

- Announcements
- Lab Quiz
- Pre-lab Lecture
 - ❖ Writing a methods section
 - ❖ Library screen
 - ❖ Engineering tools/analogues
 - ❖ Today in Lab (Mod 2 Day 3)

Homework review

- HW comments for everyone
 - for figures, get *average* activity of duplicates
 - consider presentation: absolute or fold-change?
 - inherent cell activity or dilution variability?
 - ideal case outside of linear range
 - coupling of culture volume and OD600?
 - A420 vs. OD420?
 - meaning of ~~OD550~~ ^{less} < 0 ? round to \emptyset

Announcements

- Next time
 - come to *lab* first to set up experiment
 - no quiz!
 - we'll go together to 16-336 for j. club talks
- J. club grading rubric available up front
- Discuss tablet vs. iPad
- Error: Kan^R is for the EnvZ deletion, not the integration (see strain description)

Methods section tips

- Organizing sub-sections

start w/ an overview sentence → step-by-step details

topical, not necessarily day-by-day in lab

- Methods should be concise and complete

- Space-wise, avoid lists/tables when a sentence will do

- Sentence-wise, avoid extra words

- Content-wise, cover what's needed and only that needed to understand and replicate your work

- Concentrations are more useful than volumes; or you can state amounts, plus total volume.

- Read the guidelines!

Methods section exercise

- Consider the following passage: “Template DNA (5 ng) and primers were mixed with 20 uL of 2.5X Master Mix in a PCR tube. Water was added to 50 uL. A tube without template was prepared and labeled control.”
 - What information is missing?
 - What information can be cut?

Measuring LacZ

$$1 \text{ Miller Unit} = 1000 * \frac{(Abs_{420} - (1.75 * Abs_{550}))}{(t * v * Abs_{600})}$$

Team Color	B-gal units (dark)	B-gal units (light)
Red	1203	538
Orange	1183	736
Yellow	546?	305?
Green	8249? 600?	2881? 200?
Blue	1196	594
Pink	1163	423

Slide from N. Kuldell

Genetic library screen: concepts

- Goal: improve contrast of bact. photographs
- Specifically, make plates light er in the light
- Which mutations should have this effect? blue
- Why randomize at those sites?

EnvZ	A239T	G240E	V241G	S242D	H243A		blue = K-P+
Cph8 = Cph1/EnvZ	A553	G554	V555	S556	H557		red = K+ P-

Image from wiki

Genetic library screen: methods

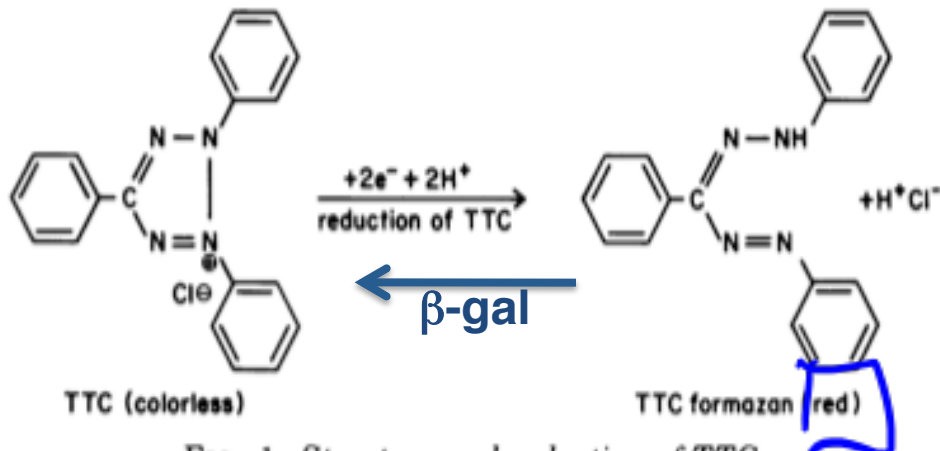
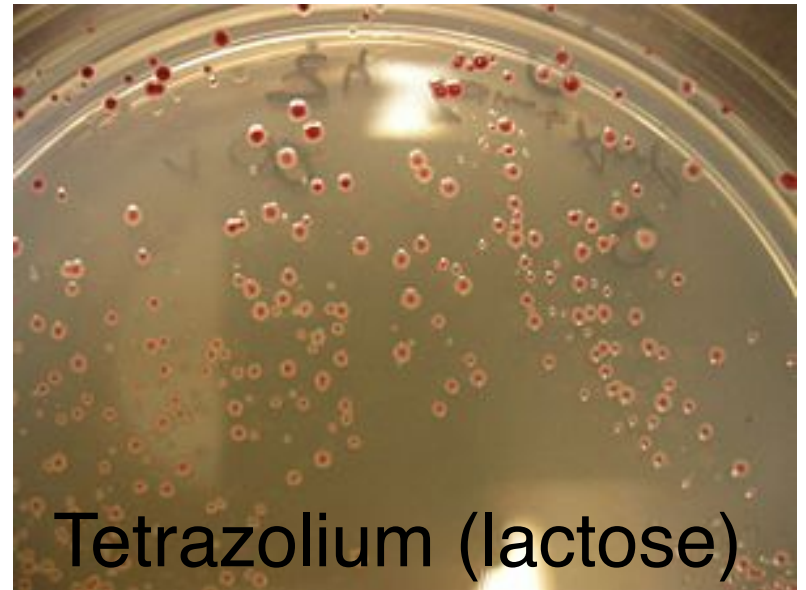


FIG. 1. Structure and reduction of TTC.



Tetrazolium (lactose)

APPLIED AND ENVIRONMENTAL MICROBIOLOGY, Feb. 1977, p. 434-444
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Generalized Indicator Plate for Genetic, Metabolic, and Taxonomic Studies with Microorganisms

BARRY R. BOCHNER^{1*} AND MICHAEL A. SAVAGEAU

Slide modified from N. Kuldell

Registry of Std Bio Parts

levels of abstraction

Goal: design protein generator that functions in *E. coli*

Browse parts and devices by function

This section replaces the previous *Featured parts* pages.

Browse parts by type

Catalog

List



Promoters (?): A promoter is of the downstream DNA sequence.



Ribosome Binding Sites (?): can bind and initiate translation.



Protein domains (?): Protein up a protein coding sequence target the protein for cleavage.



Protein coding sequences: Note that some protein coding protein from start codon to stop also included here.



Translational units (?): They begin at the site of translation.

RBS



Biosynthesis: Parts involved in the production of a product.



Cell-cell signaling and quorum sensing: Parts involved in cell-cell communication.



Cell death: Parts involved in killing cells.



Coliroid: Parts involved in taking a bacterial cell.

Browse parts and devices by standard

Unless otherwise specified, most parts in the Registry comply with this assembly standard.

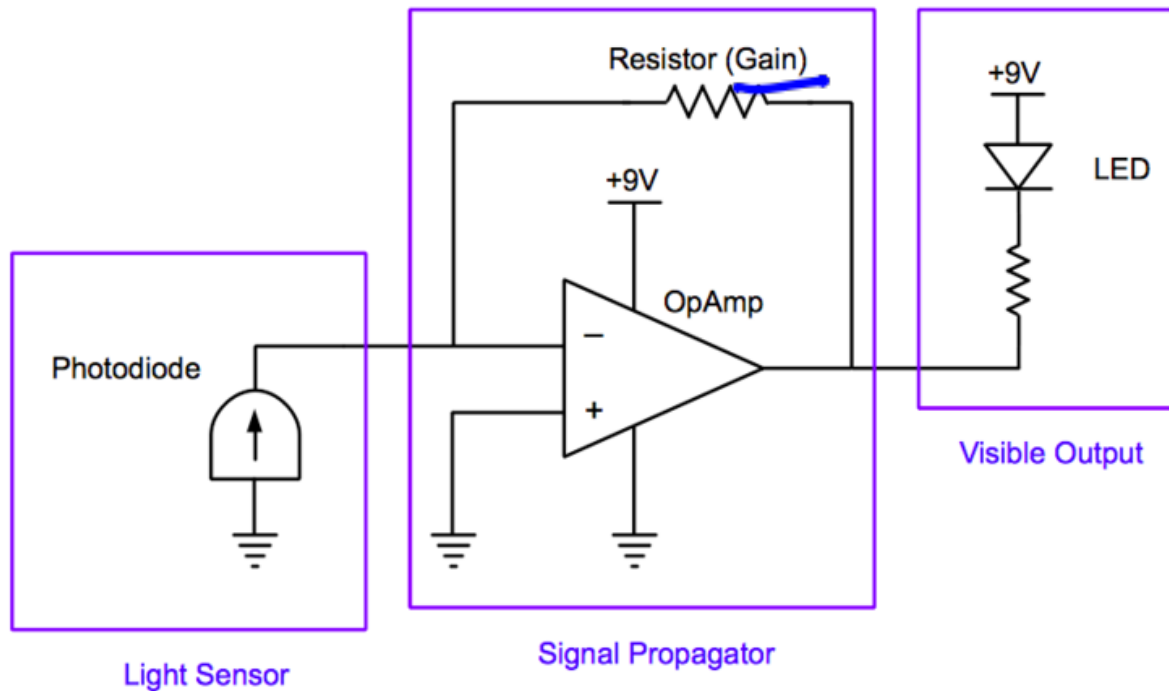
EX-SP

Assembly standard 10 (?): Assembly standard 10, parts that comply with this assembly standard.

Assembly standard 23 (?): Assembly standard 23, parts that comply with this assembly standard.

Images from parts.mit.edu

EE analogue to BP system



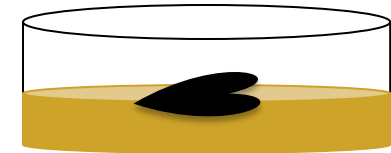
Goal: vary R and observe outcomes

limit cases: $R=0, \infty$

digital vs. analog

Today in Lab: M2D3

Observe/document *coli*roid from last time

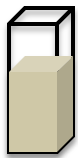


Set up library screen

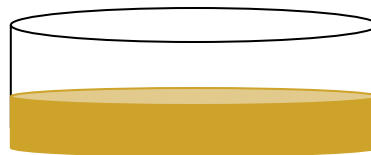
safety glasses

1. electroporate

2. plate cells (BPΔCph8)



incubate 1 hr*

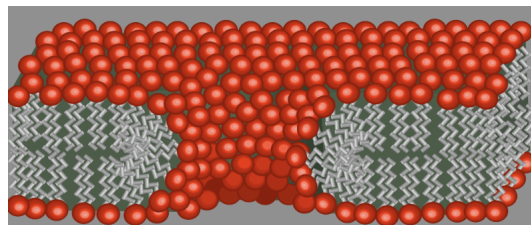
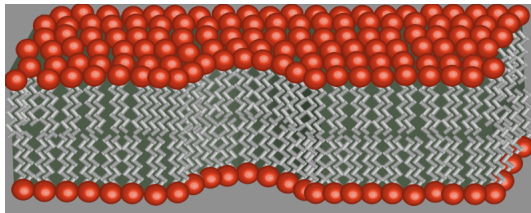


Explore EE analogue

Explore parts.mit.edu

Explore TinkerCell

*transfer to SOC medium
(has extra sugar, etc.)



Membrane model from Wikimedia Commons, public domain image