

- Announcements, Review HW
- Lab Quiz
- Pre-lab Lecture
 - ❖ Your Colony Results
 - ❖ Tissue Culture
 - ❖ Safety + Technical Tips

Announcements, old HW

- OH will be in [REDACTED] (Su 7:30-9, Tu 4-5 pm)
- #1 – ^{units} fine, #2 – careful with backbone size
- Methods are much improved! → structure/story
 - Explain controls (briefly), don't just list them * show your *
e.g. single-enzyme digests, no ligase rxn. logic
 - Carefully consider the purpose of each gel
 - first gel → isolate DNA fragments for cloning
 - verify that enzymes work
 - * infer that double-digest worked, not directly see

Interpreting Your Ligation Results

single-cut plasmid,
uncut

uncut
plasmid

Group Colour	pCX-EGFP (#)	bkb + lig (#)	bkb + ins, no lig (#)	bkb + ins, lig 1 (#)	bkb + ins, lig 2 (#)
Hypothetical Data	1000	0	2	100	100
Yellow	[redacted]	[redacted]	[redacted]	[redacted]	[redacted]
Green	[redacted]	[redacted]	[redacted]	[redacted]	[redacted]
Blue	[redacted]	[redacted]	[redacted]	[redacted]	[redacted]
Pink	[redacted]	[redacted]	[redacted]	[redacted]	[redacted]
Purple	[redacted]	[redacted]	[redacted]	[redacted]	[redacted]

Consider...

- Why did only the positive control work for many groups?
- What does the *no ligase* vs. the *no insert* sample control for? Which one do you expect to have more colonies? 2 > 3
think of 1-2 reasons
- How big is the variation for identical samples, and what are the possible sources of error causing the variation?

Tissue Culture (TC) Environment

- What will “feel” physiological to a cell?

Temp = 37°C

pH = 7.2-7.4 \longleftrightarrow CO₂ = 5%

ambient O₂ levels

Salts → (cells can burst or shriveled)

humidity

• something to stick to

* sterility

Tissue Culture (TC) Medium

- What do cells need to survive?

food : glucose = C-source = energy
glutamine = alt. energy source

rxns. building blocks, cofactors { essential amino acids → (cells don't make them, varies by cell type)
vitamins, minerals, lipids

optional: non-essential aa
Na pyruvate = alt. energy

serum = growth factors, cytokines

non-food: antibiotics P/S (10^{-6})
phenol red = pH indicator

Today in Lab

- Set up gels with diagnostic digest samples

uncut, single, double
PCX-WNX
4X
C1 C2 (or C3)

- While the gels run, you will join me for a TC demonstration and practice lab

cell "passaging"
cell counting
STERILE
TECHNIQUE

- At 3:30 pm, you will discuss the paper by Sonoda, et al. with Prof. Bevin

nitrile gloves
eye protection for MSJ)

Announcements, old HW

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- #1 – fine, #2 – careful with backbone size
- Methods are much improved!
 - Explain controls (briefly), don't just list them
 - Carefully consider the purpose of each gel

Interpreting Your Ligation Results

Group Colour	pCX-EGFP (#)	bkb + lig (#)	bkb + ins, no lig (#)	bkb + ins, lig 1 (#)	bkb + ins, lig 2 (#)
Hypothetical Data	1000	0	2	100	100
Yellow	4000	0	0	0	0
Green	1884	1	144	380	656
Blue	2000	0	168	520	1031
Pink	426	0	0	0	0
Purple	300	0	0	0	0

Consider...

- Why did only the positive control work for many groups?
- What does the *no ligase* vs. the *no insert* sample control for? Which one do you expect to have more colonies?
- How big is the variation for identical samples, and what are the possible sources of error causing the variation?