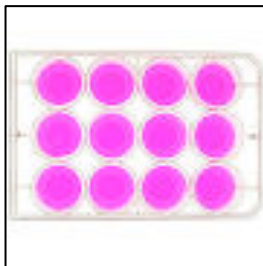
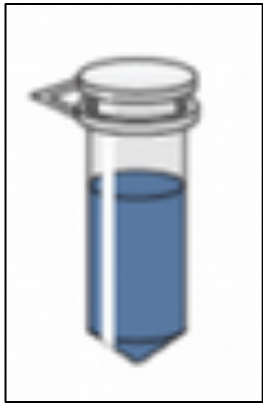


**M2D3:**  
**Finish Western blot;**  
**Extract RNA for qPCR assay;**  
**Treat cells for viability assay.**

03/16/2017

# In lab today



Student # 1 takes the lead

1. Finish Western blot analysis of cell lines

Student # 2 takes the lead

2. Prepare qPCR samples

- extract RNA from DLD-1  
BRCA2-/- +/- etoposide
- synthesize cDNA

Student # 1 takes the lead

3. Treat cells for viability assay

- Compound 401 (+/- etoposide)
- olaparib

# Key assignments of M2



- Journal club presentation [video-recorded](#)
  - 15%
  - individual
  - in class at 1pm on March 23 or April 11



- Research article
  - 20%
  - individual
  - due 10pm on April 22
  - no draft/revision this time around

**From Prof. Samson's lecture 03/09/17:**

**What experimental question will you ask in Module 2?**

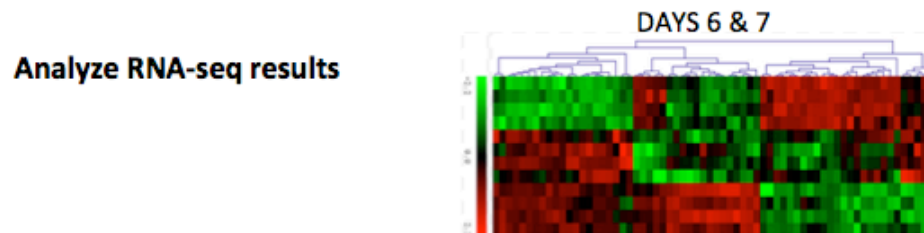
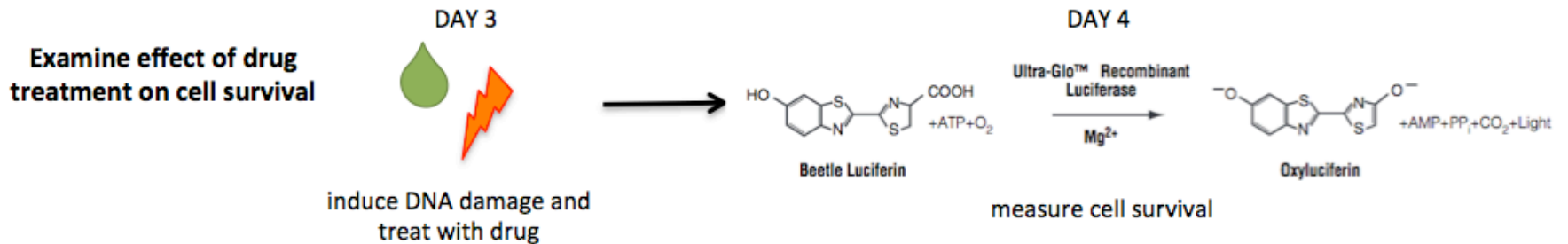
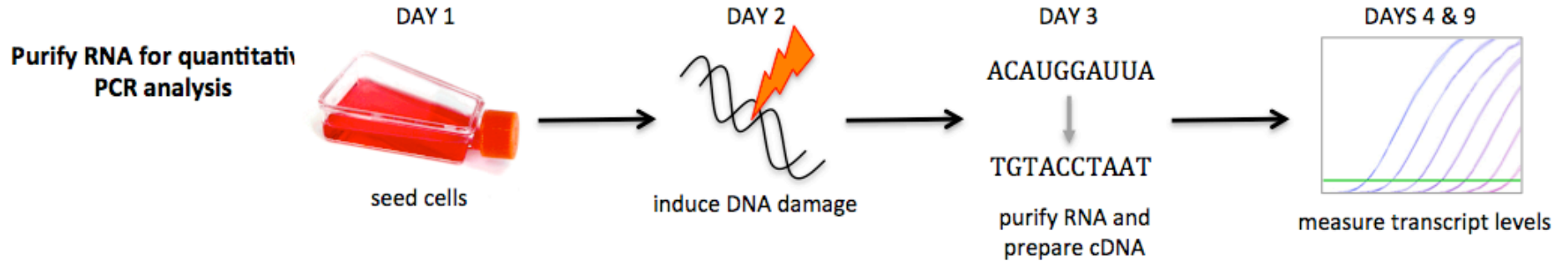
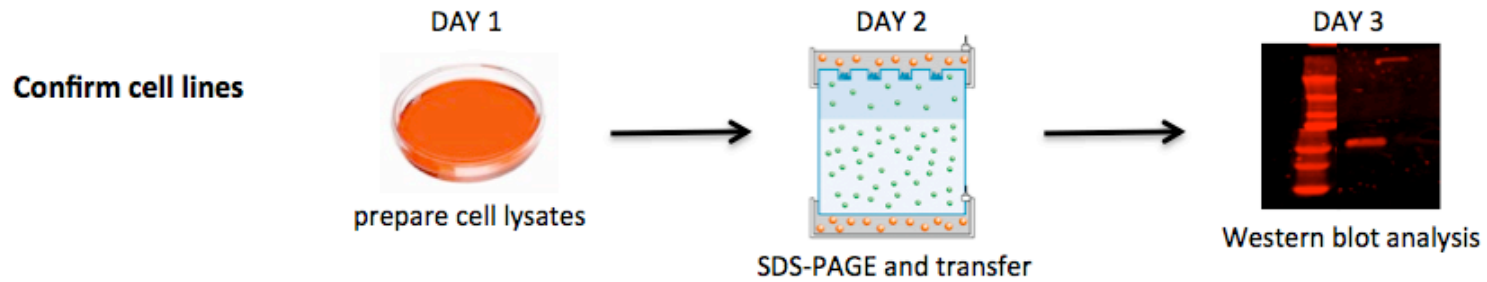
How does DNA repair affect the ability of cancer chemotherapy drugs to kill cancer cells?

How does cancer chemotherapy affect gene expression?

**This raises the following questions**

- How does DNA get damaged?
- What is DNA repair?
- Why does DNA repair exist?

# M2: Experimental overview

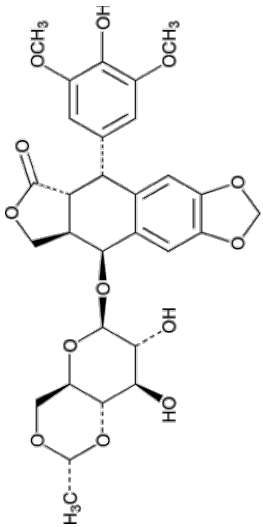


# In lab on snow day (M2D2)



1. Verify cell lines by **Western blot** protein analysis:  
“immunofluorescence on PAGE-separated proteins”

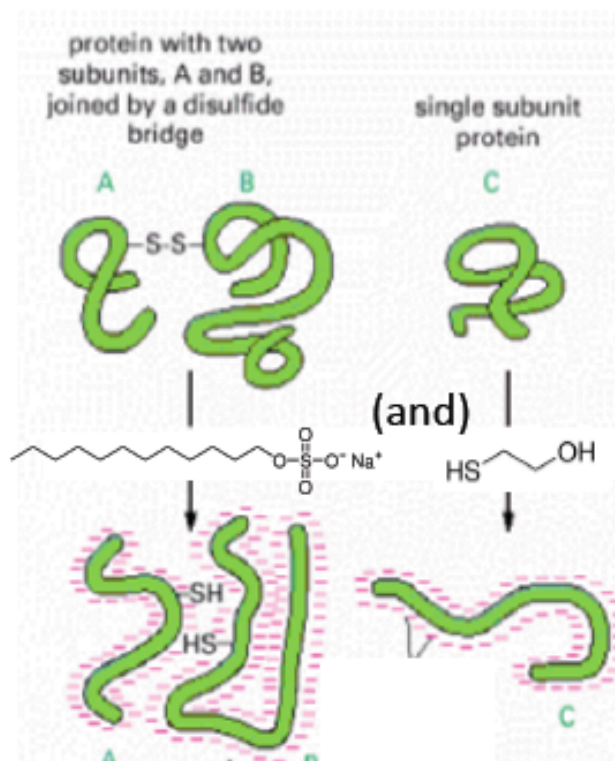
- Lyse DLD-1 and BRCA2-/- cells
- Measure protein concentration
- Separate proteins by SDS-PAGE
- Transfer proteins onto nitrocellulose membrane
- Label BRCA2 with primary+secondary antibodies



2. Treat cells with cancer drug **etoposide**

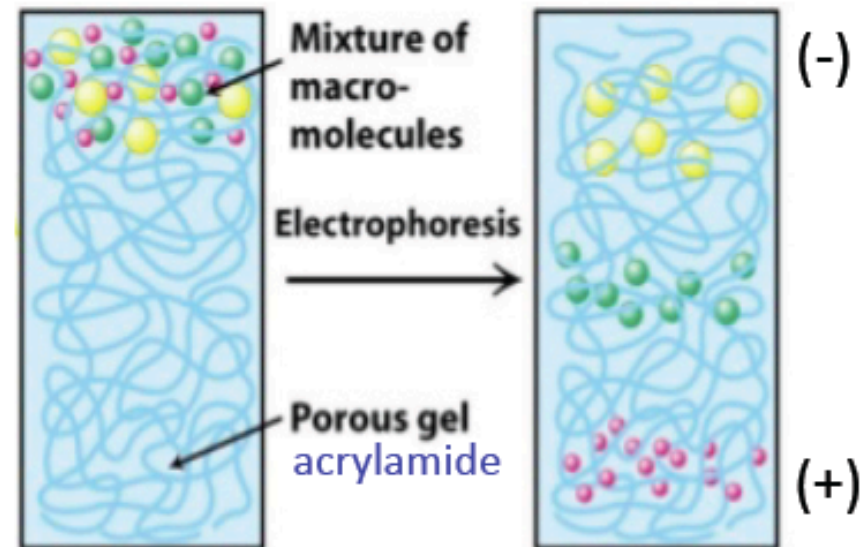
# Review: SDS-PAGE

- What gives proteins uniform charge and linear structure?



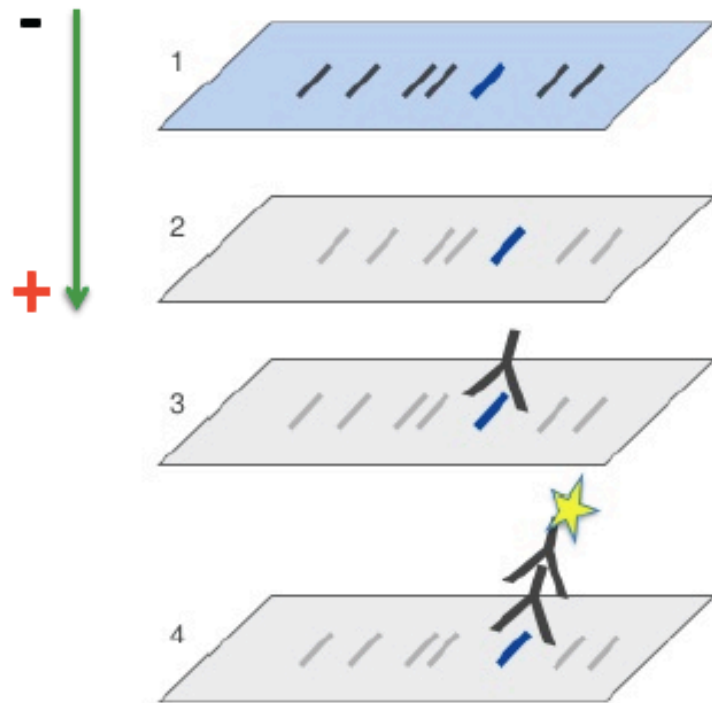
Laemmli buffer's  
SDS + beta-mercaptoethanol

- How are proteins separated?



by size (now proportional to charge)

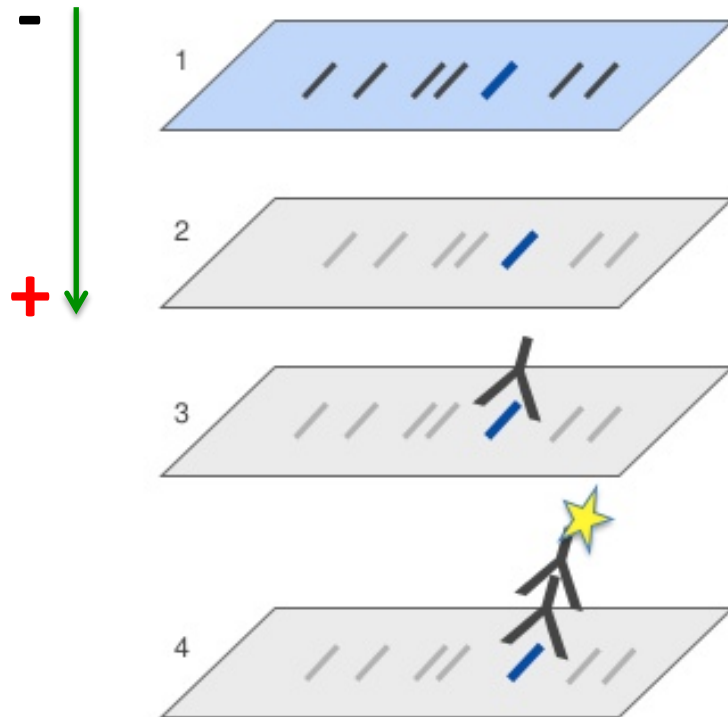
# Western blot workflow **on snow day**



1. Protein separation by SDS-PAGE
  - HiMark stained ladder **bands 31-460 kDa**
2. Protein transfer to nitrocellulose membrane
  - **high affinity for proteins**
  - **immobilizes proteins**
3. (Blocking and) probing with primary antibodies specific to
  - **BRCA2 (380 kDa)**
  - **tubulin**
4. Probing with labeled secondary antibodies specific to primary antibodies
5. Image fluorescence signal

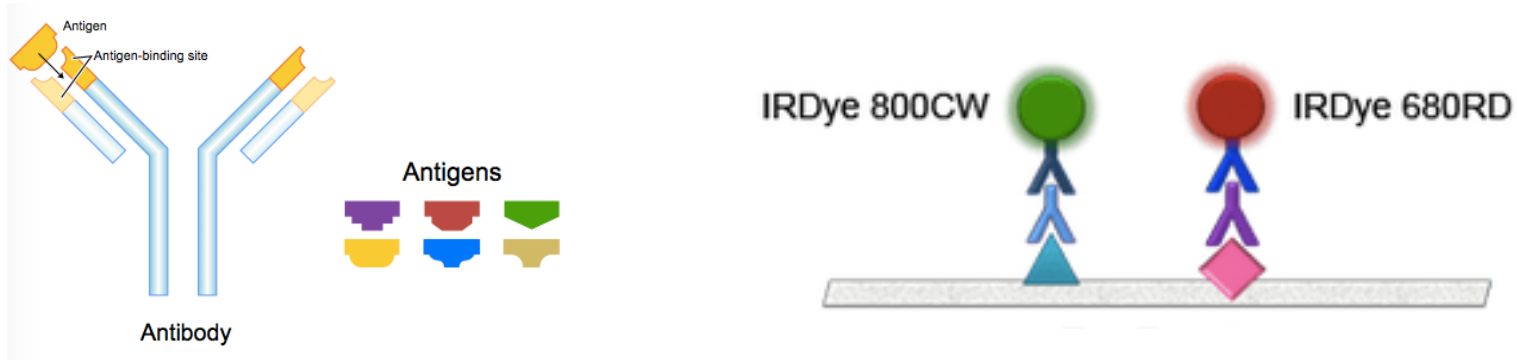


# Western blot workflow: today











1. Protein separation by SDS-PAGE
2. Protein transfer to nitrocellulose membrane
3. Block membrane **address non-specific binding**
4. Probe with primary antibodies specific to
  - BRCA2 **24 hours ago**
  - tubulin
5. Wash with TBS-T **Tris-buffered saline**
  - **Tween, a mild detergent**
  - to **get rid of unbound primary antibody**
  - and to **detach weak interactions**
6. Probe with labeled secondary antibodies specific to primary antibodies
7. Wash
8. Image *LI-COR* fluorescence signal

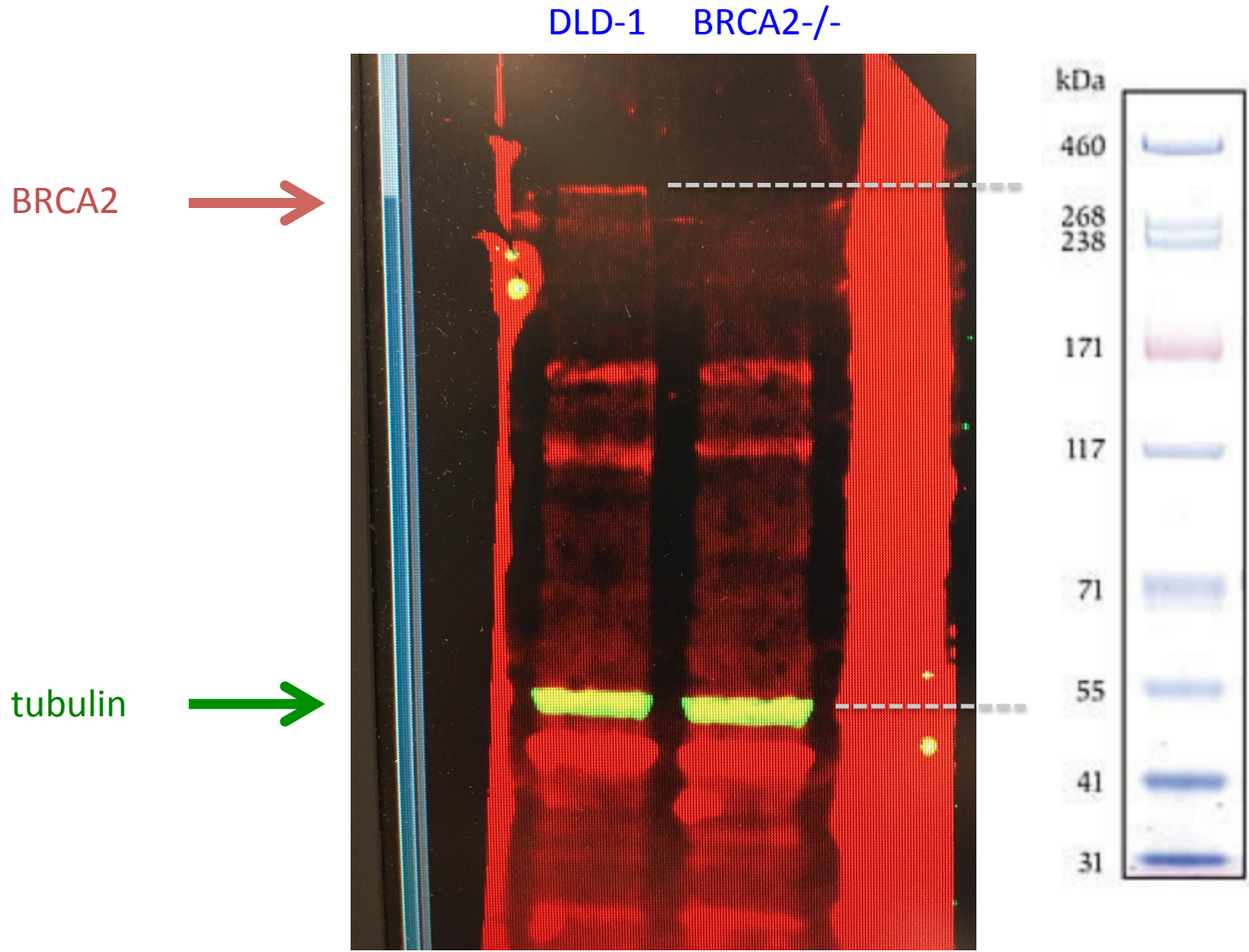
# Suite of antibodies for *LI-COR* Western blot




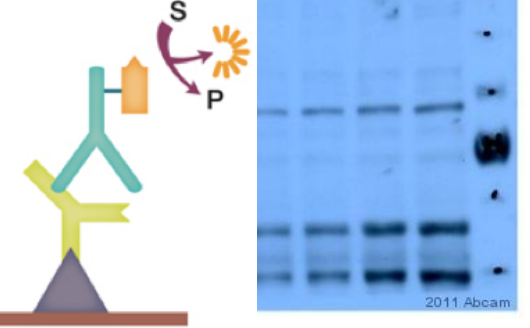
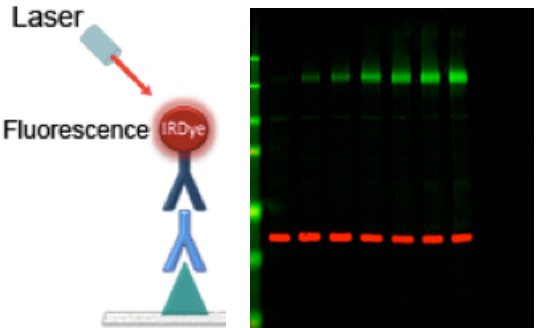
antigens

protein of interest	 tubulin	 BRCA2
primary antibody	 mouse anti-human anti-tubulin	 rabbit anti-human anti-BRCA2
secondary antibody	 goat anti-mouse	 donkey anti-rabbit
fluorescent dye IR wavelength	800 nm	680 nm
pseudo-color	 green	 red
molecular weight	~ 50 kDa	~ 380 kDa

# Verify BRCA2 is missing by *LI-COR* Western blot



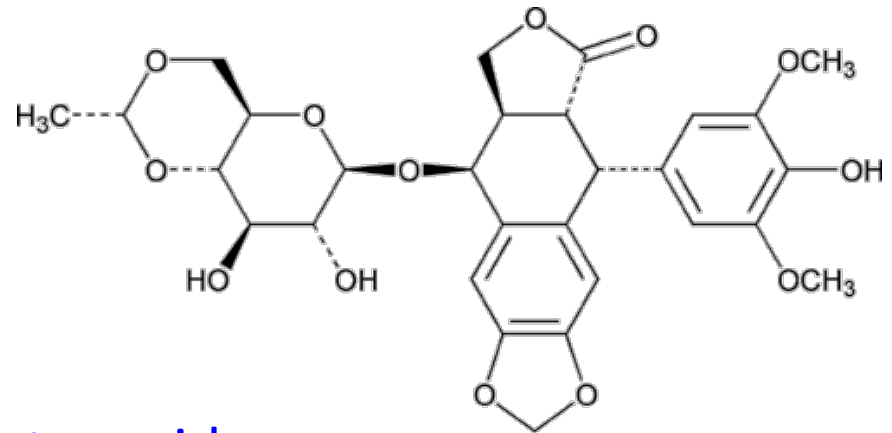
# Western blot detection / visualization

Colorimetric	Chemi-luminescent	Fluorescent (Li-COR)
		
<p>Upon incubation with a substrate that reacts with reporter (<i>e.g.</i> peroxidase), dye rendered insoluble and colored precipitates on membrane.</p>	<p>Incubation substrate luminesces when exposed to reporter on secondary antibody.</p>	<p>The fluorescently labeled probe is excited by light and the fluorescence emission is detected by a photosensor such as a CCD camera.</p>
<p><b>Pro:</b> inexpensive, easy, no equipment required</p>	<p><b>Pro:</b> sensitive, fast, film developer is common</p>	<p><b>Pro:</b> sensitive, stable, able to multiplex</p>
<p><b>Con:</b> medium sensitivity</p>	<p><b>Con:</b> requires trial and error, time-dependent snapshot</p>	<p><b>Con:</b> expensive</p>

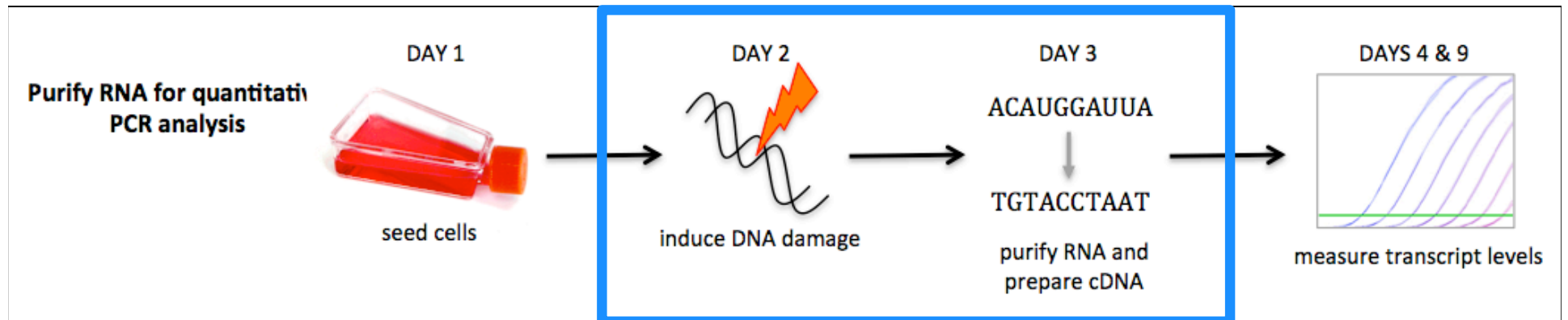
# Homework due M2D4: use Western blot

- Figure (with title and caption)
- Results (no more bullet points, paragraph form)
  - one title (but several paragraphs for several topics)
  - introductory topic sentence
  - state findings
  - conclude / transition to next result
- (Discussion / interpretation will be separate in M2 research article)

# Also on snow day

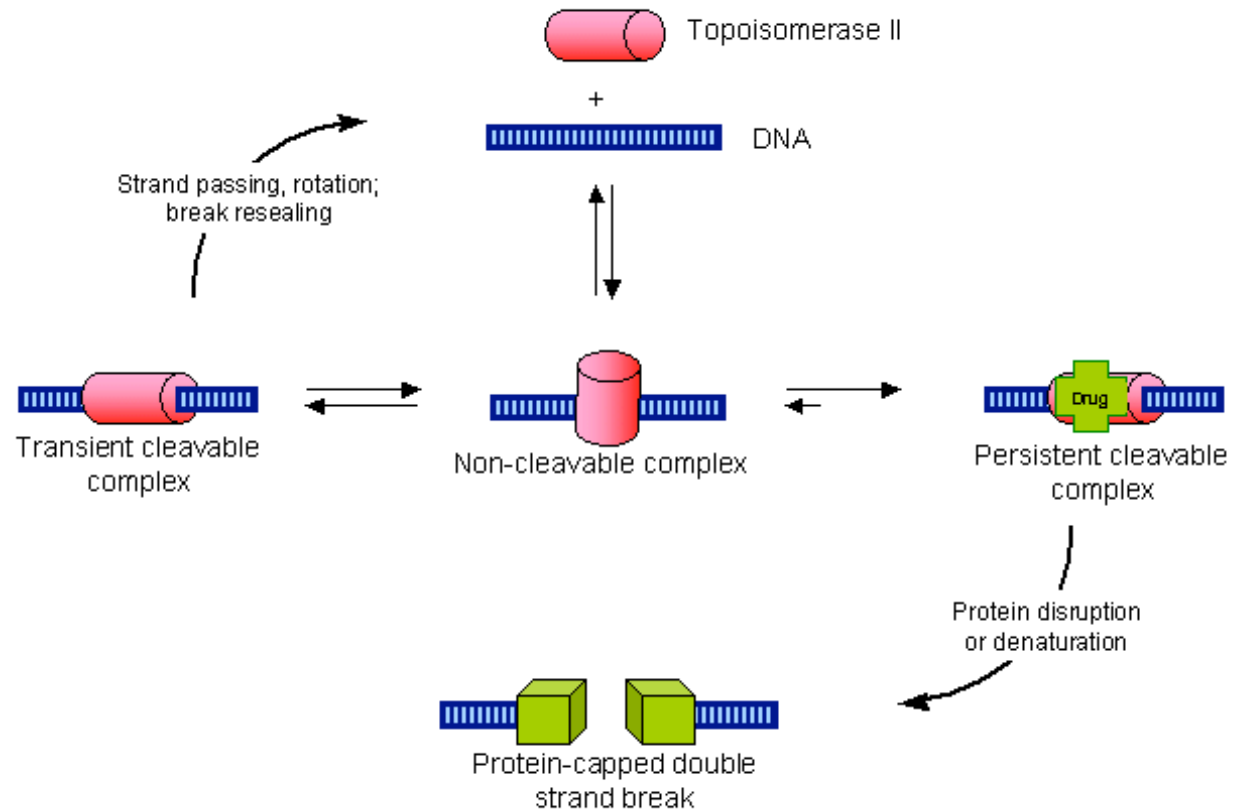


2. Treat cells with cancer drug **etoposide**



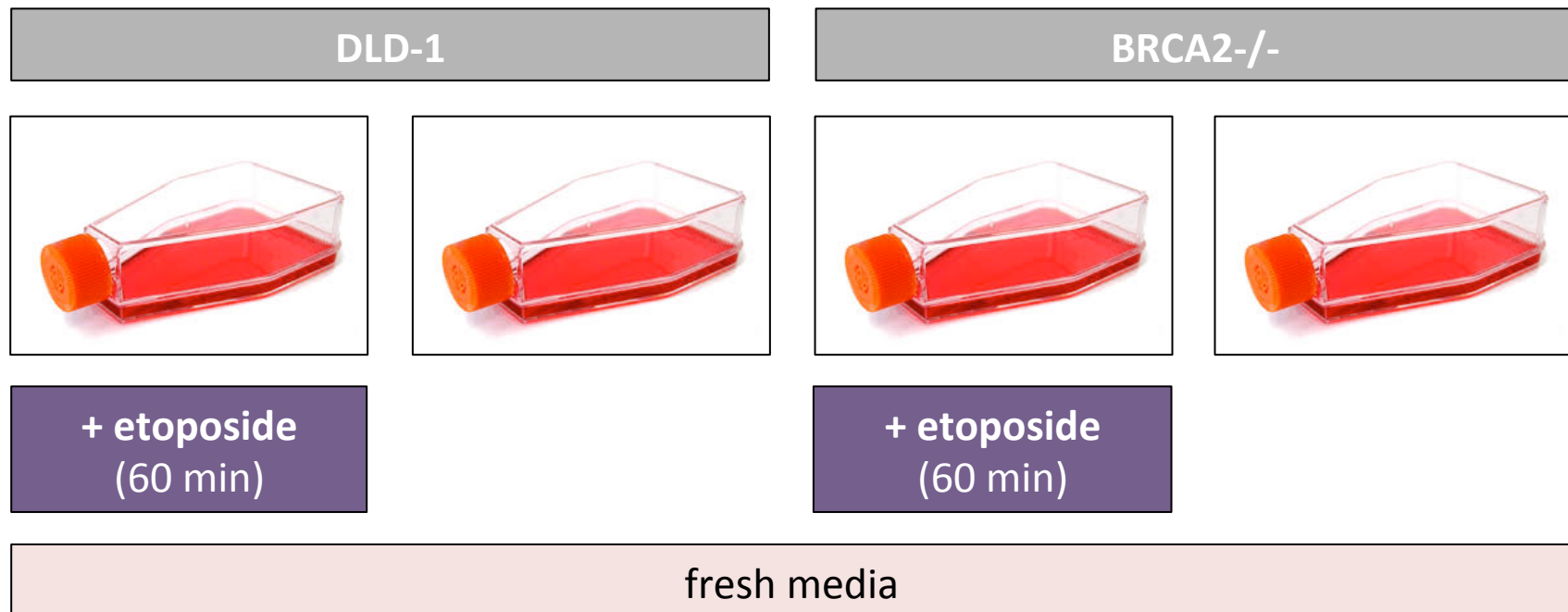
# Etoposide's mechanism of action

- forms a ternary complex with DNA and the topoisomerase II enzyme (which aids in DNA unwinding), **to remove supercoils of DNA**
- prevents re-ligation of the DNA strands,
- so causes DNA strands to break,
- and (cancer) cells to undergo apoptosis.



# Treat cells with etoposide

Thank you, Noreen!

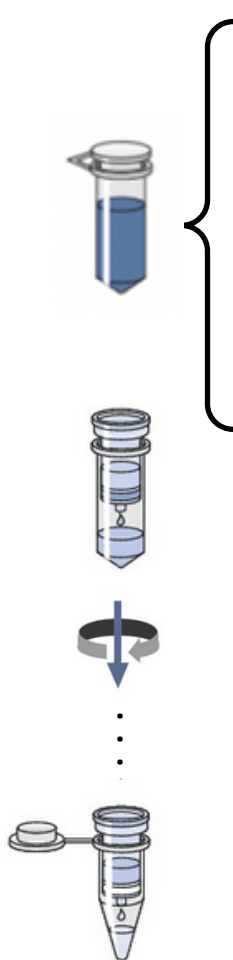


M2D3: extract RNA **today**

Has cancer drug affected transcriptional profiles / gene expression?



# Isolate RNA: QIAshredder + Rneasy kit

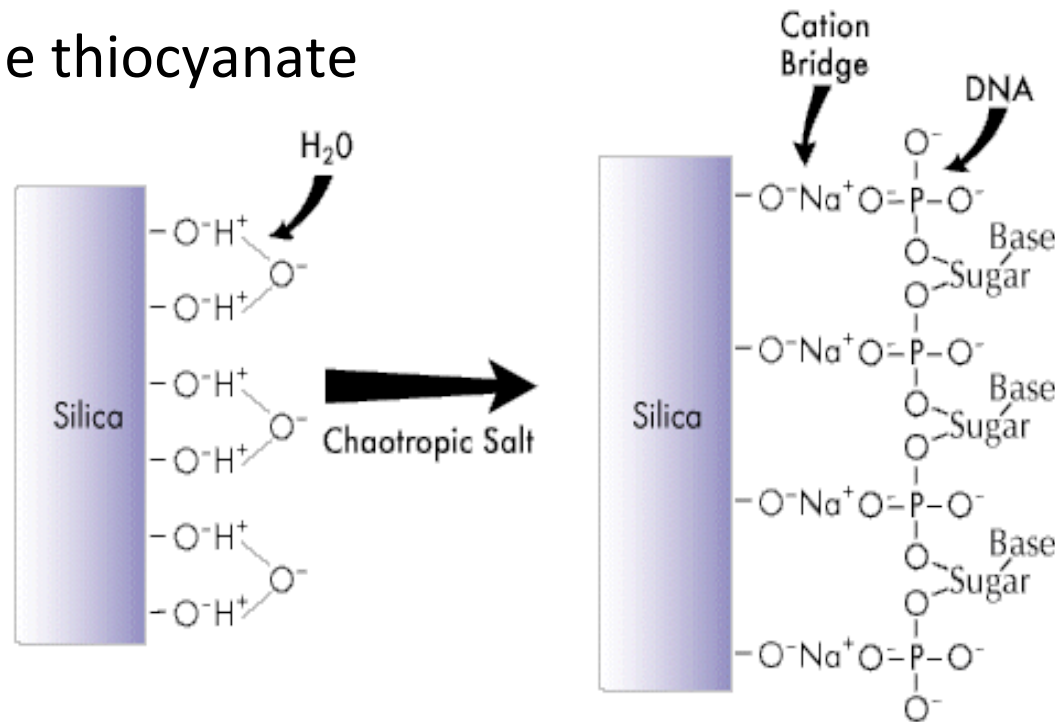


The diagram on the left illustrates the RNA isolation process. It shows a sequence of steps: 1. A microcentrifuge tube containing a blue liquid. 2. A microcentrifuge tube with a white pellet and a blue liquid supernatant. 3. A downward-pointing arrow with a circular arrow around it, indicating a transition or a specific step. 4. A microcentrifuge tube with a white pellet and a blue liquid supernatant, with a small amount of liquid being added from a pipette tip. A large curly bracket on the left side of the table groups the 'lyse' and 'prepare' steps.

steps	contents	purpose
lyse	RLT (with highly denaturing guanidine-thiocyanate salt) <b>detergents</b> + QIAshredder	inactivate RNase, disrupt membranes, helps bind column  homogenize (shear high-MW genomic DNA)
prepare	ethanol	promote efficient binding
bind	silica membrane in column, retain mRNA (> 200 nucleotides)	
wash	RW1 RPE	<b>get rid of contaminants</b> ** then, get rid of <i>all</i> ethanol
elute	water, RNase-free	high-purity RNA

# Chaotropic salts help DNA/RNA bind to column

*e.g.* guanidine thiocyanate



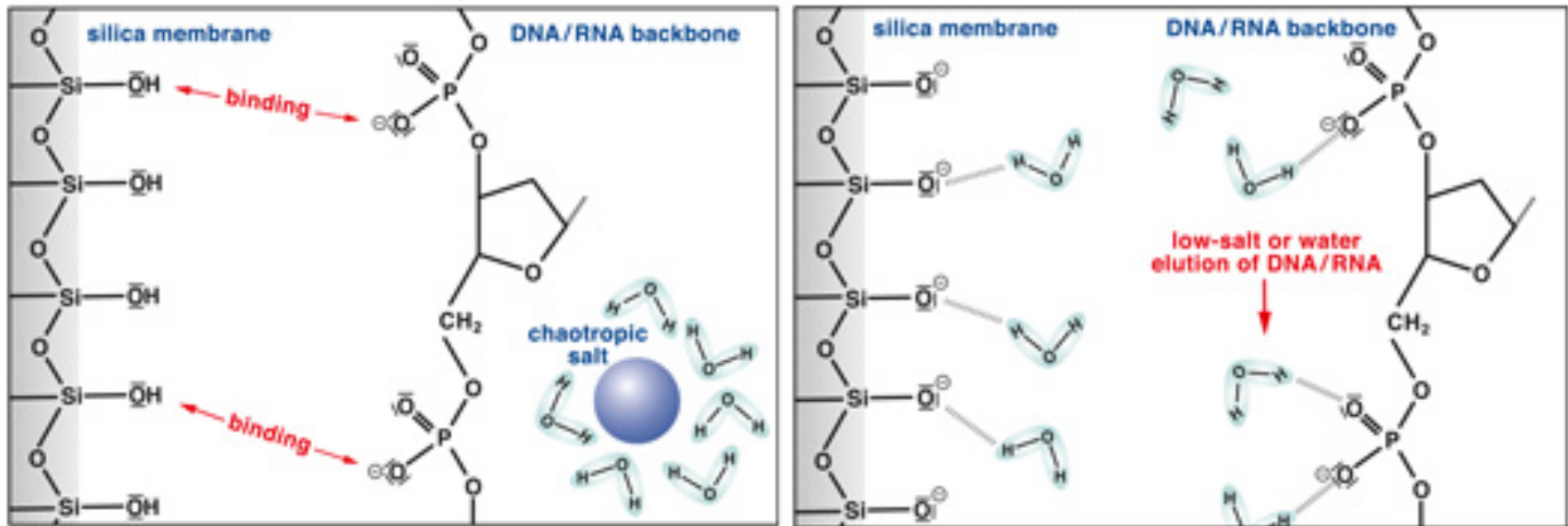
- Washes with RW1 and RPE remove residual contaminants
  - RW1 contains a guanidine salt, as well as ethanol, and is used as a stringent washing buffer that efficiently removes biomolecules such as carbohydrates, proteins, fatty acids, etc, that are non-specifically bound to the silica membrane
  - RPE contains ethanol and is a mild washing buffer

from Qiagen

# Water is used to elute nucleic acids

or low ionic-strength solvent

- Water competes RNA off of column



before elution

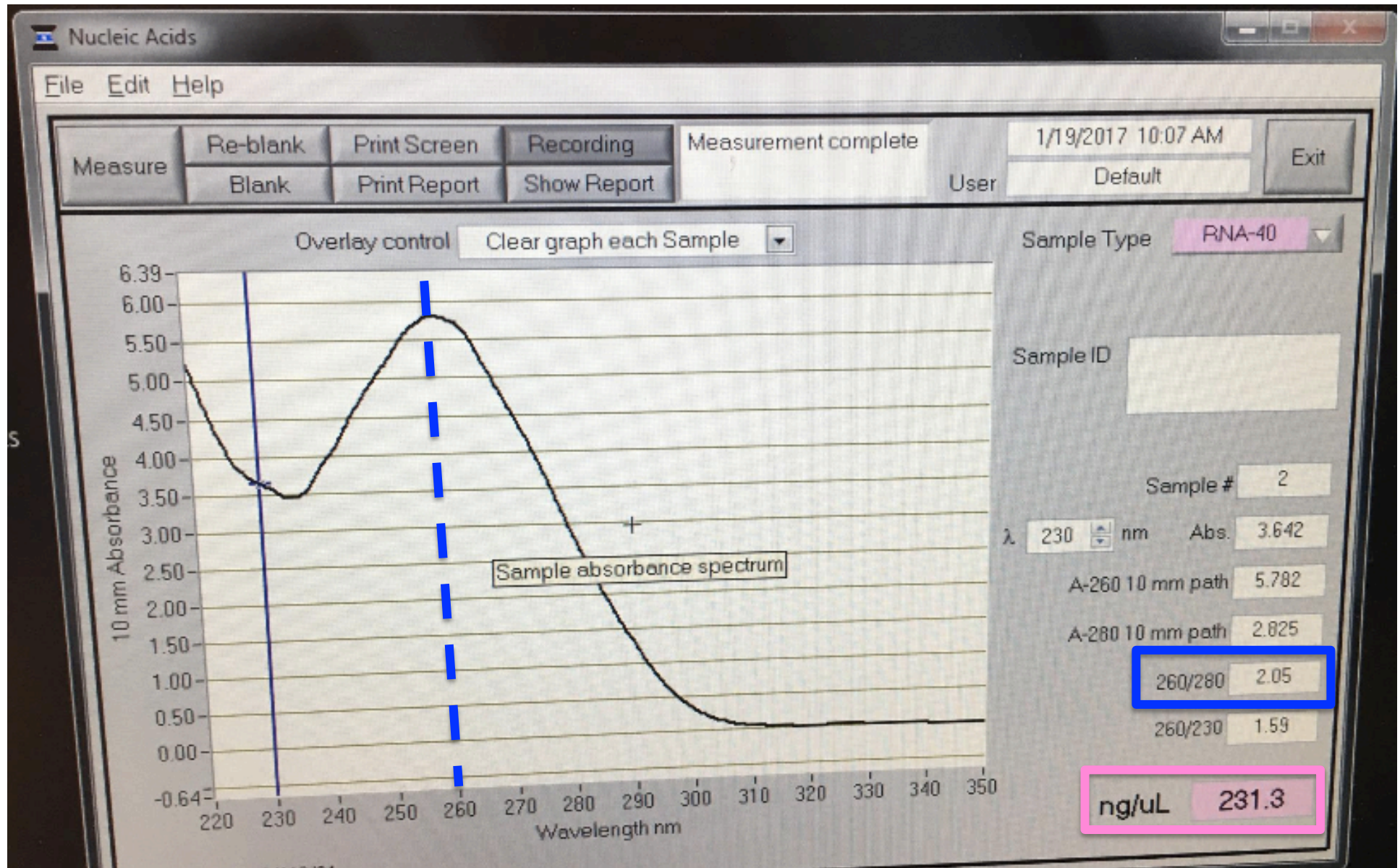
at elution

# RNA concentration from NanoDrop spectrophotometer

- $A_{260}/A_{280}$ 
  - nucleic acids absorb at 260 nm
  - proteins absorb at 280 nm
  - ratio ~ 1.8 “pure” DNA
  - ratio ~ 2.0 “pure” RNA
  - note:  $A_{230}$  from contaminants (phenol, guanidine, carbohydrates,..)



# RNA concentration from NanoDrop



# Synthesize cDNA from purified RNA

- cDNA: **complementary DNA**
- RT-PCR: **reverse transcription polymerase chain reaction**

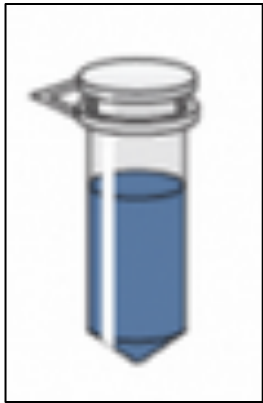


steps	conditions	ingredients added
denature	65°C 5 min on ice 1 min	1 µg RNA + primers + dNTPs
anneal	oligo (dT) <sub>20</sub>	primer: oligo (dT) <sub>20</sub>
synthesize	50°C 50 min	SuperScript III RT, RNaseOUT, MgCl <sub>2</sub> , DTT, buffer
terminate	85°C 5 min	
remove RNA	37°C 20 min	RNase H
PCR amplify	see M2D4	

# In lab today

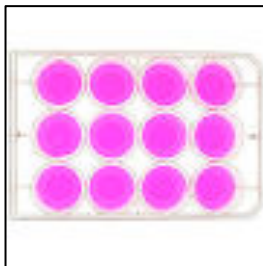


1. Finish Western blot analysis of cell lines



2. Prepare qPCR samples

- extract RNA from DLD-1  
BRCA2-/- +/- etoposide
- synthesize cDNA



3. Treat cells for viability assay

- Compound 401
- olaparib

# Treat cells to examine **viability**

