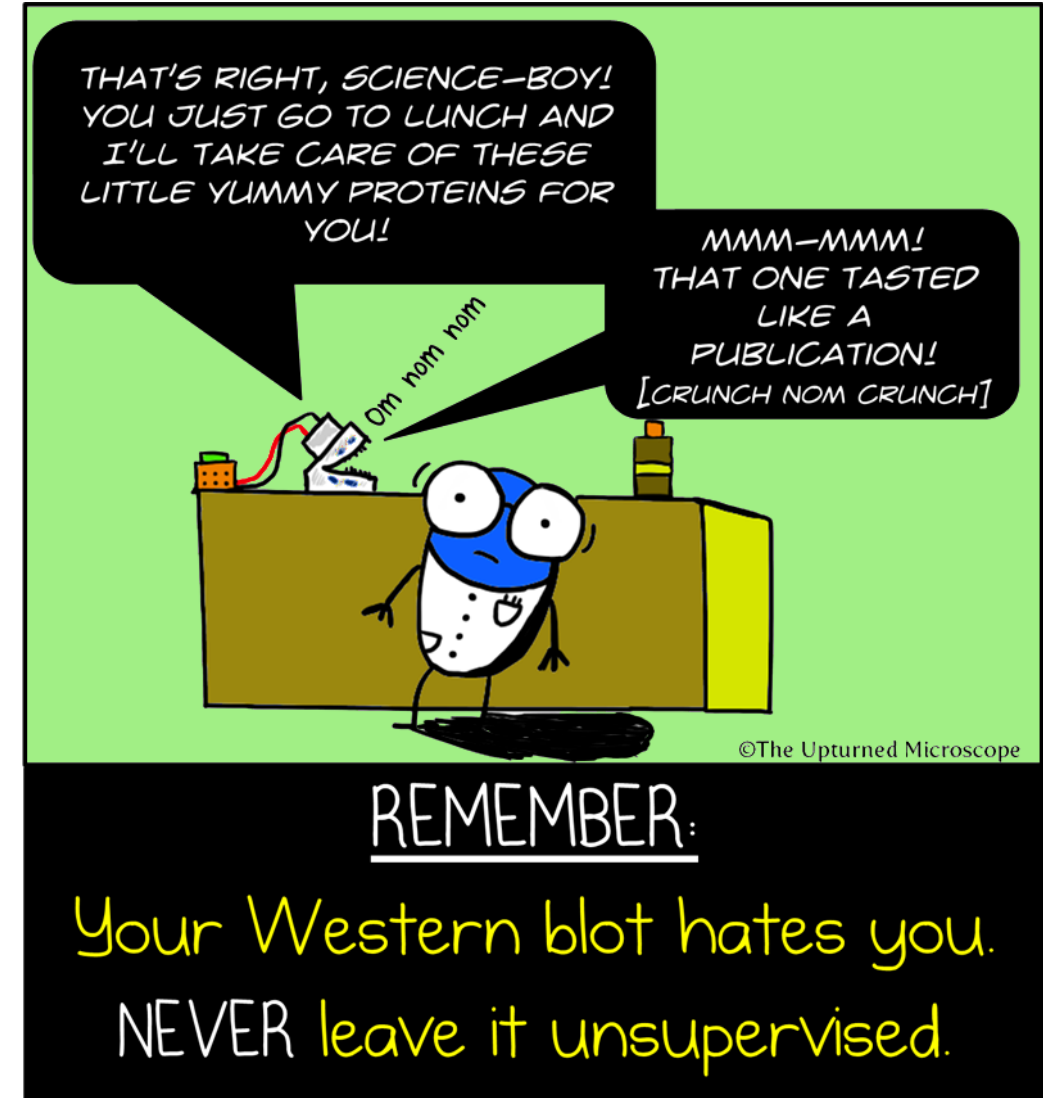


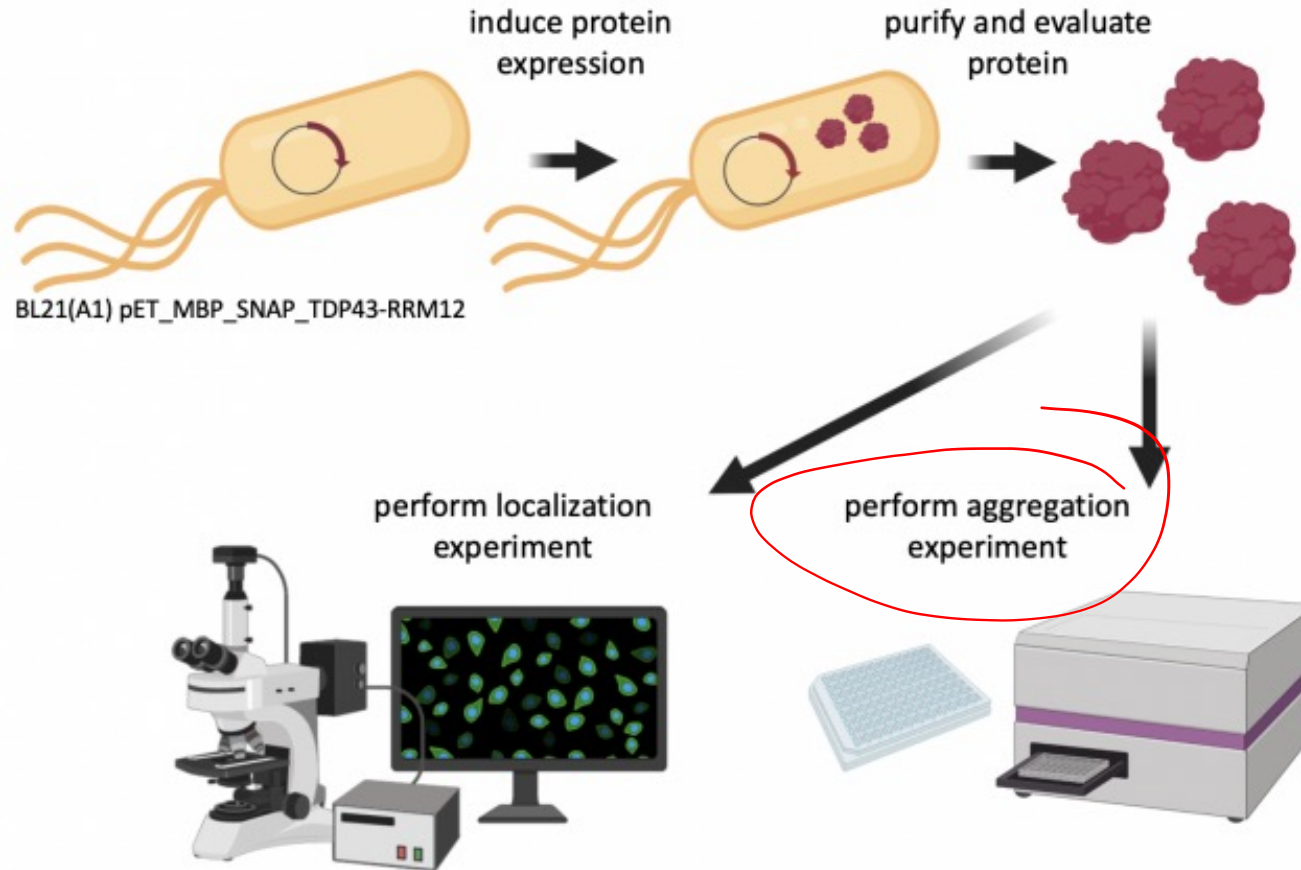
# M1D4: Assess purity and concentration of purified protein

1. Comm Lab
2. Quiz
3. Prelab discussion
4. Visualize protein purity with SDS-PAGE
5. Measure protein concentration with BCA assay



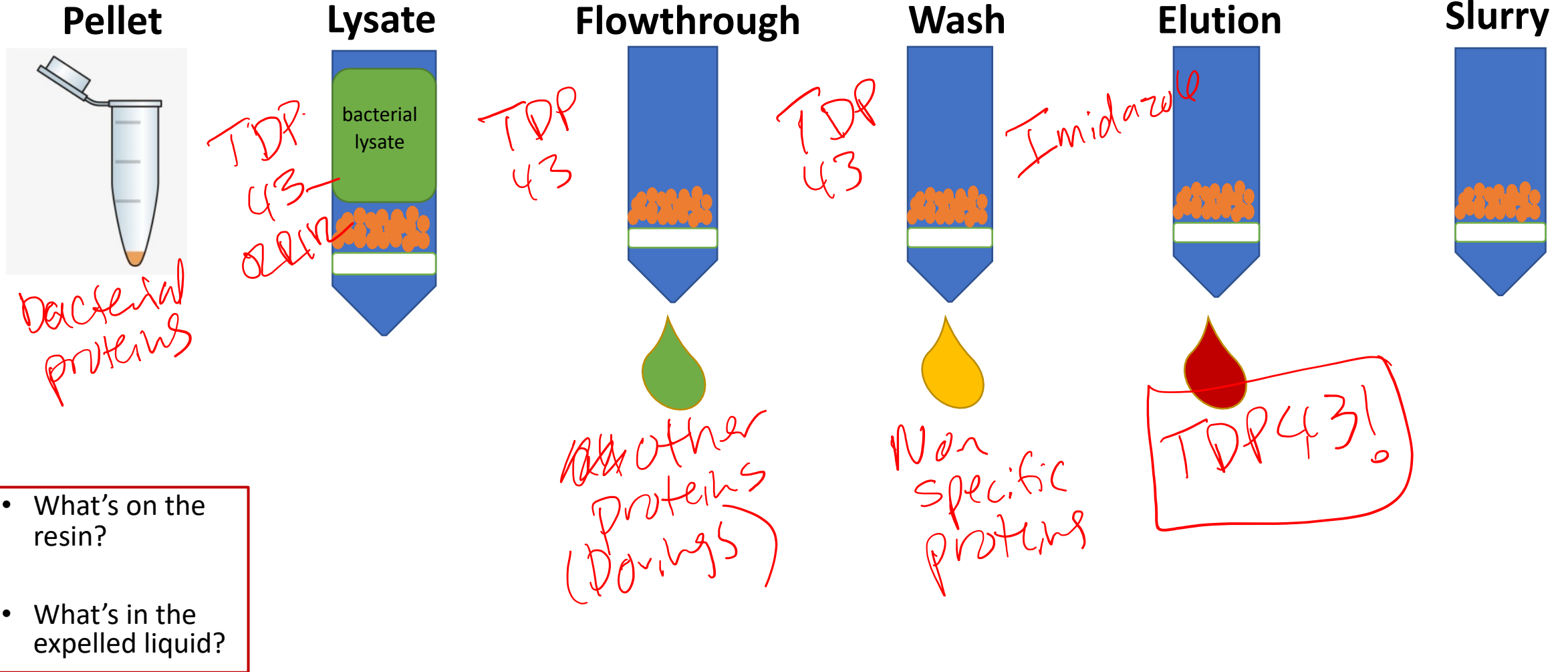
# Overview of Mod 1 experiments

**Research goal: Use functional assays to characterize ligands identified as binders to TDP43 from SMM technology**



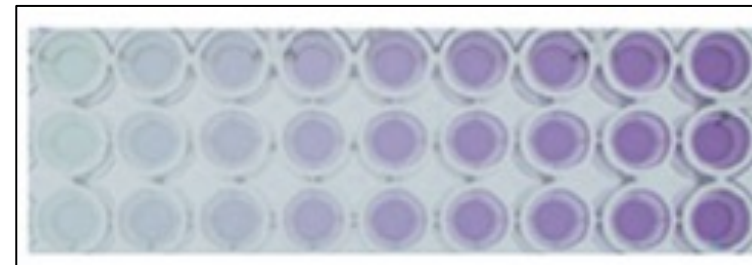
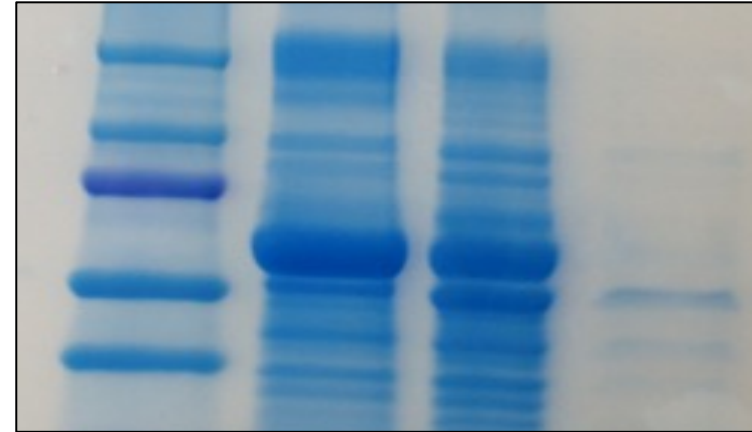
# Protein purification review

- Why this step? → *for you later*

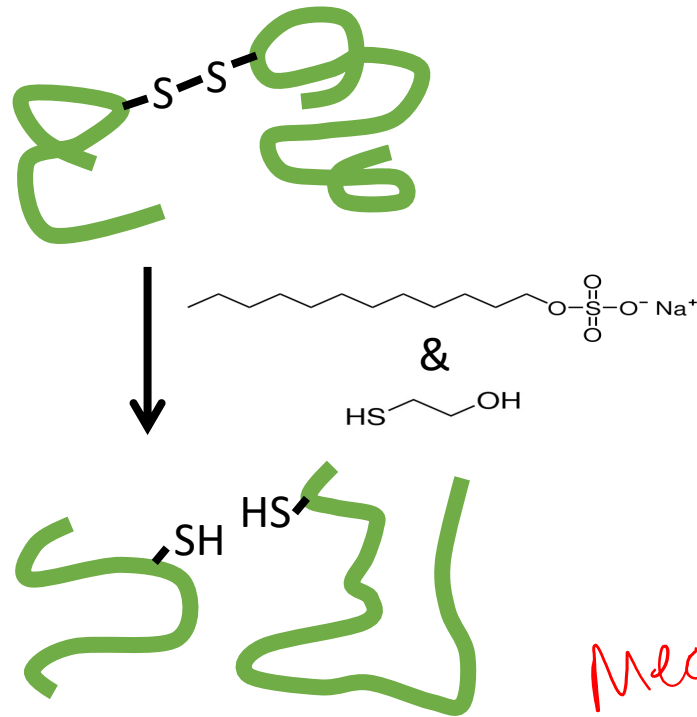


# How will you assess purity and concentration?

- Check **purity** using **SDS-PAGE**
  - Visual detection of all proteins in sample
  - Identifies purity of sample at multiple stages of purification
- Measure **concentration** using **BCA assay**
  - Colorimetric assay
  - Calculate concentration from standard curve



# Purity: Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE)



## Chemical processes

- Laemmli sample buffer / loading dye:

- SDS *detergent (surfactant -denature, (-) charge*
- $\beta$ -mercaptoethanol (BME) *-breaks disulfide bonds*
- bromophenol blue *-tracking proteins*
- glycerol *-weight sample in well*

## Mechanical

- Boiling:

*-denature*

# How are proteins separated?

- Laemmli buffer and boiling results in denature and (-) charged proteins

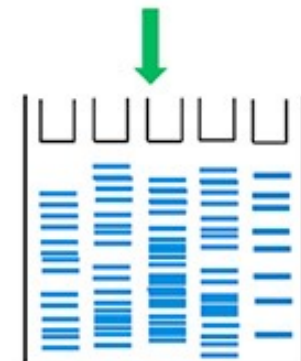
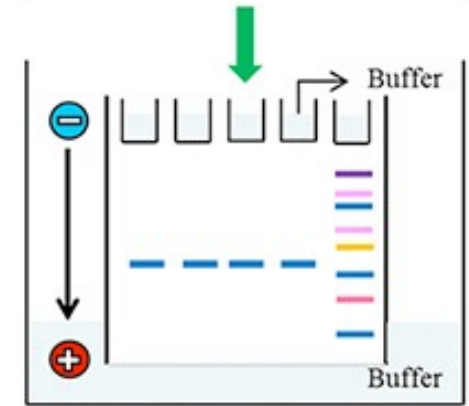
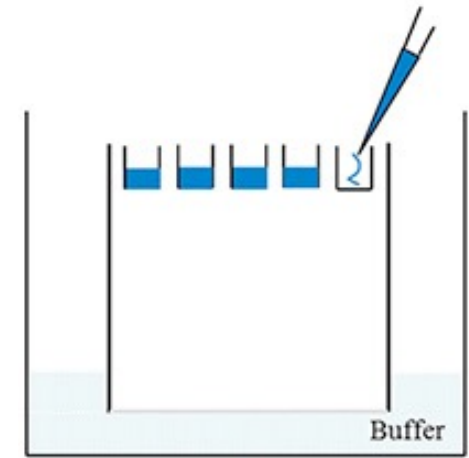
- SDS-PAGE separates proteins by Size

- Electrophoresis completed in TGS buffer

- Tris-HCl
- SDS
- Glycine

Buffers  
pH control  
Ions

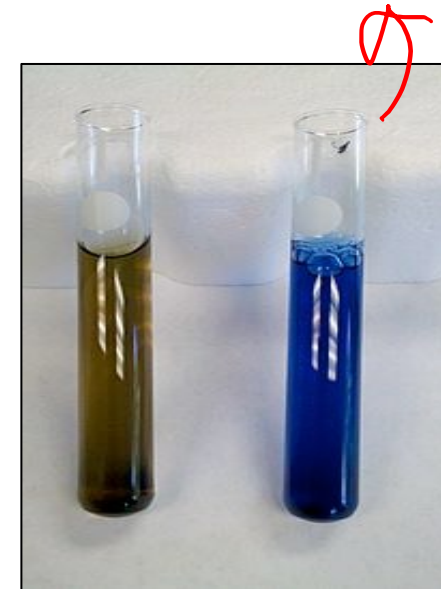
Running buffer



# How are proteins visualized?

Coomassie brilliant blue G-250 dye used to stain gel after electrophoresis

- Red if unbound (cationic form)
- Blue if bound to protein (anionic form)
- Hydrophobic and electrostatic interactions with basic residues
  - Arg (also His, Lys, Phe, Trp)



# Be mindful when assessing SDS-PAGE protein samples

Consider the order of your samples:

1. molecular weight ladder
2. pellet
3. lysate
4. flow-through
5. wash
6. elution
7. resin



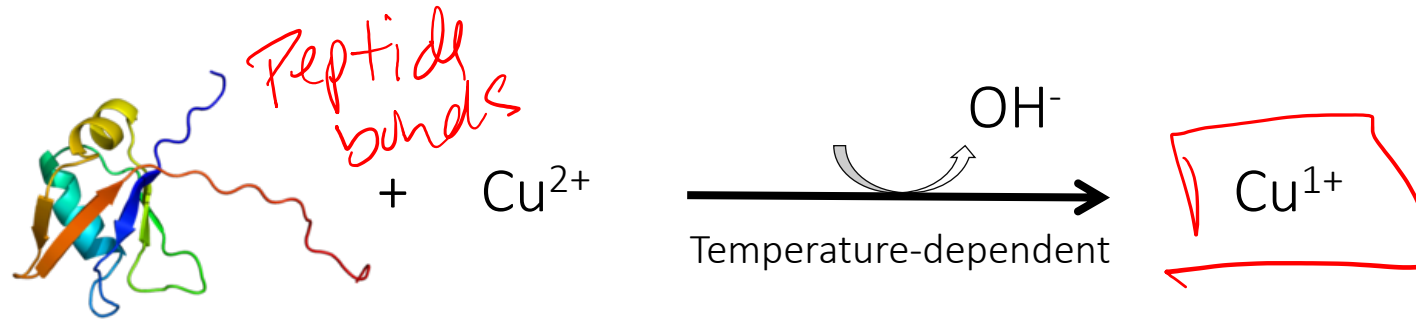
- Figure will be included in your Data Summary!



→ Elution

# Concentration: Bicinchoninic acid (BCA) protein assay

Step 1: Chelation of copper with protein, reduction of copper sulfate to copper ion



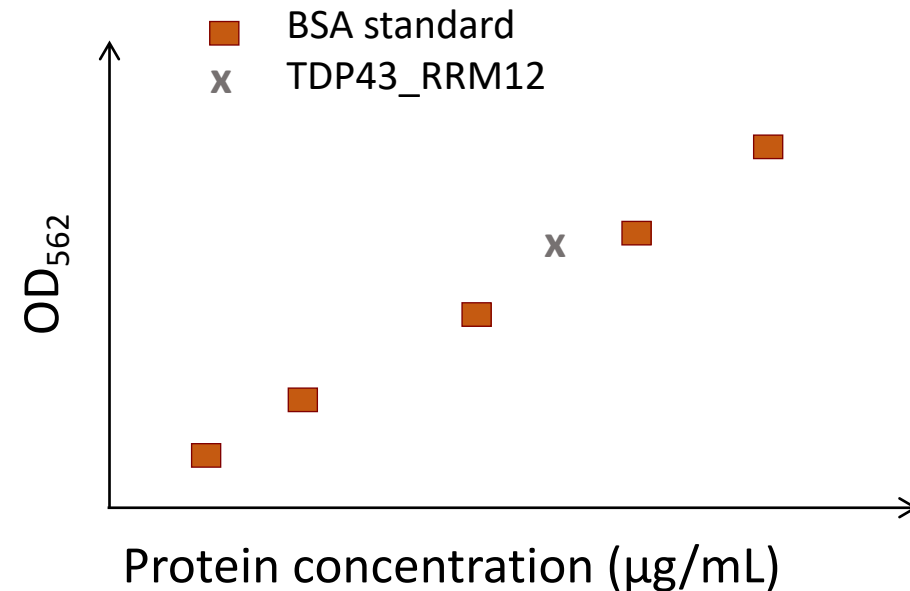
Step 2: BCA complexes with cuprous cation



# BCA/Cu<sup>1+</sup> absorbance proportional to protein concentration

Standard curve generated using serial dilutions of bovine serum albumin (BSA)

- Use fresh tips between tubes
- Mix well between dilutions
- Be mindful of volumes



# For today...

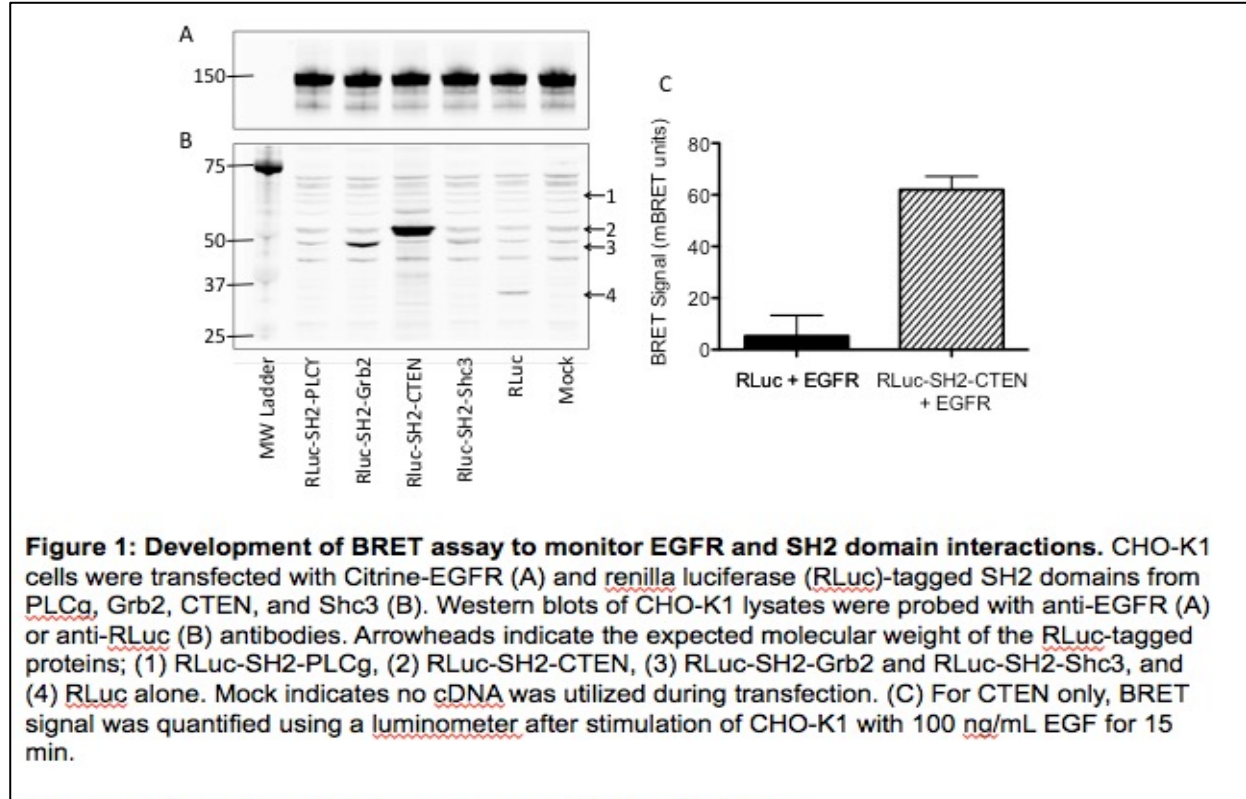
- Complete the purity and concentration assessments
  - It's good to divide the work load here!
  - Start by deciding who will be in charge of the two different techniques today

# For M1D5...

- Create a data figure of your purification results
  - must include SDS-PAGE gel
- Outline your Research Talk
  - See Assignments page for details and Homework page for checklists
  - Focus is aggregation experiment, just put a placeholder for actual results

# Data figure example

- Image **should not** be the entire page
  - Only needs to be large enough to be clear / visible
- Title **should** be conclusive
  - Don't include what you did, rather state what you found (take home message)
- Caption **should not** detail the methods or interpret the data
  - Define abbreviations, symbols, etc.
  - Info needed to “read” figure



# Notes on the Research Talk

- Individual assignment
- Three (3) minute video of you talking directly into the camera
- No visual aids allowed
  - Introduce yourself and your project
  - Highlight key results with quantitative information
  - Place your work in the scope of the larger field
- No need to state you are doing a class project or anything about 20.109
  
- Homework = outline
  - Introduction of your project
  - Key results from your research (including a statement as to the method(s) used to generate data)
  - Take-home message