

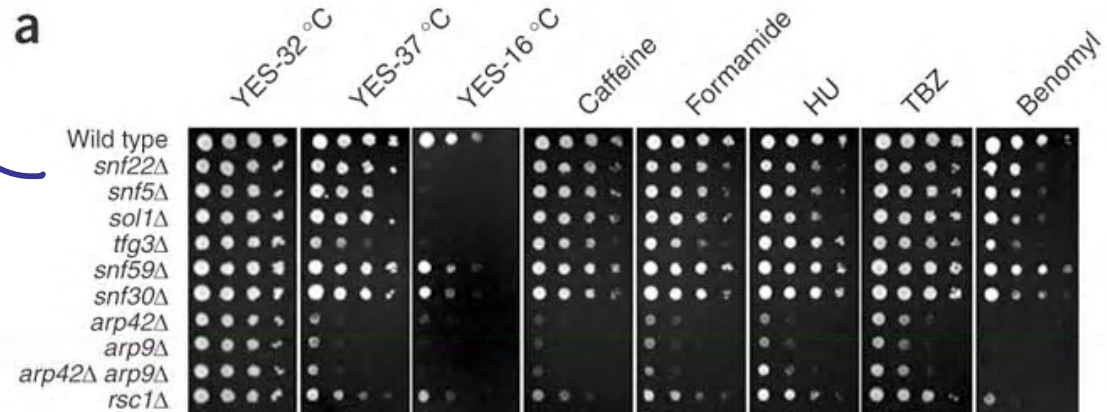
- Announcements, Old HW
- Lab Quiz
- Pre-lab Lecture
  - ❖ Microarray workflow
  - ❖ Intro to uArray analysis
  - ❖ Today in Lab, FNT

# Old HW

- Figure comments from Natalie

"at-a-glance"  
genotype labeling

First sentence tells  
how you got the data



(a) Growth phenotypes of the viable SWI/SNF and RSC mutants. Wild-type (FWP52), *snf22Δ* (FWP229), *snf5Δ* (FWP231), *sol1Δ* (FWP245), *tfg3Δ* (FWP246), *snf59Δ* (FWP247), *snf30Δ* (FWP248), *arp42Δ* (FWP239), *arp9Δ* (FWP240), *arp42Δ arp9Δ* (FWP249), *rsc1Δ* (FWP234) and *rsc4Δ* (FWP242) strains were grown in liquid YES medium to stationary phase, subjected to ten-fold serial dilutions and spotted onto solid YES medium containing caffeine, formamide, hydroxyurea (HU), thiabendazole (TBZ) or benomyl as indicated (see Methods for concentrations). The leftmost spot for each came from a culture at  $1 \times 10^8$  cells  $\text{ml}^{-1}$ , and approximately 3  $\mu\text{l}$  were spotted. (b) Cell elongation, chromosomal

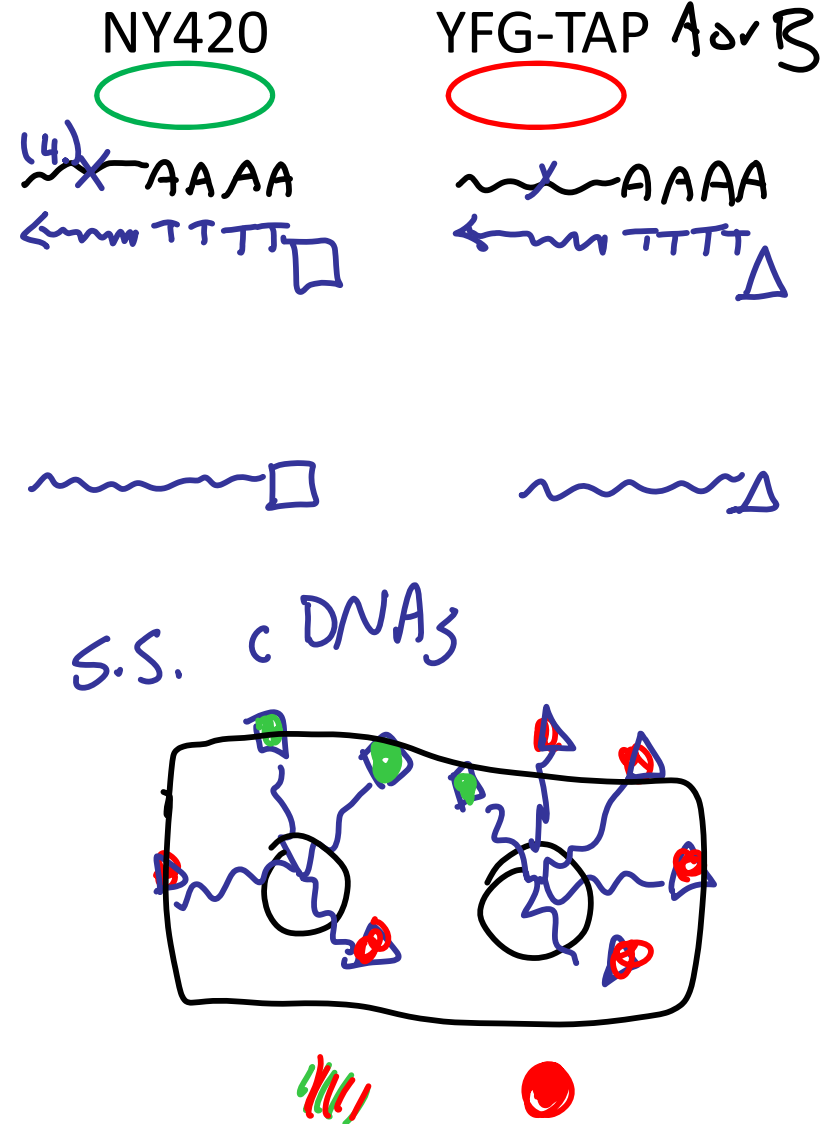
- Additional comment from me
  - In the final report, don't contradict yourself! If growth phenotype suggests TAP-TRP incorporation, and Western doesn't, find an explanation consistent with both results.

# Announcements

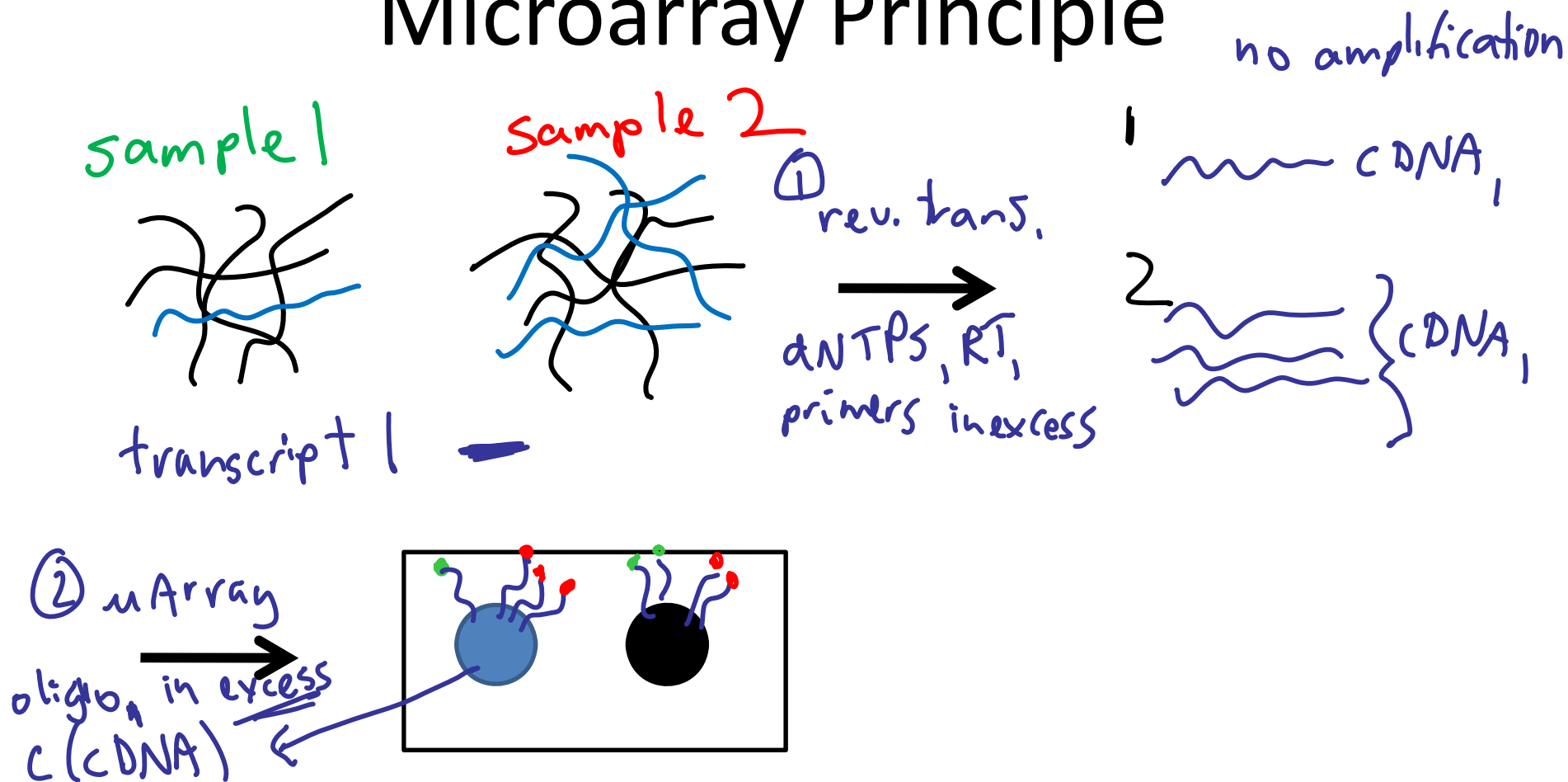
- BE seminar this week is Prof. Han, u/nfluidics
- Office hours
  - Natalie (in lab) Sunday 3-5 pm, Monday 2-4 pm
  - Agi (in 16-336) Tue 4-6 pm, other by appt.
- Brief discussion of mid-term feedback

# Microarray Workflow

1. Isolate RNA from yeast
2. Anneal primers to RNA  
(w/capture sequences)  
*80°C, 10'*
3. Extension step w/RTase  
*42°C, 1.5 hrs*
4. Destroy RNA  $\rightarrow$  NaOH
5. Hybridize cDNAs to array
6. Wash, add fluorophores  
*7 wash, scan*




# Microarray Principle



Intensity of fluorescence at a particular spot is proportional to the original amount of the relevant mRNA in that sample. Next time you'll analyze 44,000 green and red intensity values!

yeast ~6K ORFs

# Microarray Analysis

Analysis	What/why/how	Typical #'s
Background Subtraction	 <p>intensity of b/w spot areas reflects noise</p> <p><math>I - BI</math> sometimes <math>&lt; 0</math> → change it = <math>BI</math></p> <p>Excel =IF(<math>I &lt; 0</math>, <math>BI</math>, <math>I</math>)</p>	<p>bkgrnd. <math>\sim 50-100</math></p> <p>gene spots <math>\sim 10^2 - 10^5</math></p>
Normalization	<p><math>R_I = K G_I</math></p> <p>Cy 5 less incorporation, less stable (but higher quantum yield than Cy3)</p>	<p><math>&lt; 2</math>-fold</p>
Log2 Transform ratio of $G_I; R_I$	<p>"readability" "help (LOA)"</p>	

# Today in Lab

→ b/w 1 & 5 mg RNA

note: red vial = Cy 3 = green/red  
blue vial = Cy 5 = red/far-red  
\*need to know which is which

- Set up RT-PCR
  - With RNase-free technique!
- Meanwhile practice uArray analysis
- Hybridize cDNA to uArray slides
- FNT is recommended, but optional