

- Announcements
- Lab Quiz (on M1D1 material)
- Pre-lab Lecture
 - ❖ Writing a methods section
 - ❖ Gel electrophoresis
 - ❖ DNA purification
 - ❖ Today in Lab: M1D2

Announcements

- Brief discussion of orientation day quiz
- Remember, *Assignments + Schedule* = our syllabus
- A few FNTs due on Stellar

MODULE	DAY	DATE	FNT DUE TODAY
		Tue Feb 7	N/A
1	1	Thur Feb 9	Due M1D1
1	2	Tue Feb 14	Due M1D2
1	3	Thu Feb 16	Due M1D3
President's Day	Feb 20	Mon Feb 20	Feb 20

↑
wiki FNT list Stellar →

General
[edit topic](#) - [delete topic](#) - [add assign](#)

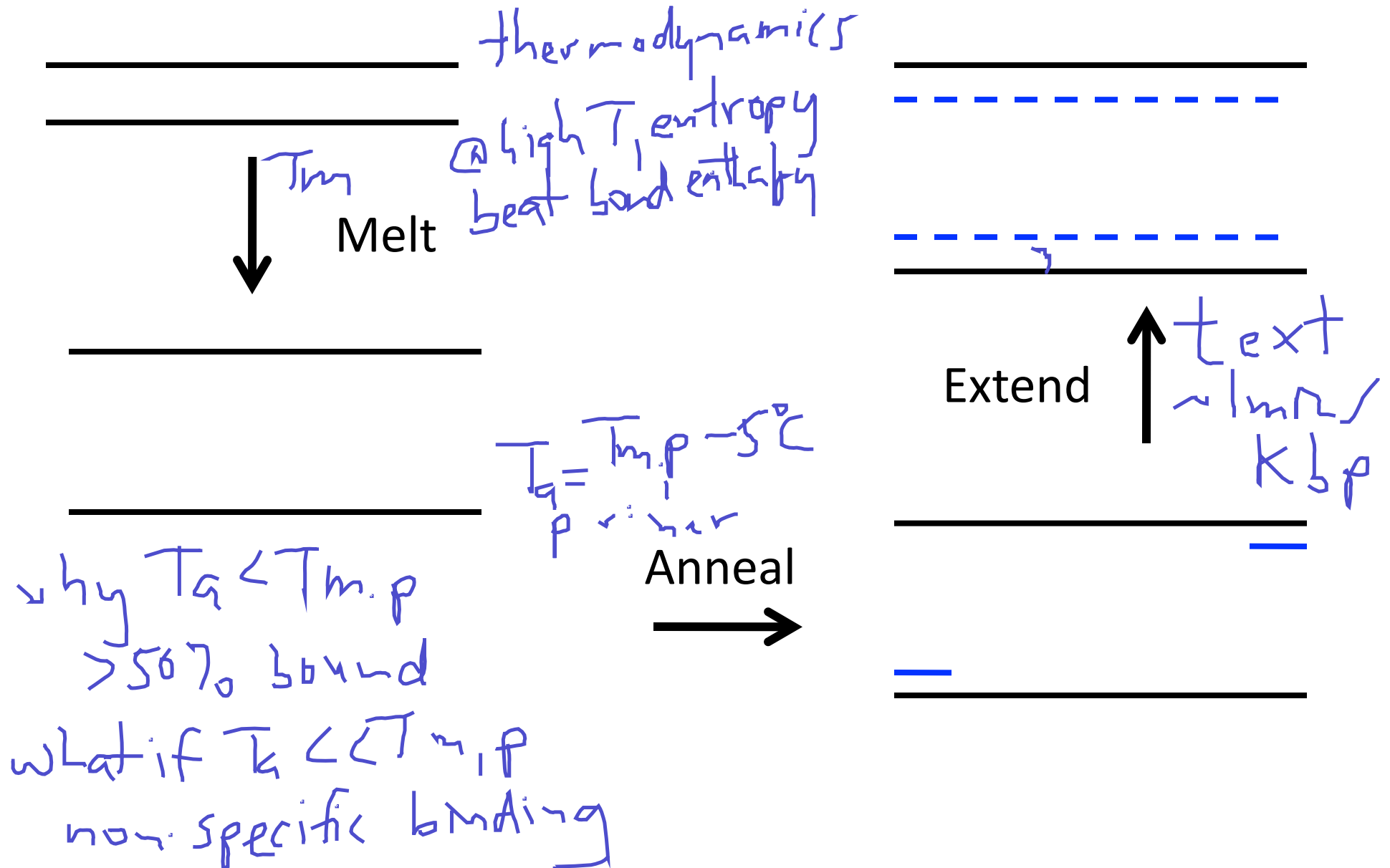
[Due M1D3, T/R](#) [edit](#) - [delete](#)
Due 16 February 2012 1:00 p.m. Post

[Due M1D3, W/F](#) [edit](#) - [delete](#)
Due 17 February 2012 1:00 p.m. Post

[Due M1D4, T/R](#) [edit](#) - [delete](#)
Due 23 February 2012 1:00 p.m. Post

[Due M1D4, W/F](#) [edit](#) - [delete](#)
Due 24 February 2012 1:00 p.m. Post

Quick note: what drives PCR?



Methods section tips

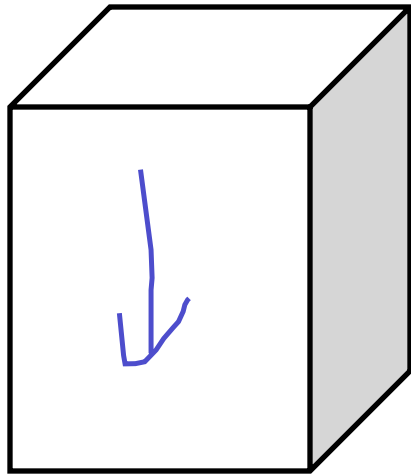
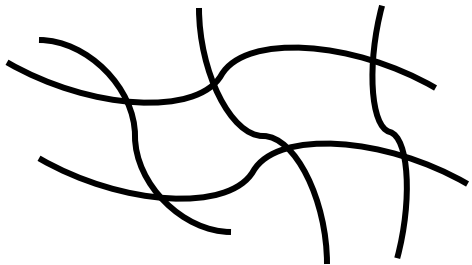
- Organizing sub-sections
 - Often start with an overview/introductory sentence (*what*, not *why*) → then give step-by-step details
- Methods should be concise and complete
 - Space-wise, avoid tables/lists when a sentence will do
 - Sentence-wise, avoid extra words
 - Content-wise, cover what's needed and only that needed to understand and replicate your work
- Concentrations are more useful than volumes; or you can state amounts, plus total volume.

Methods section exercise

- Consider the following passage: “Template DNA (5 ng) and primers were mixed with 20 uL of 2.5X Master Mix in a PCR tube. Water was added to 50 uL. A tube without template was prepared and labeled control.”
 - What information is missing?
 - What information can be cut?

DNA Electrophoresis (EP): Principle

Agarose gel



DNA



Agarose and DNA are both
polymers: Have
molecular entanglements

Driving force for separation: mass: charge

DNA moves - to + because of phosphate groups

Separation is according to: size

smaller

DNA moves faster because

entang. \uparrow size

as wt \uparrow , pore size \downarrow

DNA EP: Visualization

Loading dye: glycerol: sink into wells

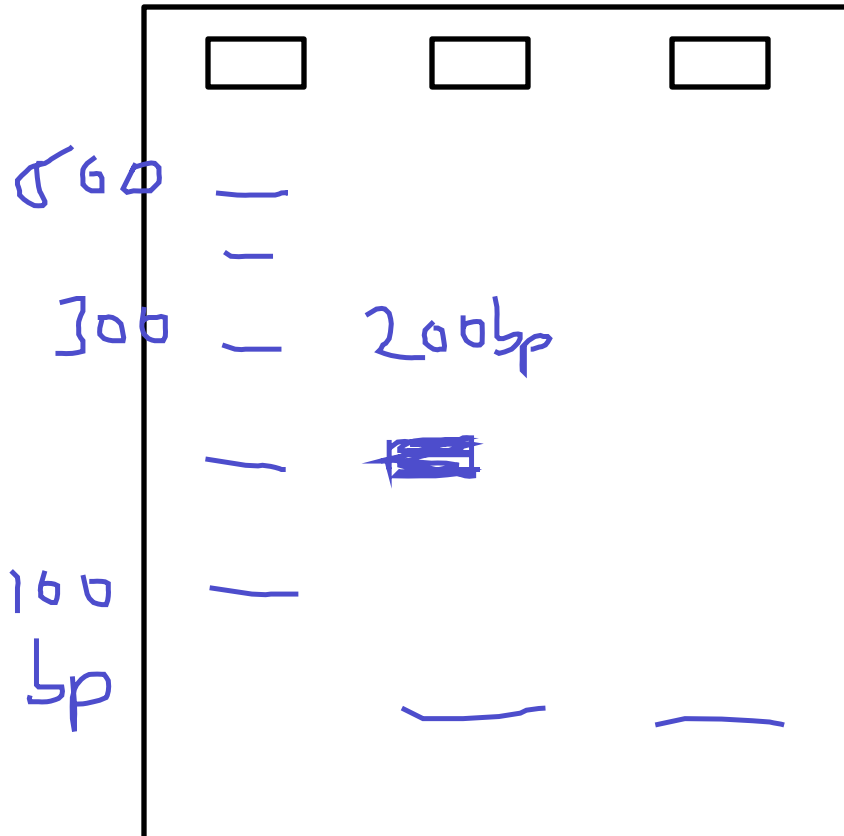
xylene cyanol: visual/real-time
tracking dye

Ethidium bromide
(or SYBR Safe, etc.):

fluoresces under UV/blue
if DNA-bound

DNA EP: Analysis

ladder PCR* control



DNA ladder: known size and conc. standards

Relationship:

distance of $\frac{1}{\log(MW)}$

more details in Mod 2

DNA EP: Clean-up and Safety

- Use **nitrile gloves** when handling DNA gels and all equipment used for gels.
- Gels and gel-contaminated papers are disposed of in solid chemical waste.
- Wear **amber glasses (blue light) or face shields (UV)** when cutting DNA bands out of a gel.

DNA extraction from agarose gel

why? get rid of dNTPs, etc.; switch buffers



1. bind DNA \rightarrow high salt, low pH
chaotropic salts disrupt H-bonds
DNA sticks to column

2. Keep DNA, wash rest
ethanol precipitate

Silica resin
column

3. elute DNA \rightarrow low salt, high pH



\rightarrow O^- - DNA \leftarrow c-salts, repulsion

[qiagen.com]

Note: initial mixture should look yellow, not blue



Today in Lab (M1D2)

- Prepare PCR products for running on gel.
- Gel runs 60 min.
 - Mark your area of the gel box with colored tape.
 - After, learn to photograph and cut out bands.
- Meanwhile, WAC discussion on lab reports.
 - T/R Leslie 2:30-3:15
 - W/F Marilee 2:45-3:30
- Finally, DNA extraction from gel.
- FNT: methods section, read journal article.

prep tubes!