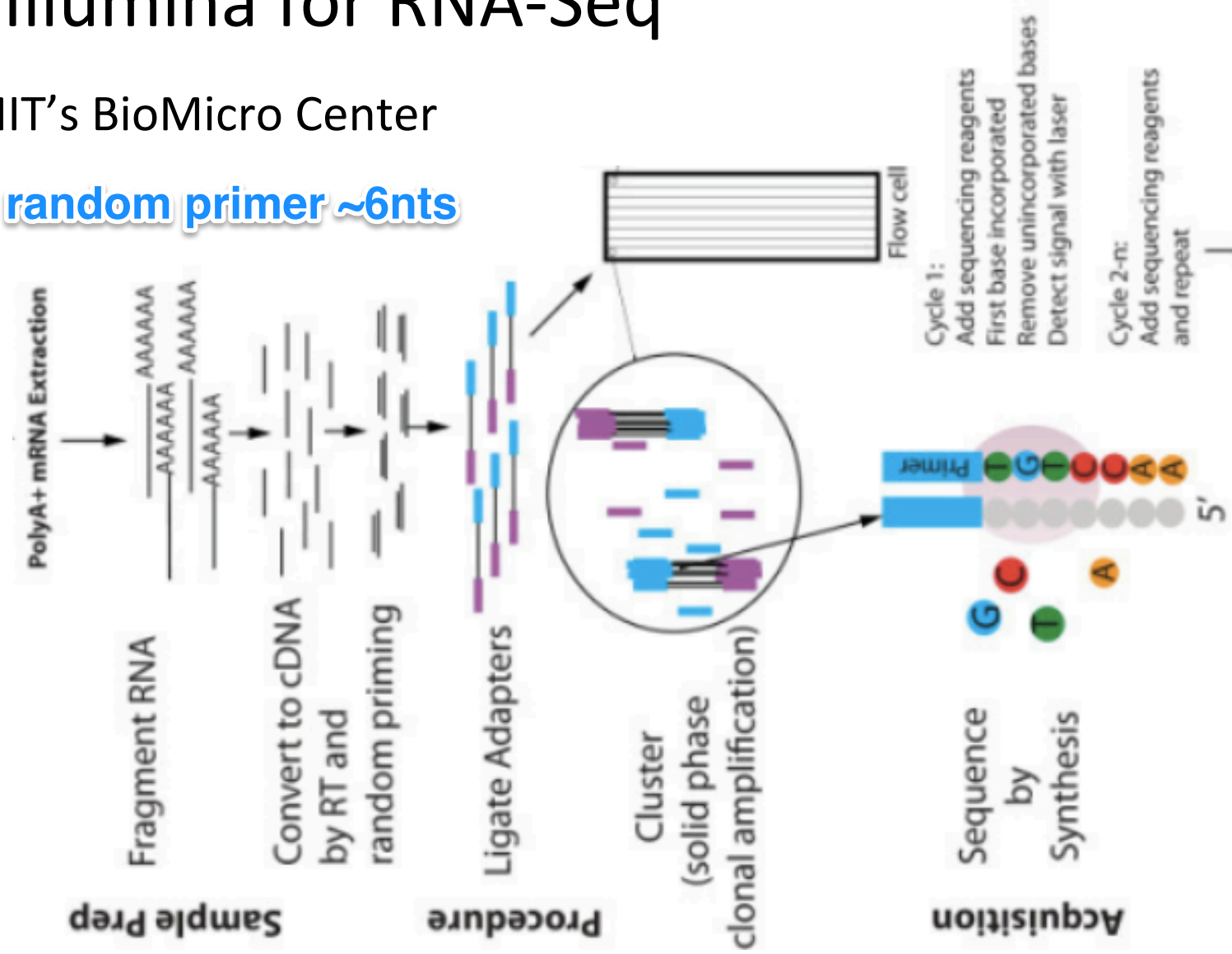


# OVERALL

## Next generation sequencing (NGS) by Illumina for RNA-Seq

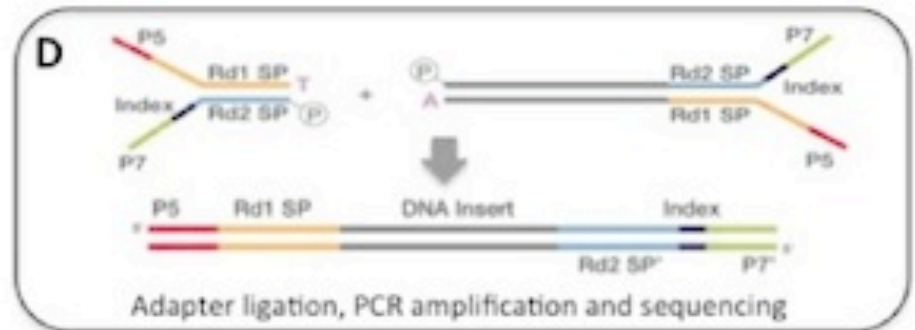
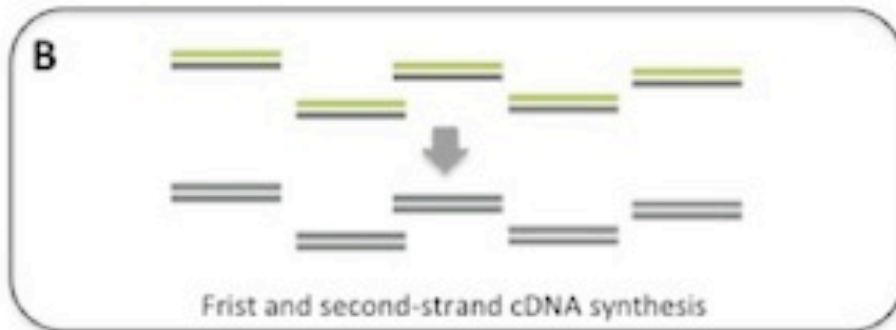
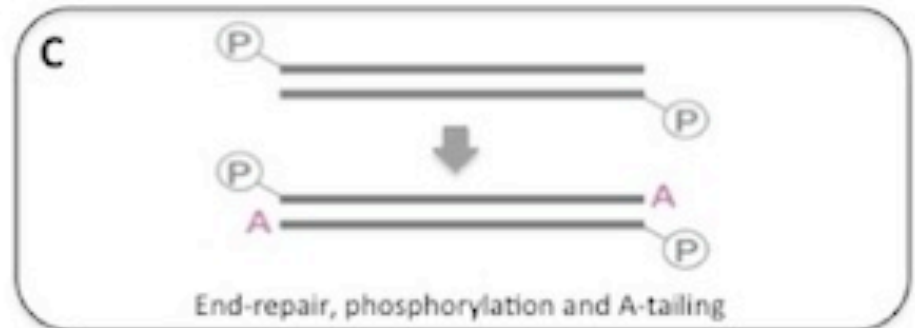
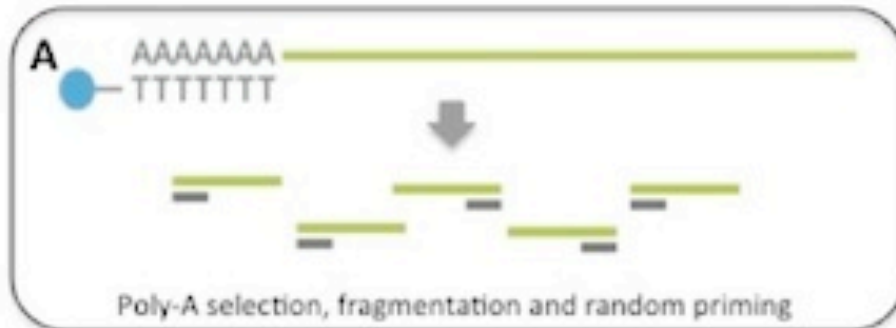
at MIT's BioMicro Center

random primer ~6nts



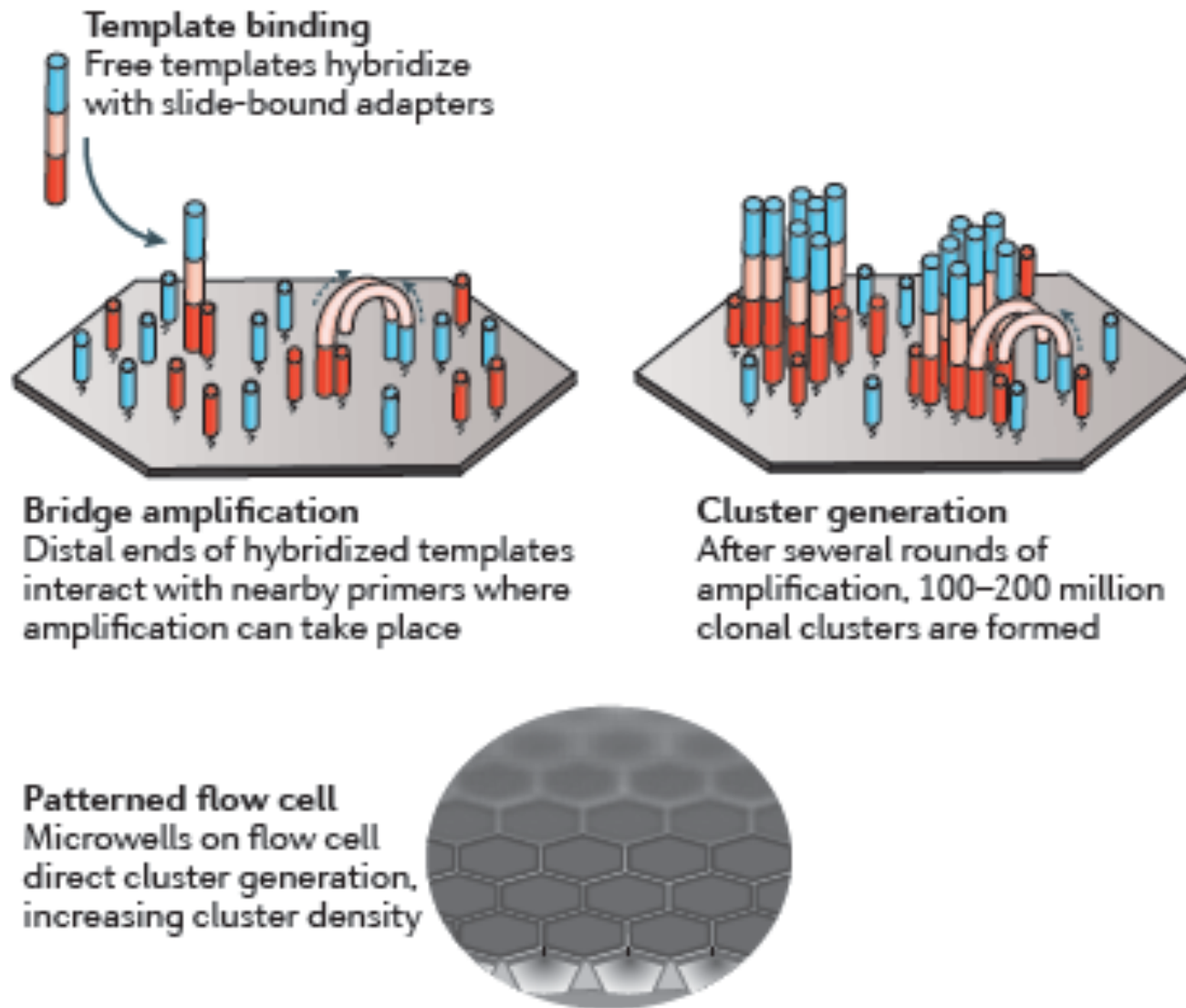
## 1ug RNA for library prep

# Illumina Tru-Seq RNA-seq protocol

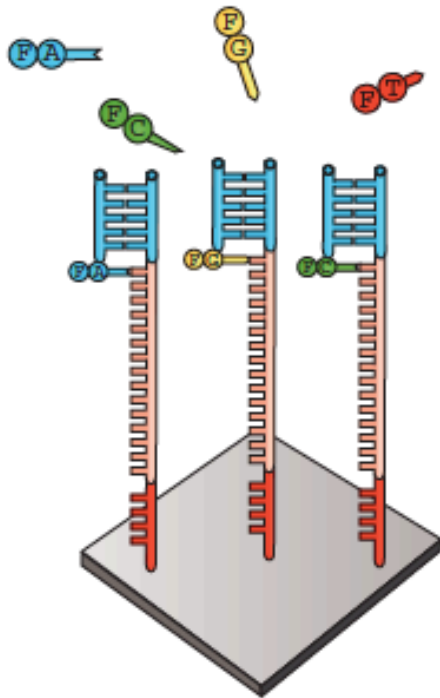
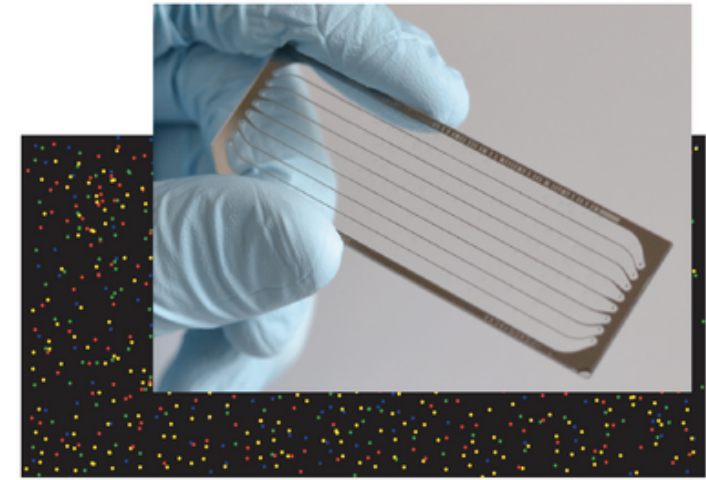


Library prep begins from 100ng-1ug of Total RNA which is poly-A selected (A) with magnetic beads. Double-stranded cDNA (B) is phosphorylated and A-tailed (C) ready for adapter ligation. The library is PCR amplified (D) ready for clustering and sequencing.

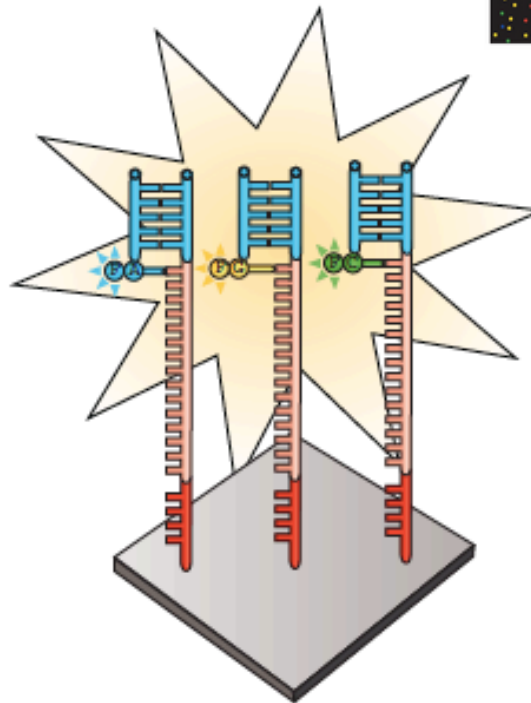
# Solid-phase bridge amplification for clonal amplification of cDNA



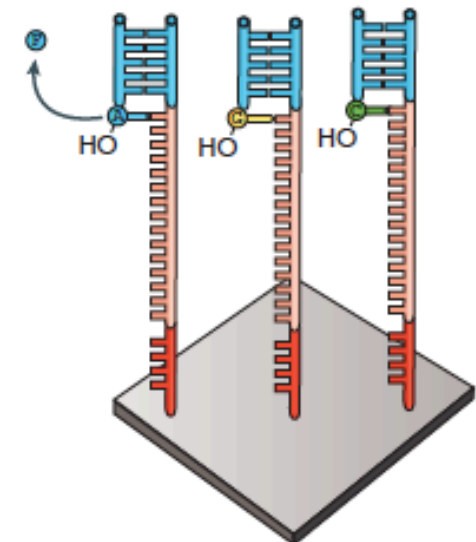
# Sequencing by synthesis (SBS)



**Nucleotide addition**  
Fluorophore-labelled, terminally blocked nucleotides hybridize to complementary base. Each cluster on a slide can incorporate a different base.



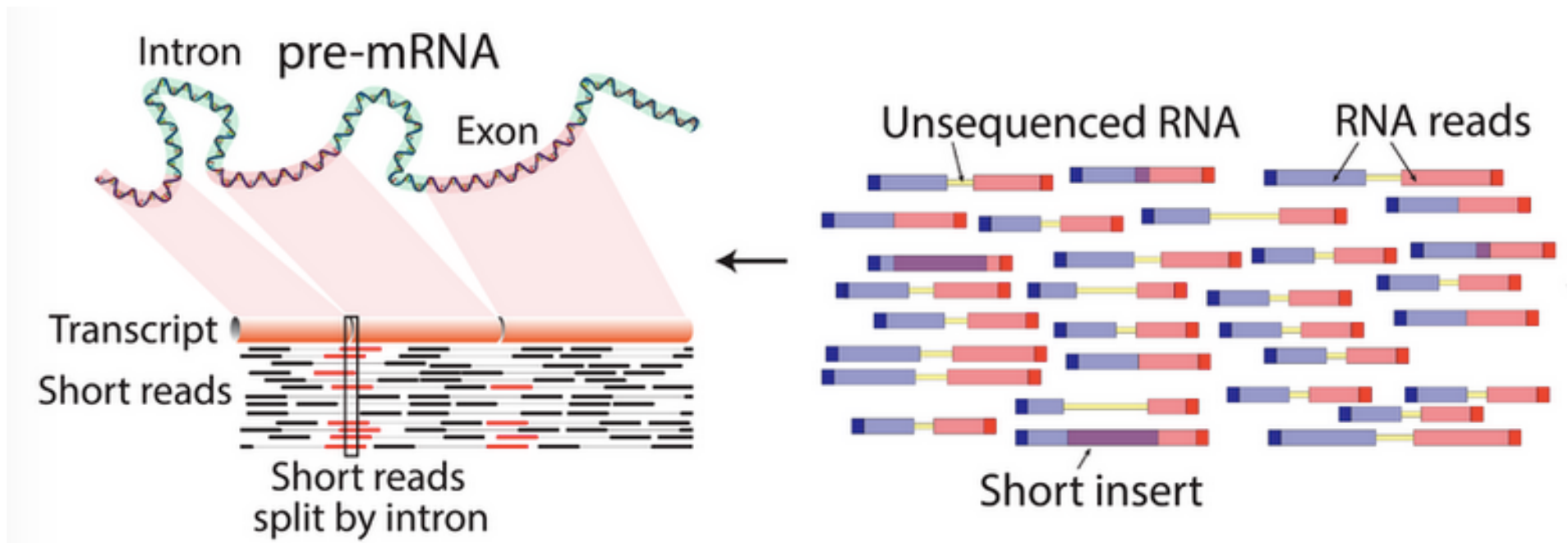
**Imaging**  
Slides are imaged with either two or four laser channels. Each cluster emits a colour corresponding to the base incorporated during this cycle.



**Cleavage**  
Fluorophores are cleaved and washed from flow cells and the 3'-OH group is regenerated. A new cycle begins with the addition of new nucleotides.

# Finally, map to genome, transcriptome, and predict exon junctions

## reference based analysis



# Bioconductor, genomic alignments package

## Data structure

(preprocessed by Amanda)

### experimental conditions



### samples

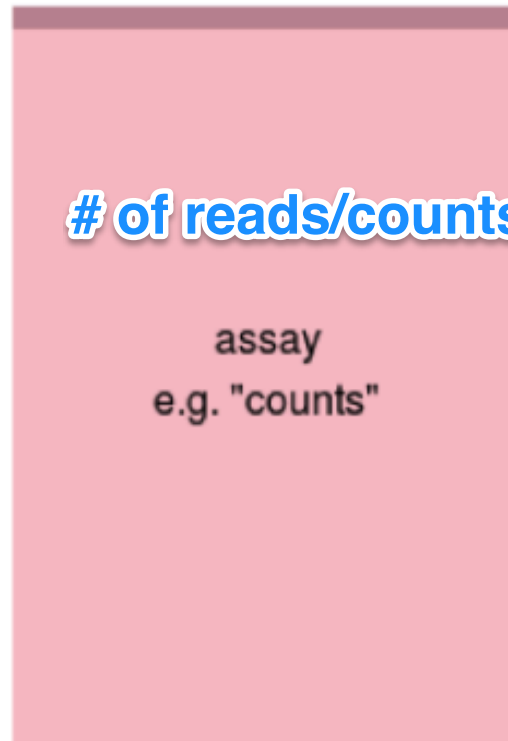
(DLD-1 or BRCA2-/-,  
+/- etoposide)

### genes

(chromosome,  
number of exons,  
ID, etc)



### # of reads/counts



### genomic region



**From Prof. Samson's lecture 03/09/17:**

**What experimental question will you ask in Module 2?**

How does DNA repair affect the ability of cancer chemotherapy drugs to kill cancer cells?

How does cancer chemotherapy affect gene expression?

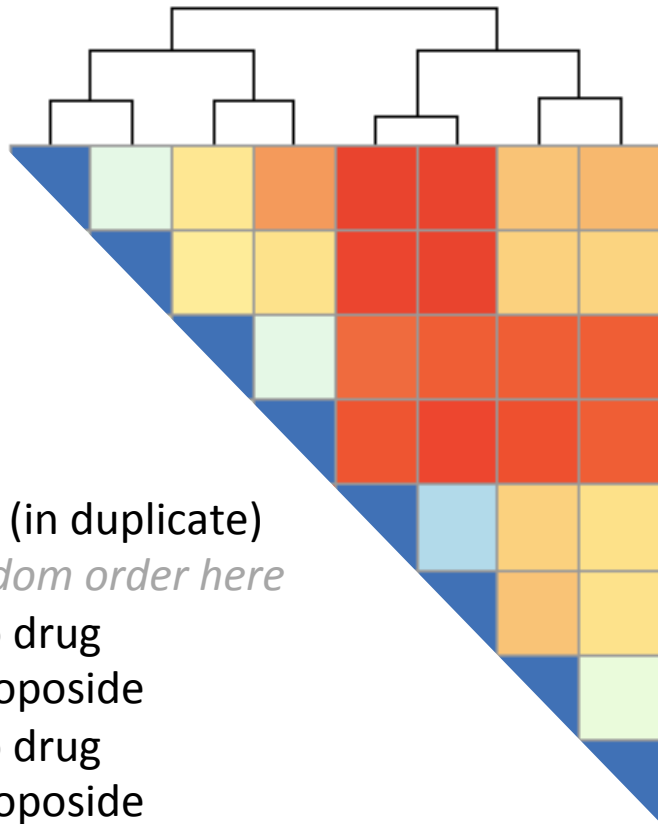
**This raises the following questions**

- How does DNA get damaged?
- What is DNA repair?
- Why does DNA repair exist?

We ask:

Which experimental conditions are **overall** most similar (least similar)? **Clustering:**

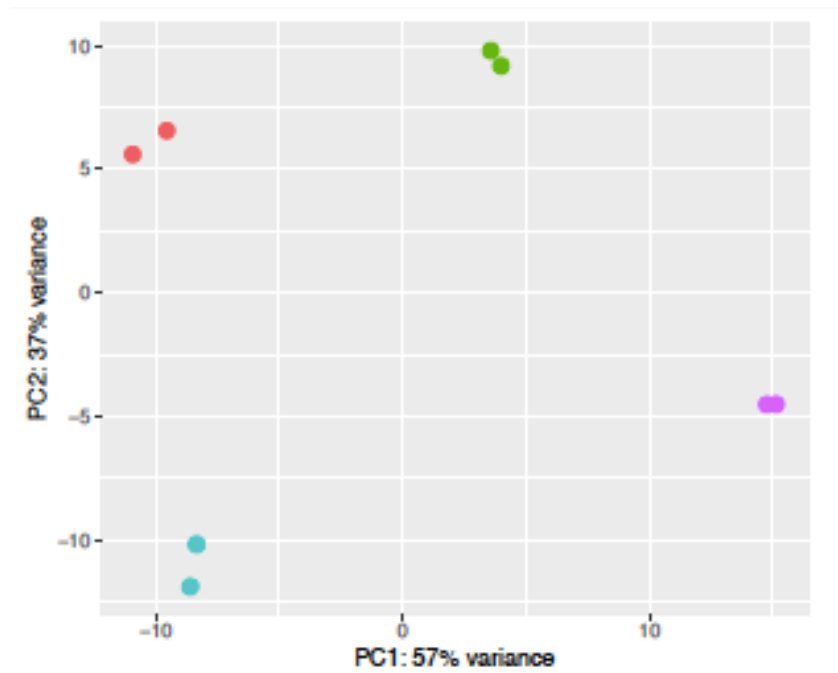
Hierarchical clustering



columns are (in duplicate)  
*listed in random order here*

DLD-1 no drug  
DLD-1 etoposide  
BRCA2-/- no drug  
BRCA2-/- etoposide

Principal component analysis (PCA)

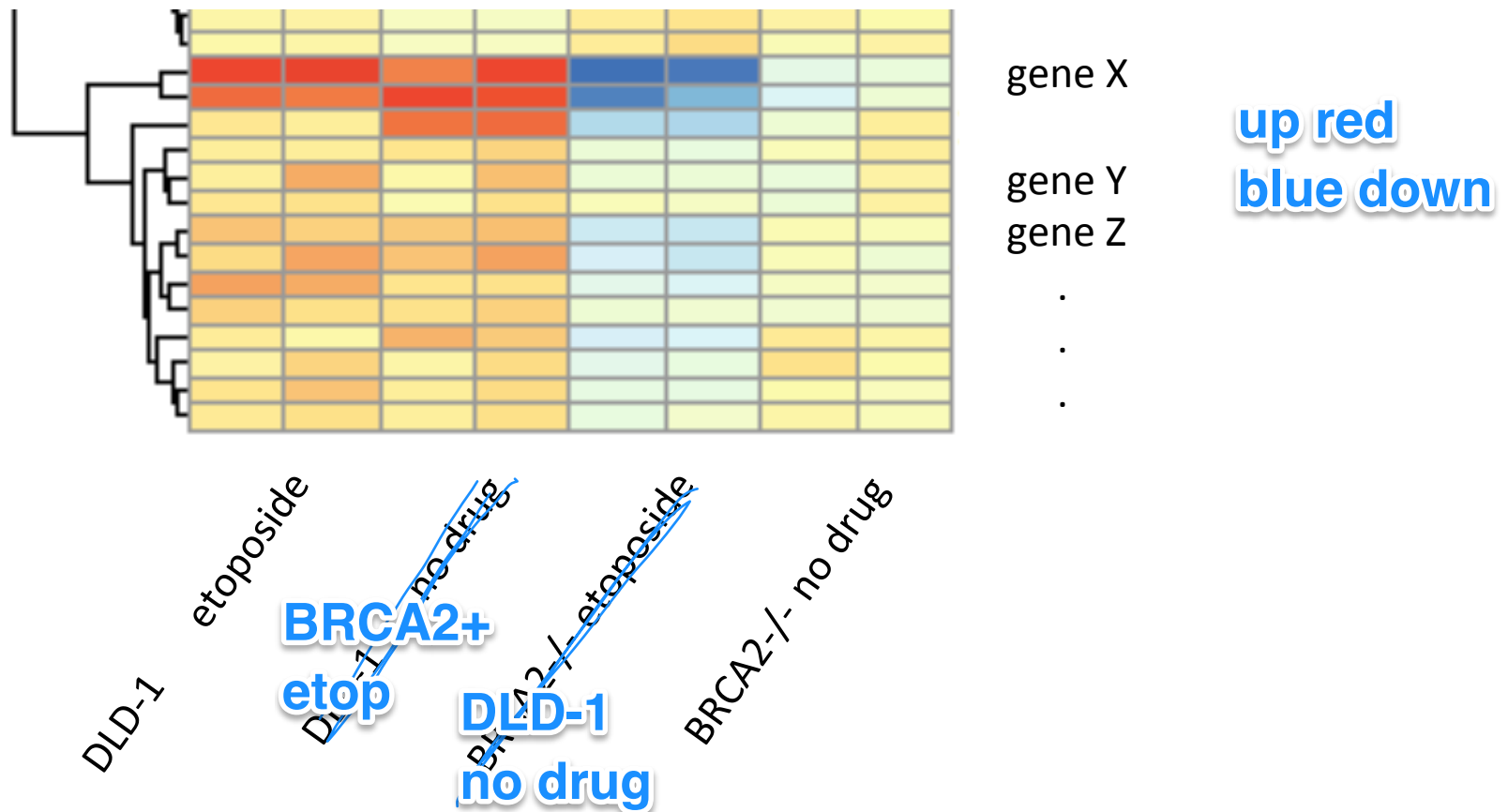


## DNA damage genes

We ask:

Are specific genes differentially expressed?

up or down (or unchanged) compared to average of entire data set



command + return saves so much time!

```
p53_targets = c('CDKN1A', 'BTG2', 'FBXW7', 'GADD45A', 'SFN', 'GTSE1', 'ZNF385A',  
               'PCBP4', 'GPX1', 'GPX2', 'SESN2', 'ALDH4A1', 'SOD2', 'CFLAR',  
               'PTGS2', 'CCNG1', 'DDR1', 'HBEGF', 'PPM1D', 'MYO6', 'TNFRSF10D',  
               'TNFRSF10B', 'APAF1', 'BAX', 'FAS', 'PMAIP1', 'PERP', 'TP53AIP1',  
               'TP53I3', 'BBC3', 'SIVA1', 'PTP4A3', 'PML', 'PTPRVP', 'PIDD1',  
               'DDB2', 'ERCC5', 'FANCC', 'XRCC5', 'MGMT', 'MLH1', 'MSH2',  
               'RRM2B', 'POLK', 'XPC')  
  
mat = assay(rld)[p53_targets, ]  
mat = mat - rowMeans(mat)  
pheatmap(mat, annotation_col=df)
```

However, note the **erratum**: On page 4,

```
pheatmap (sampleDists, labels_row=filenames)
```

should be replaced by

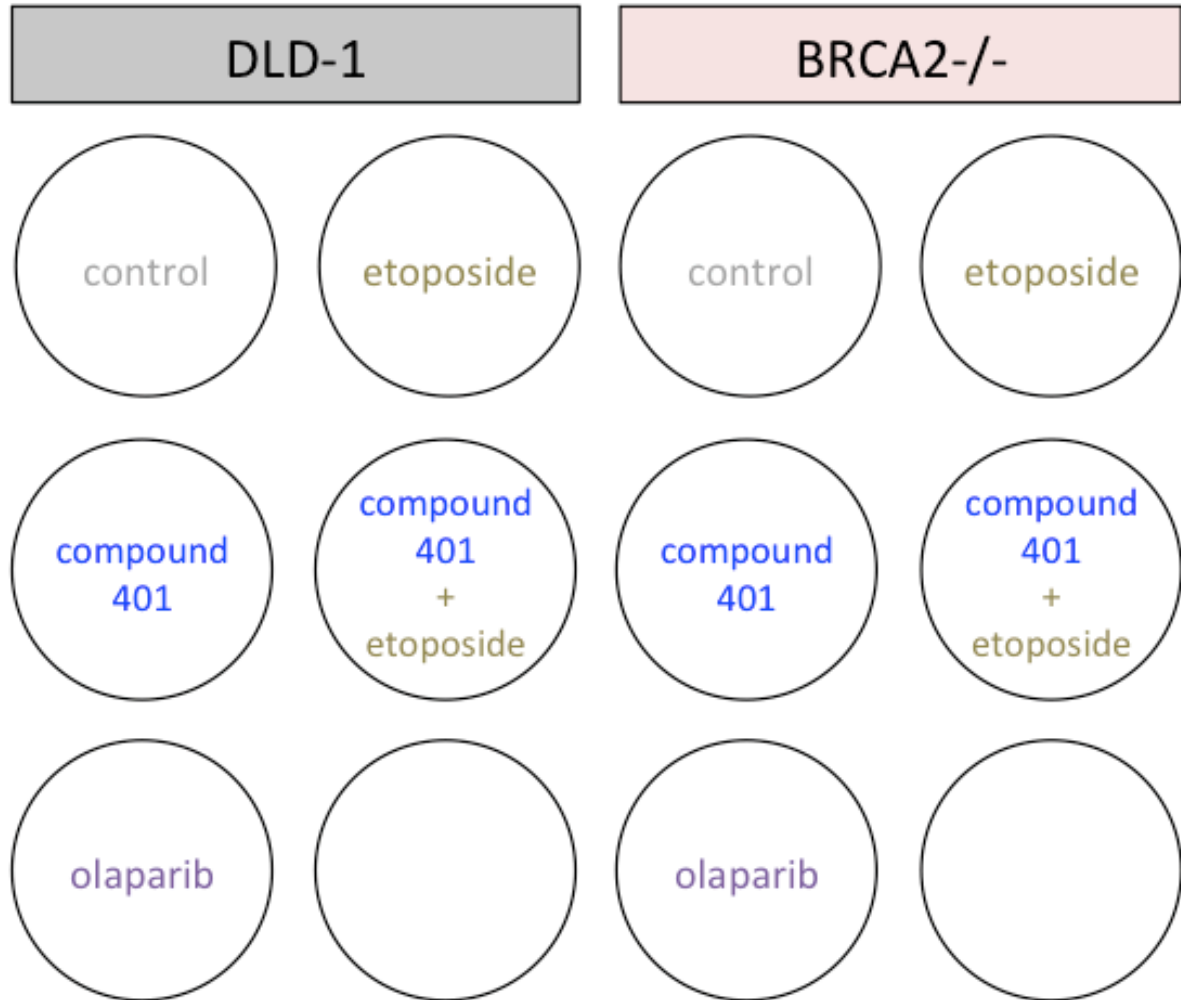
```
pheatmap(sampleDists, labels_row=rownames(colData(dds)))
```

on M2D3: you treated cells to examine viability

- etoposide:  
creates double-stranded  
breaks

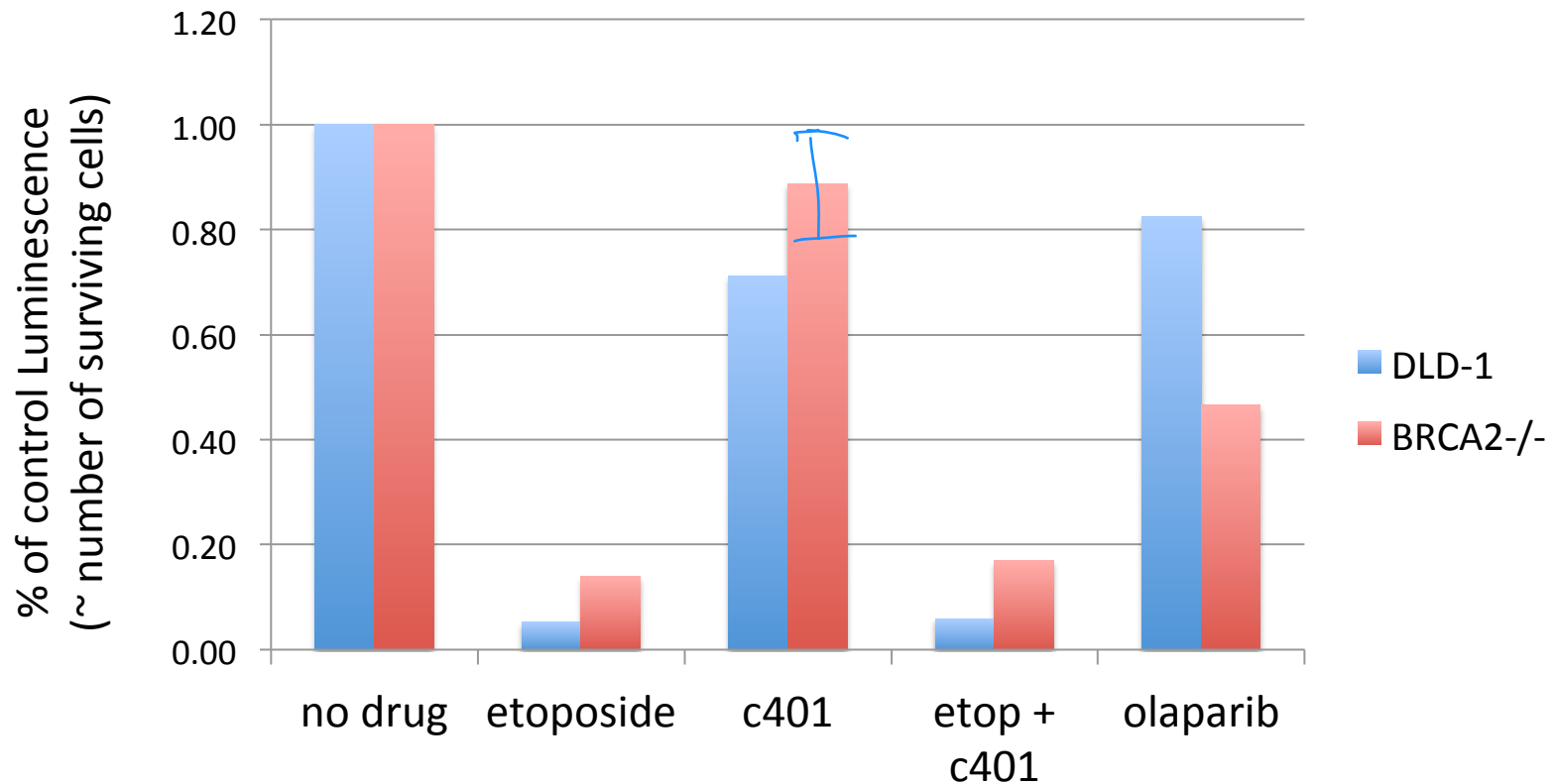
- compound 401:  
inhibitor of DNA-PK  
NHEJ

- olaparib:  
inhibitor of PARP  
BER



If you're done early,  
analyze your CellTiter Glo cell viability data

[http://engineerbiology.org/wiki/Talk:20.109\(S17\):Module\\_2](http://engineerbiology.org/wiki/Talk:20.109(S17):Module_2)



## Today in lab:

1. Get lab computer and analyze RNA seq data set
  2. Make sure to take notes and save images where noted.
  3. You should have enough time in lab today to analyze Cell titer glo data and start your Mod2 report figures!
- No homework due Wednesday! Journal Club 2
  - Don't forget to Journal Club blog (April 15)
  - HW for M2D9 (Friday) could take time, start now!