

M2D1: Tissue culture and confirm cell lines

03/09/2017

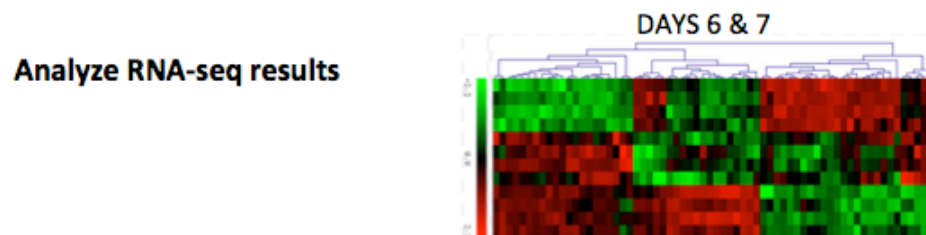
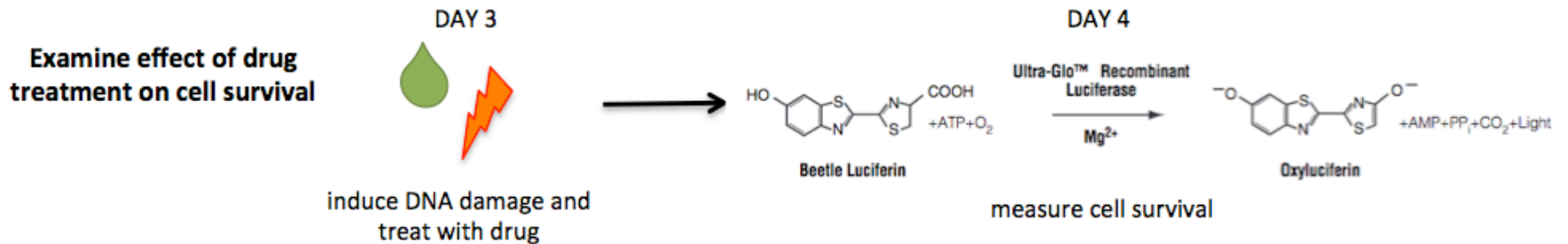
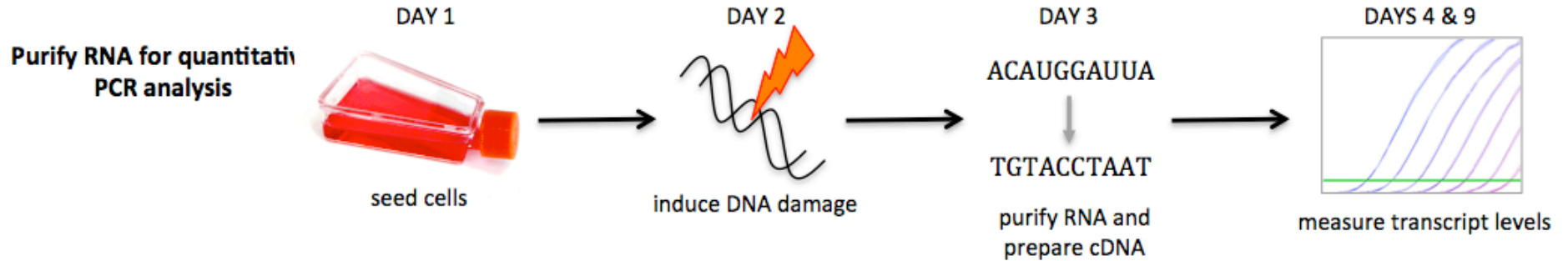
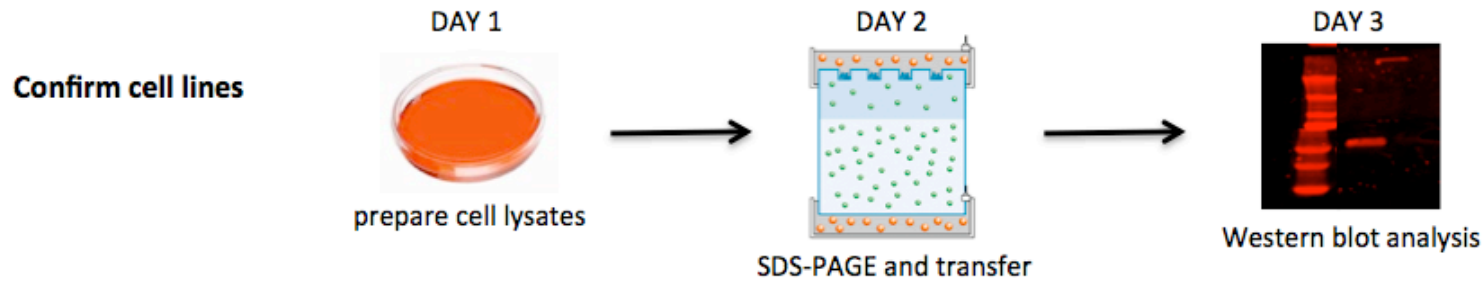
Homework due M2D2: Sign up for journal club

- Pick 1 of 22 papers, or suggest your own
- Present M2D5 (March 23) or M2D8 (April 11)
- Sign up by adding your name next to paper [MJ/WF/Rainbow] [JC/TR/blue]
 - first come first serve!
 - you cannot switch paper after M2D2
 - only one T/R and one W/F per article

M2D8

Slot	Day 5 (T/R)	Day 8 (T/R)	Day 5 (W/F)	Day 8 (W/F)
1	Micayla Flores			
2				
3				
4				
5				
6				
7				

M2: Experimental overview



In lab today

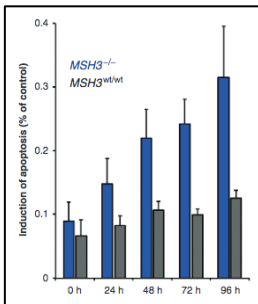


1. Learn/practice cell culture



2. Verify cell lines by Western blot protein analysis

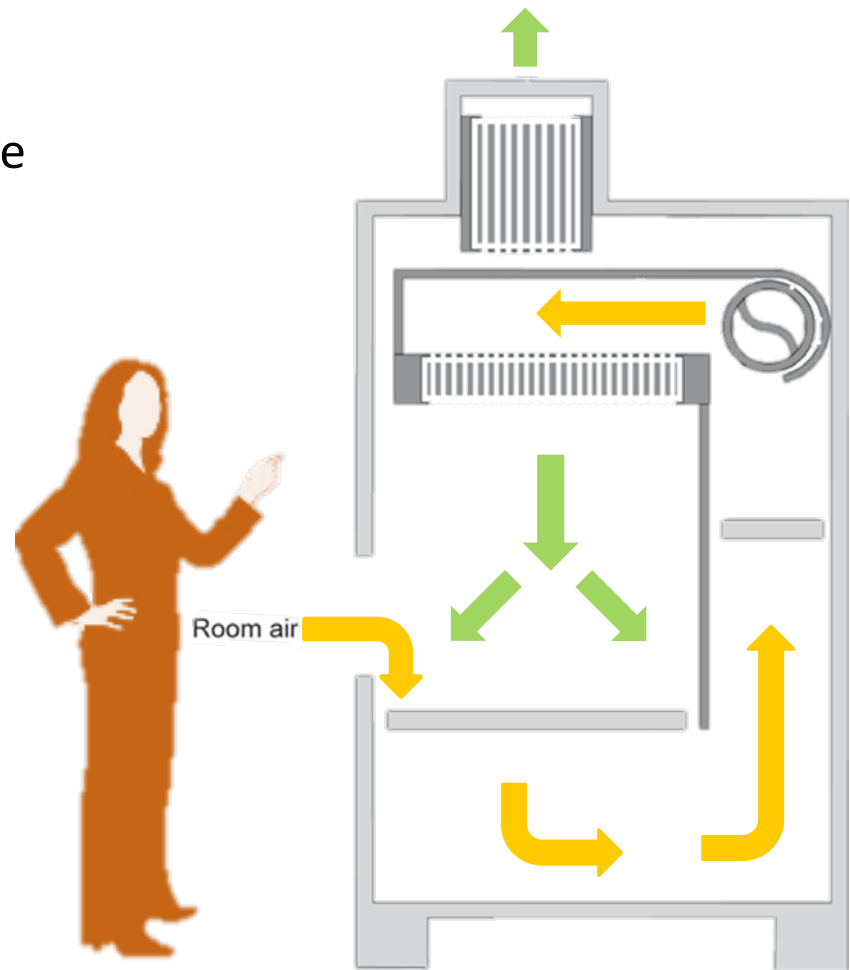
- Lyse DLD-1 and BRCA2^{-/-} cells
- Measure protein concentration
- Separate proteins by SDS-PAGE
- Transfer proteins onto nitrocellulose membrane



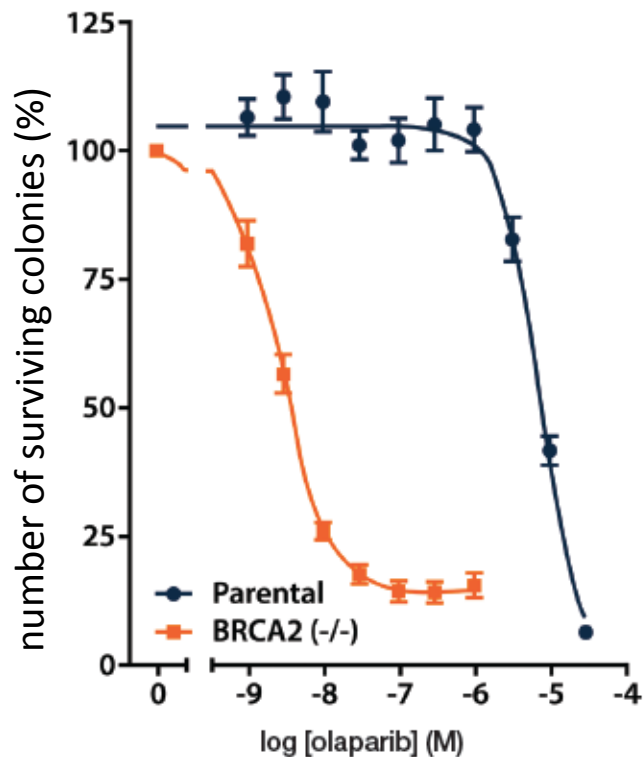
3. Paper discussion

Tissue culture sterile technique

- **70% ethanol** is your BFF:
 - wipe cabinet before and after use
 - wipe everything that enters the cabinet
- Do not disturb air flow:
 - Do not block grille or slots
 - Minimize side-to-side arm movements
 - Work > 6" away from sash
 - Leave blower *on*
- Do not talk into incubator!
- Only open in hood



Our cell lines: DLD-1 and BRCA2-/-



- DLD-1 = wild-type
 - from the colon of a male with colorectal adenocarcinoma
- BRCA2-/- = mutant
 - disruption of exon 11 from BRCA2 gene
 - deficient in DNA repair (by homologous recombination) **HR**

Note: olaparib is a PARP inhibitor

Mammalian cell culture medium



Food:

- RPMI 1640 (Roswell Park Memorial Institute)
 - (a lot of phosphate)
 - often used to culture lymphoid cells

glucose, amino acids, salts, vitamins
phenol red: pH indicator



- FBS: fetal bovine serum 10%
 - BSA and other proteins
 - growth factors, cytokines, lipids, cholesterol
 - "undefined"

Non-food:

- antibiotics:
 - penicillin 1%
 - streptomycin



Mammalian cell culture terminology

- confluence

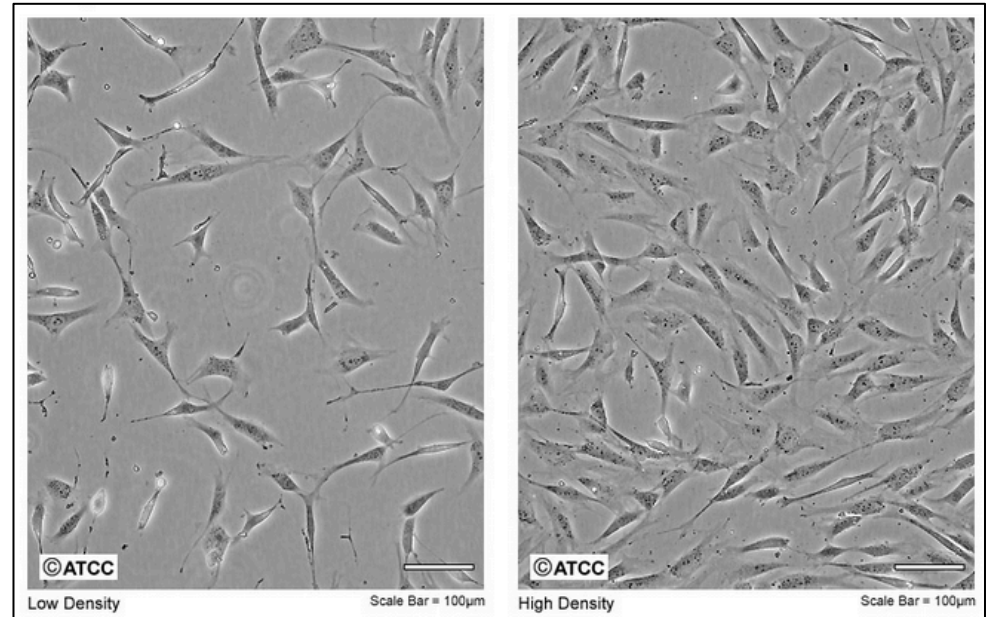
80% when split

- splitting

"sub-culturing"

- seeding

~ 20-40% of confluent culture



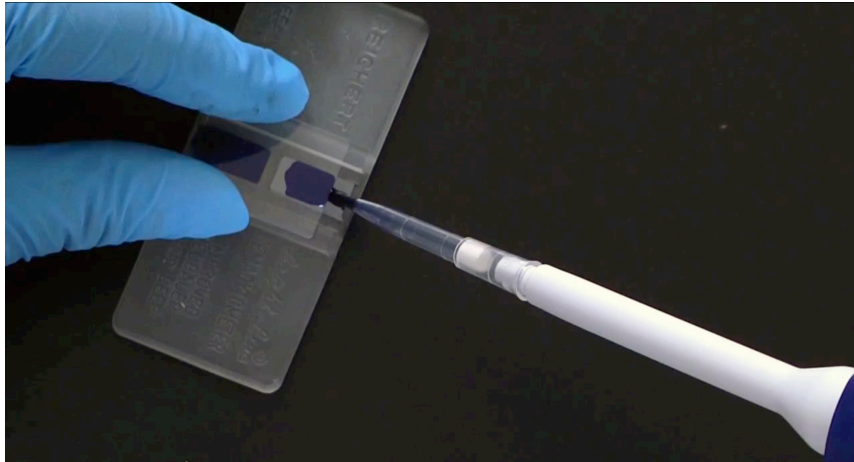
Seeding your DLD-1 and BRCA2-/- cells



why?

1. Rinse with PBS
 - get rid of debris, dead cells
 - eliminate some anti-trypsin agents (released)
2. Detach with trypsin
 - serine protease
 - breaks surface-cells adhesions
3. Calculate number of cells
 - consistency, reproducibility
4. Seed new culture dish

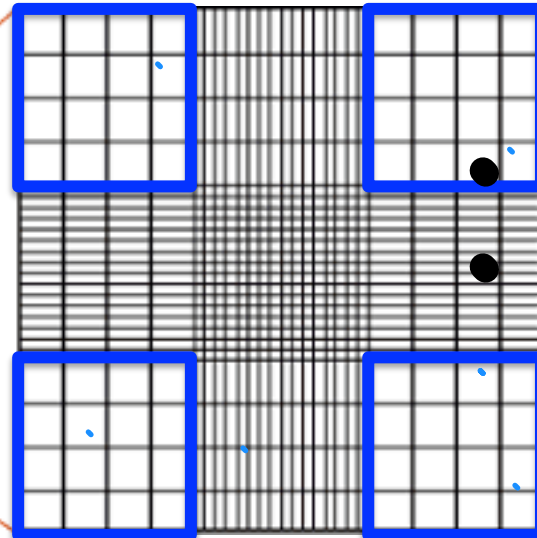
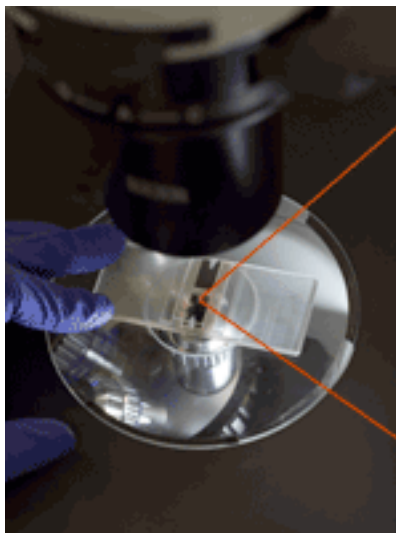
Calculating number of cells



- Hemacytometer
- Trypan blue

dye stains dead cells

- # cells / mL = 10,000 x average of 4 corners



30,000 cells/mL

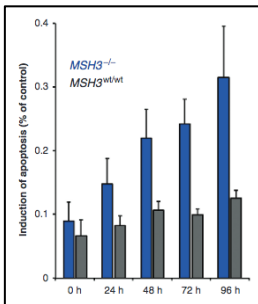
In lab today



1. Learn/practice cell culture



2. Verify cell lines by **Western blot** protein analysis
“immunofluorescence on PAGE-separated proteins”



3. Paper discussion

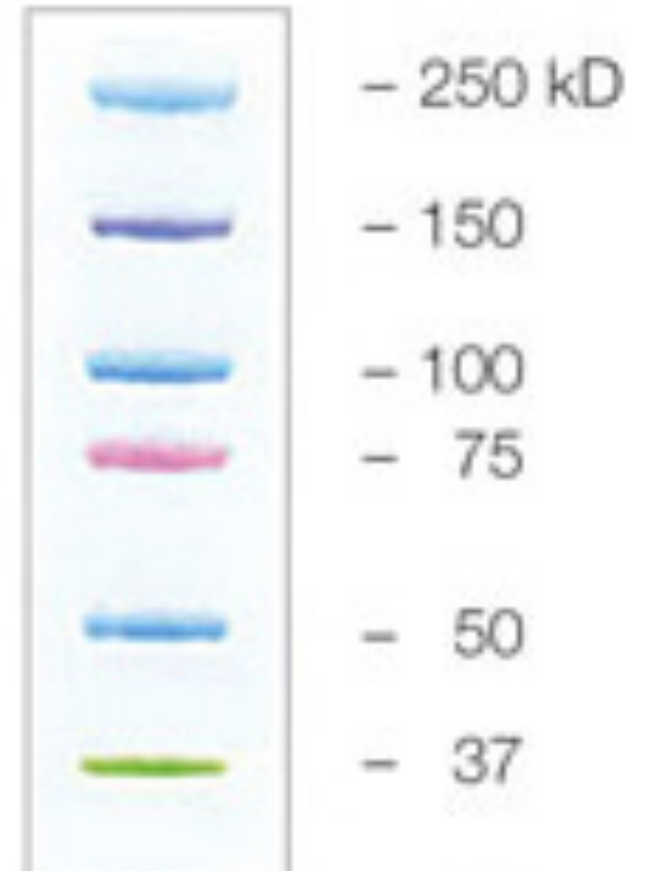
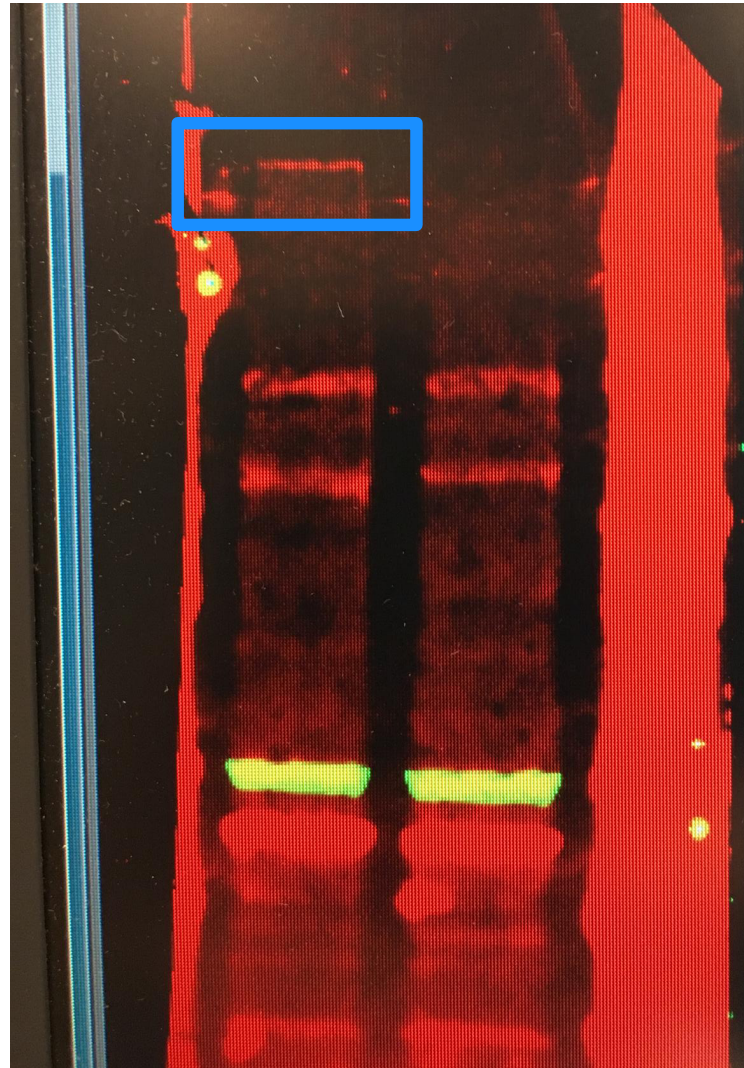
Verify BRCA2^{-/-} are missing BRCA2 (380 kDa)

wt DLD-1 mt BRCA2^{-/-}

BRCA2 →

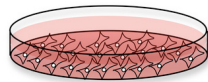


tubulin
(~50 kDa)
control →

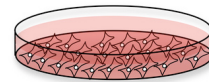


Verify cell lines by Western blot protein analysis

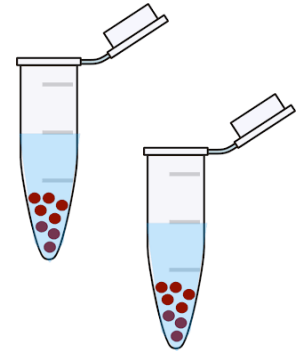
– Lyse DLD-1 and BRCA2 cells



“normal cells”
DLD-1



“DNA repair-deficient cells”
BRCA2-/-



RIPA buffer for mammalian cell lysis:

- **detergents;**
 - 1% NP-40 (nonyl phenoxyethoxyethanol)
 - 0.1% SDS (sodium dodecyl sulfate)
 - 0.5% sodium deoxycholate
- protease inhibitors **prevent protein degradation**
keep proteins intact
- Tris-HCl, pH 7.4 + NaCl
physiological pH and ionic strength

In lab today



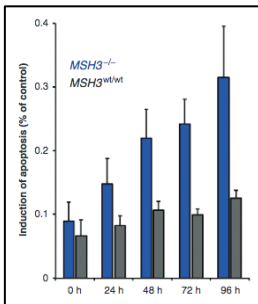
1. Learn/practice cell culture

- red, yellow, purple and silver teams



2. Verify cell lines by Western blot protein analysis

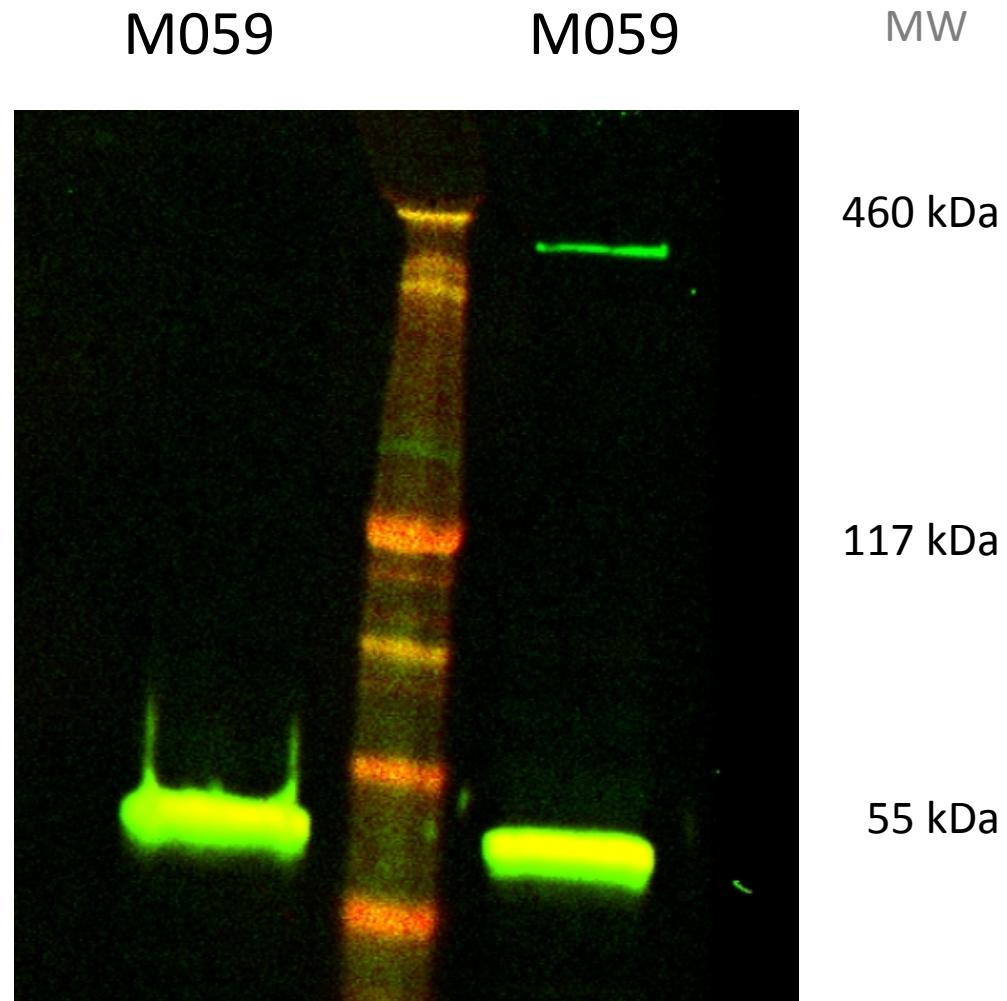
- green, blue, and pink teams



3. Paper discussion

- 4pm?

2. Verify BRCA2^{-/-} are missing BRCA2 by *LI-COR* Western blot

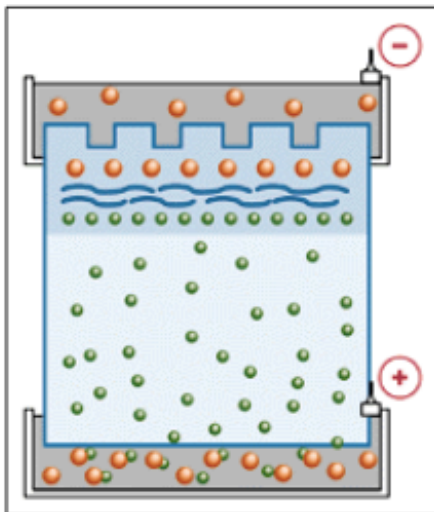
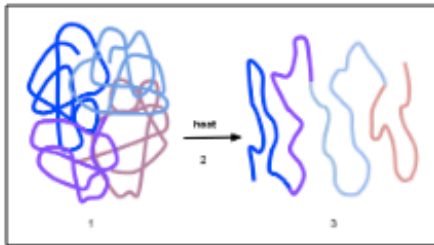
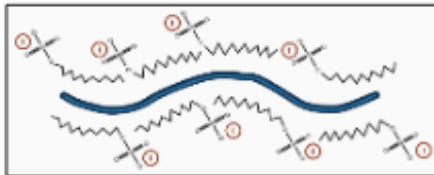


1. Verify cell lines by Western blot protein analysis

SDS-PAGE separates proteins by size

sodium dodecyl sulfate – polyacrylamide gel electrophoresis

carcinogenic



- Laemmli sample buffer / loading dye:

- + SDS: **detergent denatures proteins, coats proteins with negative charges**


- + β -mercaptoethanol **reduces disulfide bonds**

- + bromophenol blue **to follow front of migration**

- + **glycerol**

- boiling denatures higher-order structures

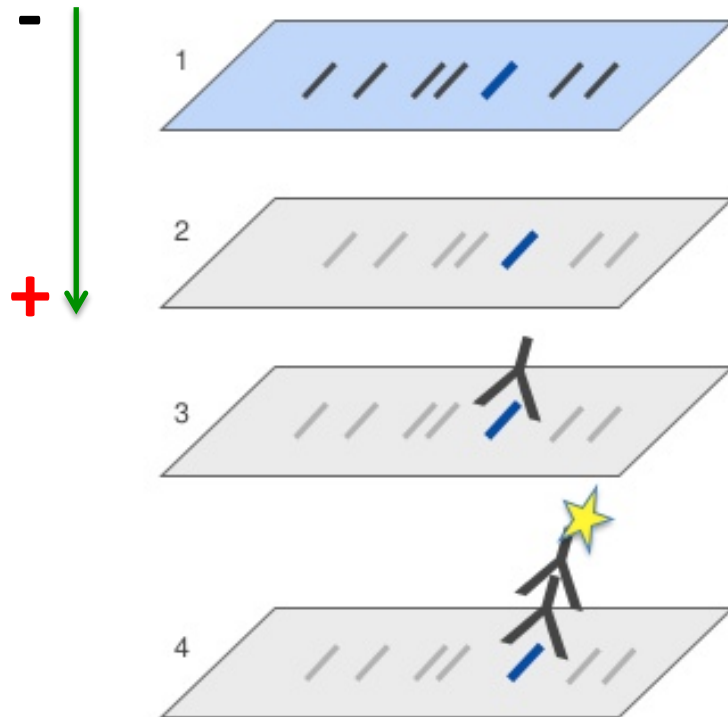
- TGS buffer **: sandwiched proteins form tight bands**

- + Tris-HCl 

- + SDS 

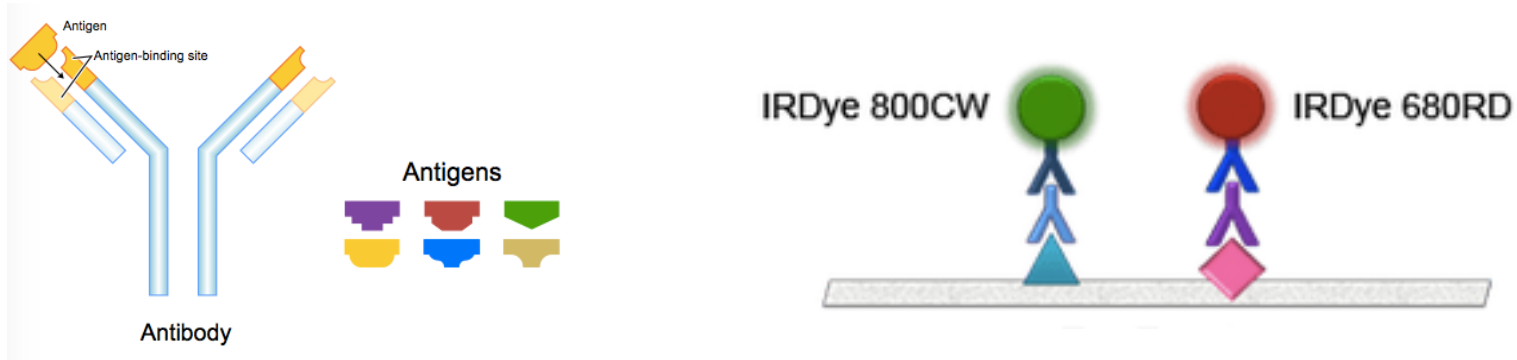
- + glycine 









Western blot workflow



1. Protein separation by SDS-PAGE
 - HiMark stained ladder
2. Protein transfer to nitrocellulose membrane
 -
 -
3. (Blocking and) probing with primary antibodies specific to
 -
 -
4. Probing with labeled secondary antibodies specific to primary antibodies
5. Image fluorescence signal

Suite of antibodies for *LI-COR* Western blot



protein of interest	 DNA-PKcs	 tubulin
primary antibody	 mouse anti-human anti-DNA-PK	 rabbit anti-human anti-tubulin
secondary antibody	 goat anti-mouse	 donkey anti-rabbit
fluorescent dye IR wavelength	800 nm	680 nm
pseudo-color	 green	 red
molecular weight	~ 465 kDa	~ 50 kDa

Key assignments of M2



- Journal club presentation
 - 10%
 - individual
 - in class at 1pm on Friday, March 18 or April 8



- Research article
 - 25%
 - individual
 - due 5pm on Monday, April 18
 - no draft/revision this time around