

RNA-seq vs. qPCR

RNA-seq:

- Measures every expressed gene
- You *enrich* for mRNA
And get rid of rRNA
- Requires sequencing

qPCR:

- Measures single gene
- No mRNA enrichment
Uses primers to amplify GOI
- Does not require sequencing

Q1:

Why are you more likely to observe sequence from the 3' end of a gene in RNA-seq data (relative to sequence from the 5' end)?

Why are you more likely to observe sequence from the 3' end of a gene in RNA-seq data?

- What is special about the 3'-end?

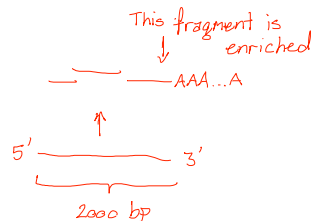
polyA tail 5' ——— AAA...A 3'

- What is special about mRNA?

It has a p(A) tail

- How do you enrich for mRNA?

Uses oligo(dT)'s



Q2:

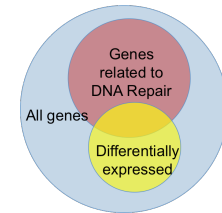
To compare two sets of RNA-seq data, you first normalize the results by calculating the RPKM value for each gene. What are the two factors to which you normalize (hint: how do you normalize between experiments AND how do you normalize between genes)?

Calculating the RPKM

- RPKM = Reads Per Kilobase Million
from RNA-seq exp.
 $\frac{\text{Total reads} / 1,000,000 = \text{per million (PM) scaling factor}}{\text{Reads} / \text{PM} = \text{RPM}}$
 $\frac{\text{RPM} / \text{gene length in Kb} = \text{RPKM}}$

Q3:

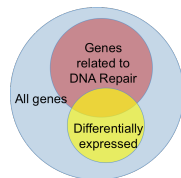
When analyzing RNA-seq data you identify a group of differentially expressed genes (yellow circle). You already know which genes are involved in DNA repair (red circle).w



Which probability distribution will tell you the probability of overlap?

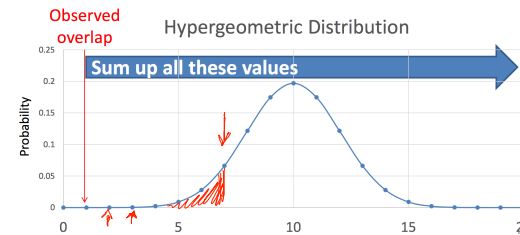
- Hypergeometric distribution

$$P(\text{overlap}) = \frac{\binom{\text{DNA repair Overlap}}{\text{Genome - DNA repair Diff. expr. - overlap}}}{\binom{\text{Genome Diff. expr.}}{\text{Diff. expr.}}}$$



What statistical function can you use to test if the overlap is significant?

- Cumulative density function (CDF)
- Fisher's Exact Test



Q4:
qPCR is used to measure expression levels of specific genes.

qPCR is used to measure expression levels of specific genes

Why measure p21?	Why measure GAPDH?
<i>Has to do w/ cell cycle.</i>	<i>Housekeeping gene</i>
<i>Stalls cell cycle in response to DNA damage</i>	<i>Normalize p21 expression</i>

Q5:
Briefly describe “synthetic lethality” and how it applies to your cell viability experiment

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What is synthetic lethality?	How does it apply to our experiment?
<i>The combination of two mutations kills the cell. The individual mutations do not.</i>	<i>Our BRCA -/- mutant is defective in HR. We treat with a drug that inhibits NHEJ. We want to see if the combined knockout of both pathways leads to cell death when we cause DNA damage (etoposide)</i>

M3D1: Grow phage-based active (cathode) material

1. Purify M13 bacteriophage (phage)
2. Prelab during 60min incubation
3. Finish M13 purification and measure concentration
4. Incubate phage with gold nanoparticles (AuNP)

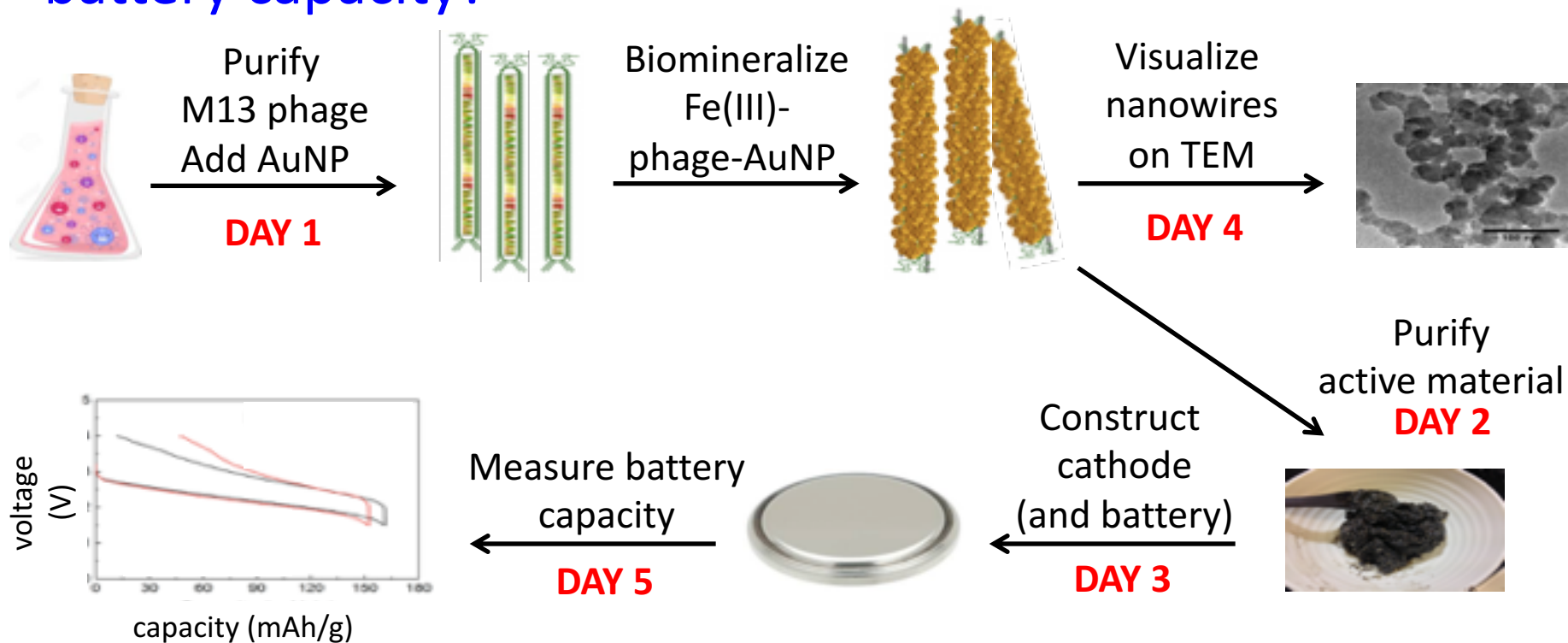
4/19/18



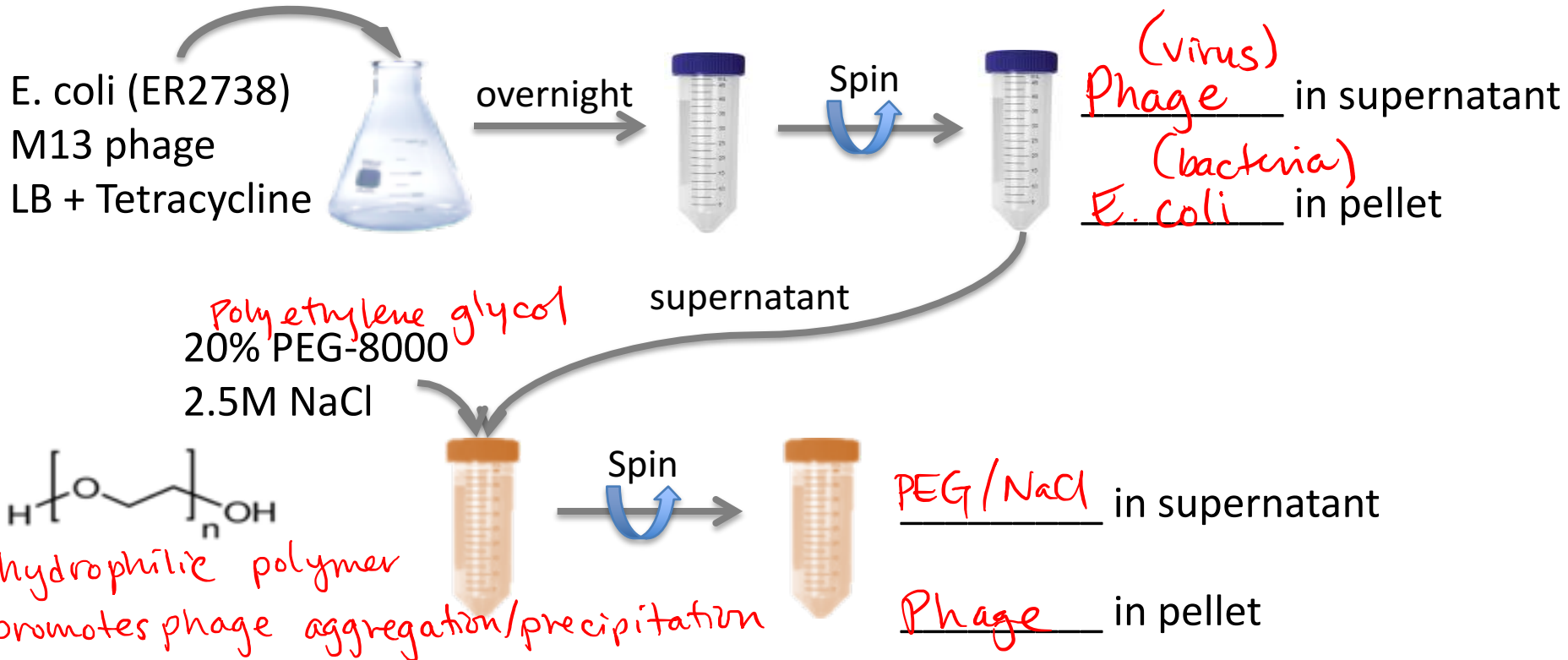
Thank you, Jifa Q. (Belcher Laboratory)!

Module 3: biomaterials engineering

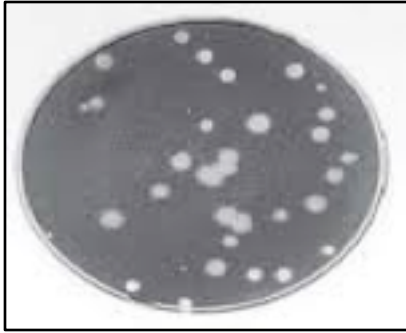
How do material choice and nanoparticle size affect battery capacity?



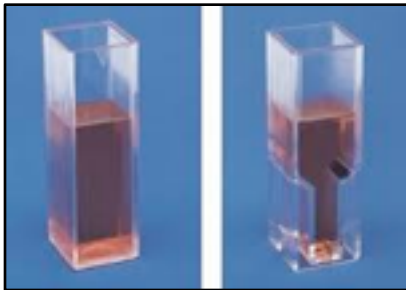
Phage purification using polyethylene glycol (PEG) in 2.5M NaCl



Determining phage titer (number of virus):



- By plating: plaque assay
 - Phage slows *E. coli* growth = plaque (cleared zone)
 - Plaque-forming units: PFU/mL



- By spectrophotometry

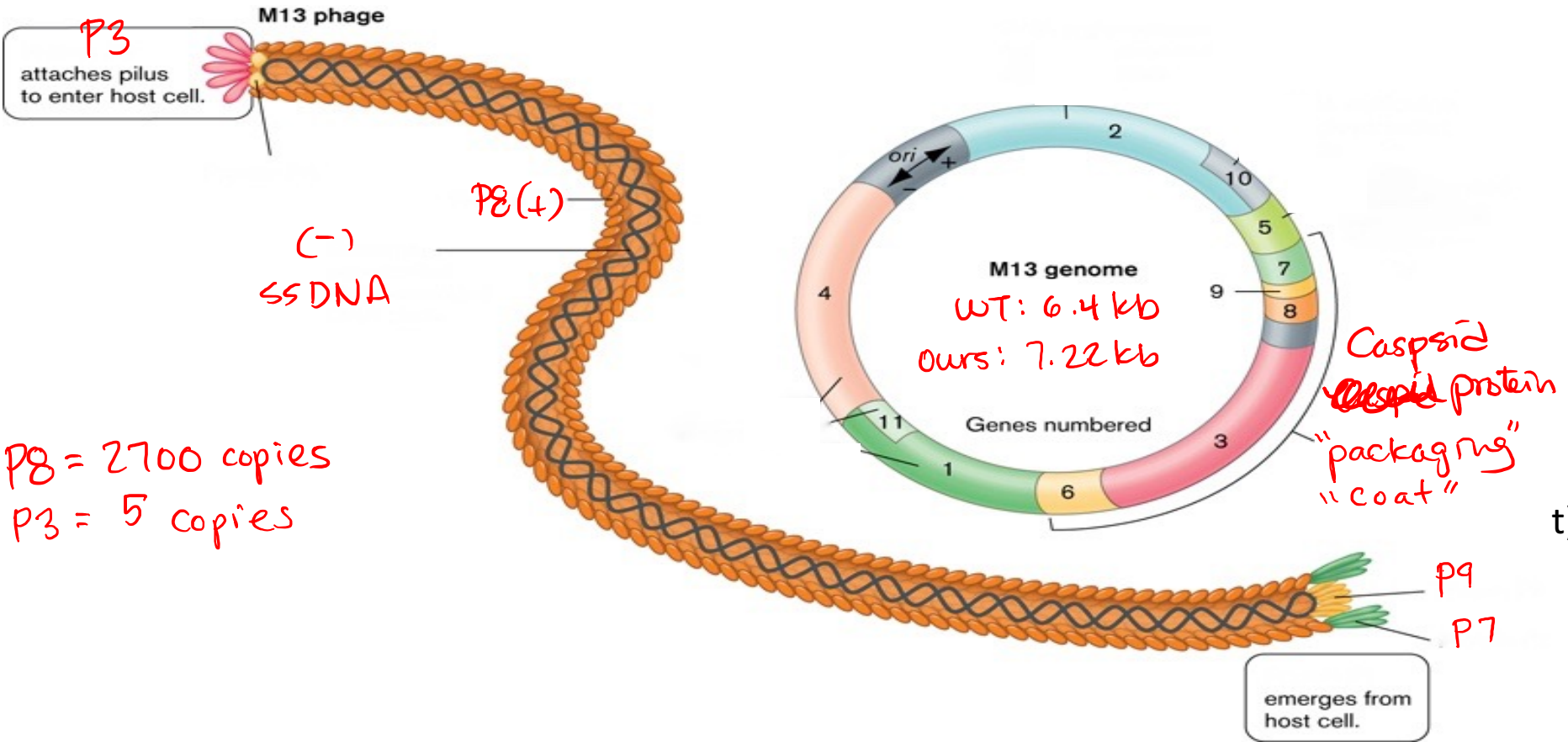
$$\# \text{ phage / mL} = \frac{(6 \times 10^{16}) (A_{269} - A_{320})}{\# \text{ bases in phage genome}}$$

- ❖ Don't forget to consider dilution in this calculation
- ❖ Quartz cuvettes are expensive!

M13 is a high aspect ratio phage

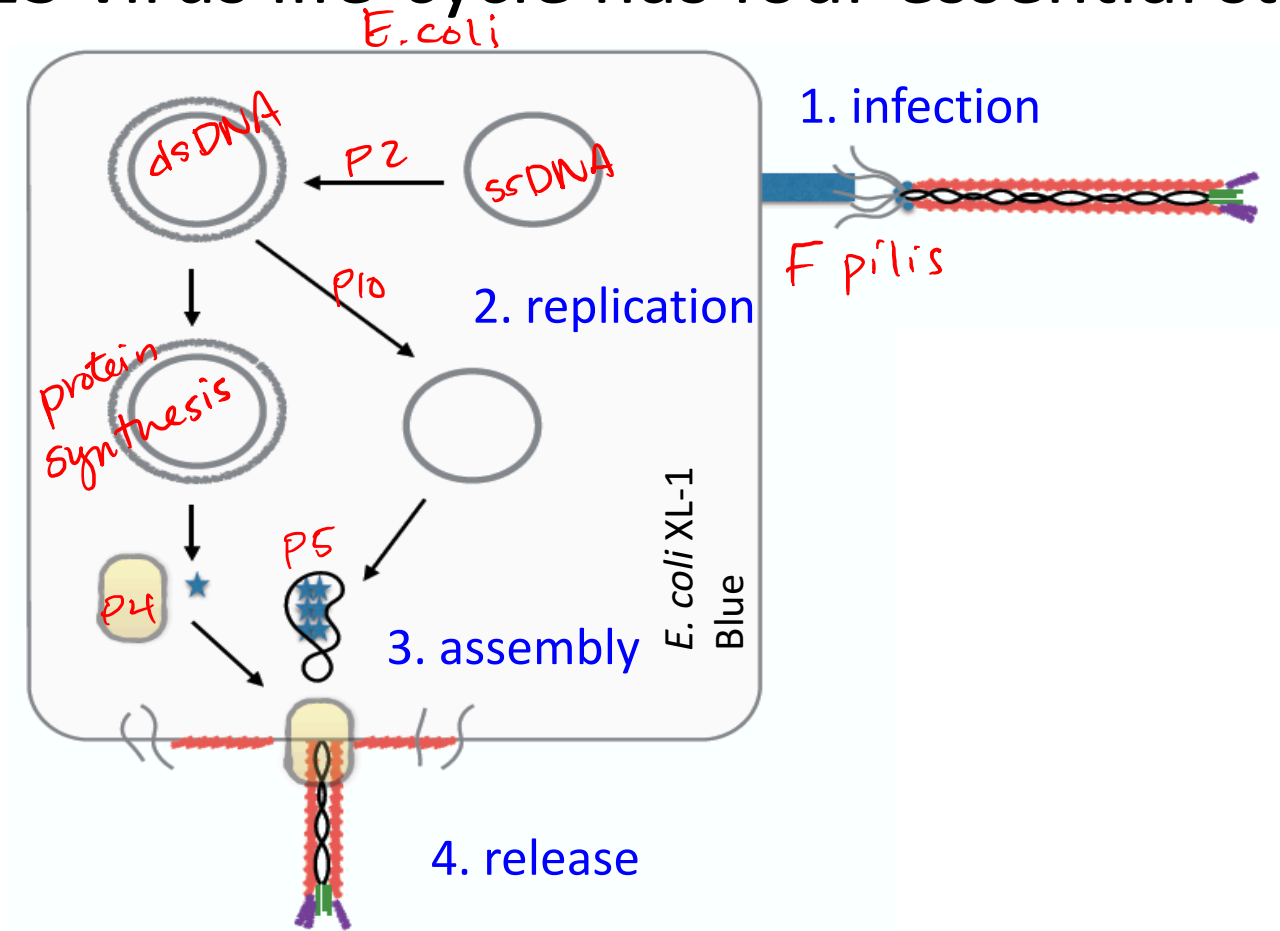
990 nm long
6 nm wide

coated in proteins encoded by ssDNA loop



P8 = 2700 copies
P3 = 5 copies

M13 virus life-cycle has four essential steps

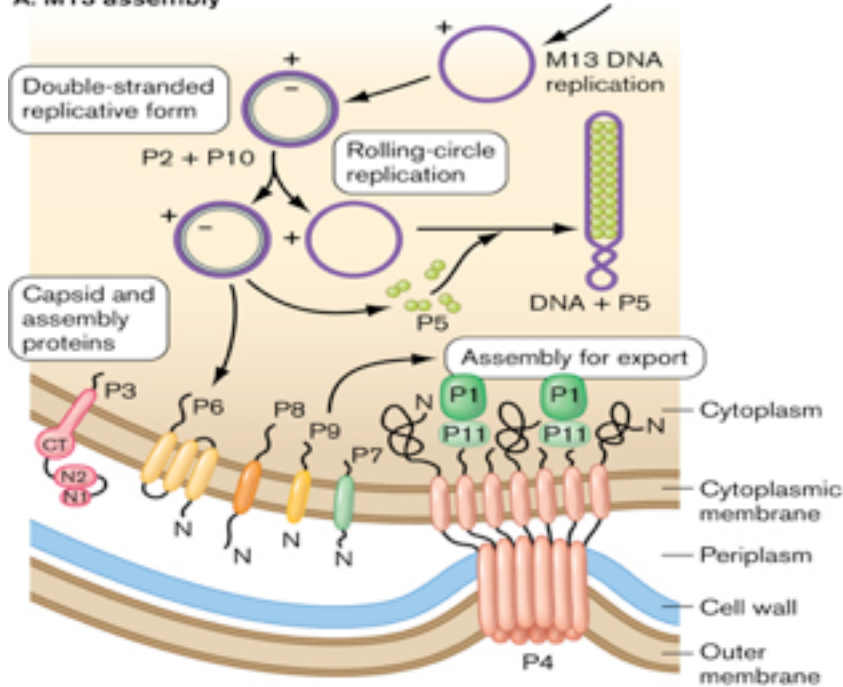


M13 is a nonlytic bacteriophage

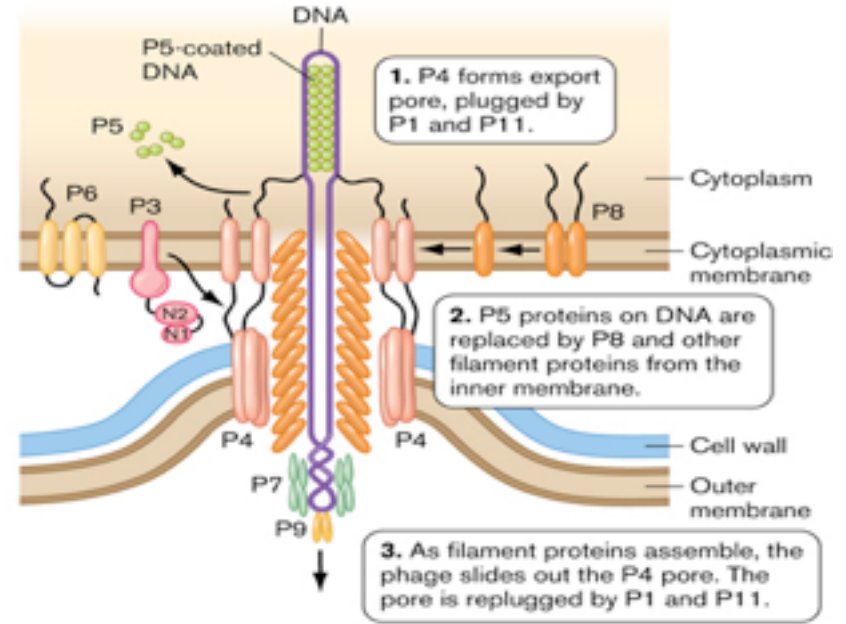
(so we can easily get lots of it)

*Zoomed in
view of
escape*

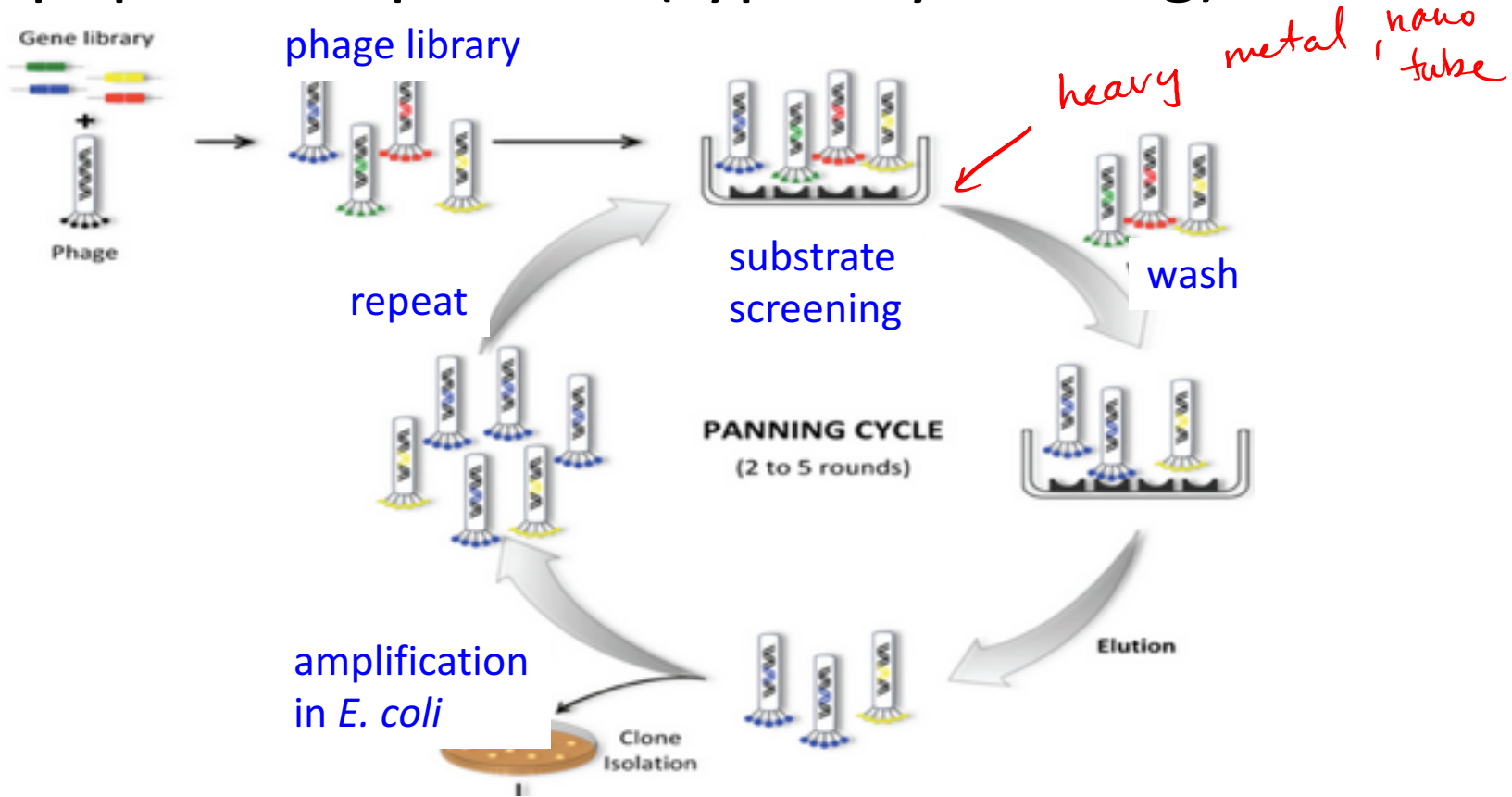
A. M13 assembly



B. M13 export



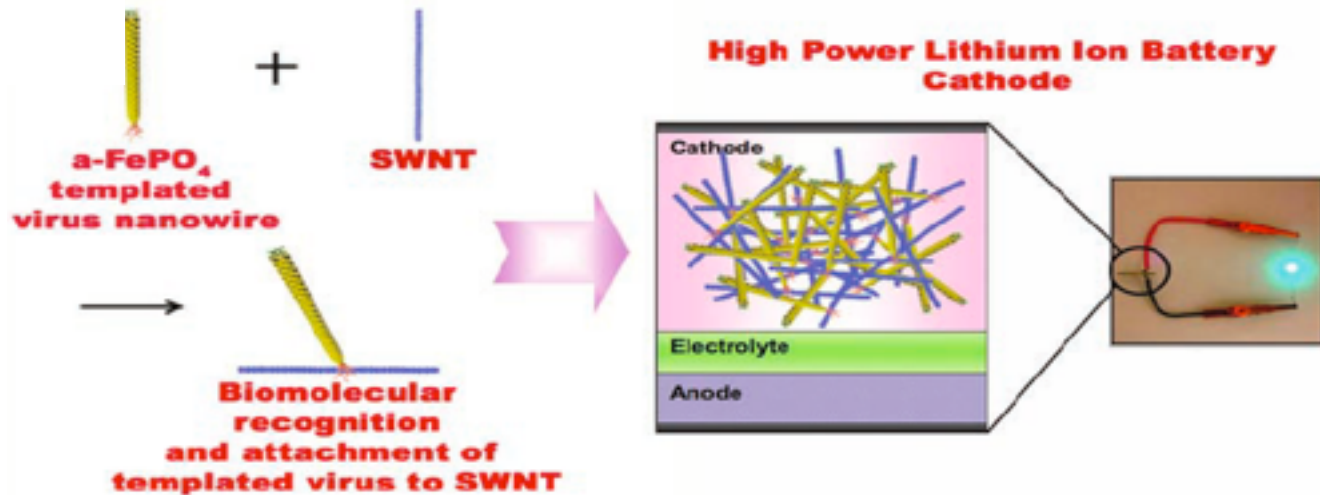
Phage display allows unbiased selection of useful peptide sequences (typically binding)



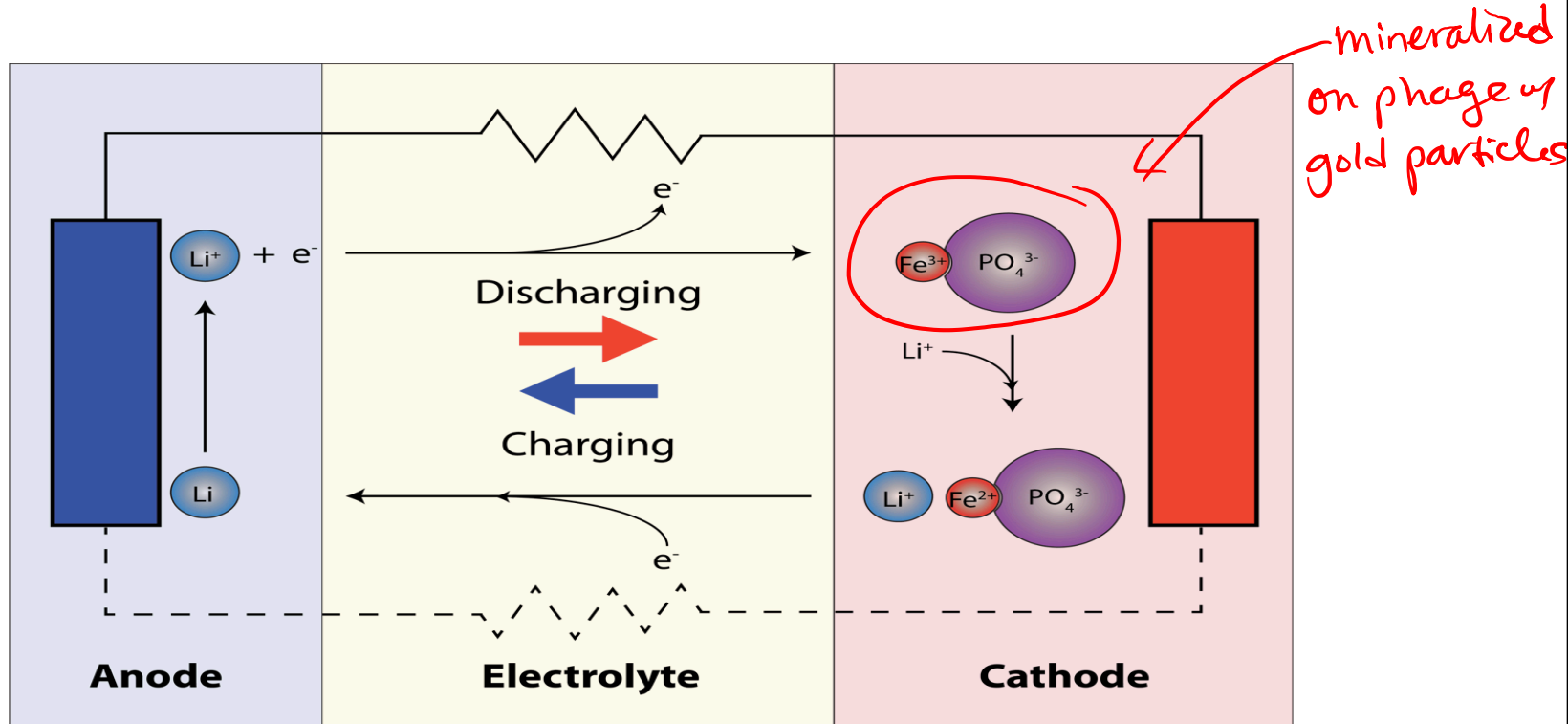
M13 are engineer-able biomaterials

negatively charged

- Our p8 coat protein was mutated to contain sequence **DSPHTELP**
- Modified p8 proteins bind single wall carbon nanotubes (SWCNT), iron, gold, and other cationic metals
- Example of this virus in literature (Science, 2009):



M13 nanowires as battery cathode



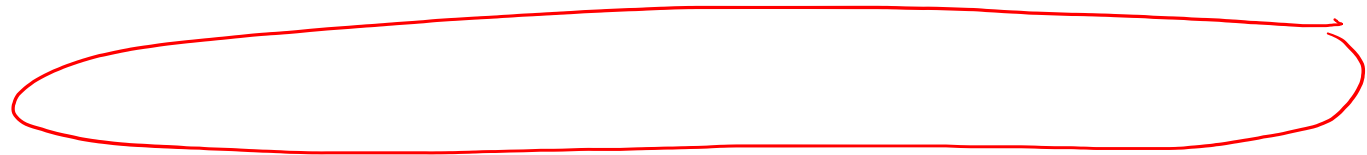
You will make a “Gold Standard” battery and an **experimental** battery

- Gold standard: 3.8nm Gold, 40 AuNP per phage
- Choice of combination: Keep total volume less than 50ml

- 3.8nm Gold
- 5nm Gold
- 9nm Gold

concentrations on wiki

990nm



Considerations for experimental battery: nanoparticle material and size

- Conductivity
 - Au is conductive, how much total gold will be in your cathode?
- Internal battery reaction catalysis
 - Li⁺ in solution → Li⁺ embedded
 - lithium embedded in iron phosphate
 - gold may catalyze this interaction *"facilitates"*
 - Surface area to volume ratio ~~✗~~
 - consider surface area of each nanoparticle
 - consider NP binding phage reduces iron phosphate binding sites

Design with your lab partner. What is your **hypothesis**?

You will make two flasks—one for each battery

Gold standard



- 4×10^{13} Phage
- 40(3.8nm) AuNPs/phage
- Water (final volume 50 mL)

Experimental



- 4×10^{13} Phage
- ? AuNPs
- Water (final volume 50 mL)

Today in lab

1. Finish phage purification
2. Calculate phage number
3. Mix components: phage, AuNP, FePO₄ nanowires (2 flasks, one per battery)

M3D2 HW: Describe **FIVE** recent findings that could potentially define an interesting research question.

- Formally cite the finding
- Write 3-5 sentences summarizing the finding