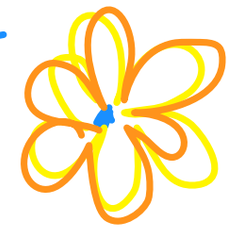
M1D6: Lipofection & Paper Discussion

10/1/13

Lab Treat



1. Pre-lab discussion

2. 1/2 lab in TC to set up HR experiment

   *White, Blue, Yellow & Orange in TC

3. 1/2 lab meet with Prof. Engelward to discuss paper

   *Red, Green, Pink, Purple, Platinum to 56-711

4. Switch!

Thank you Lizzie!!!

Review M1D3 FNT: Figures & Data Interpretation

Appreciate Class-wide view

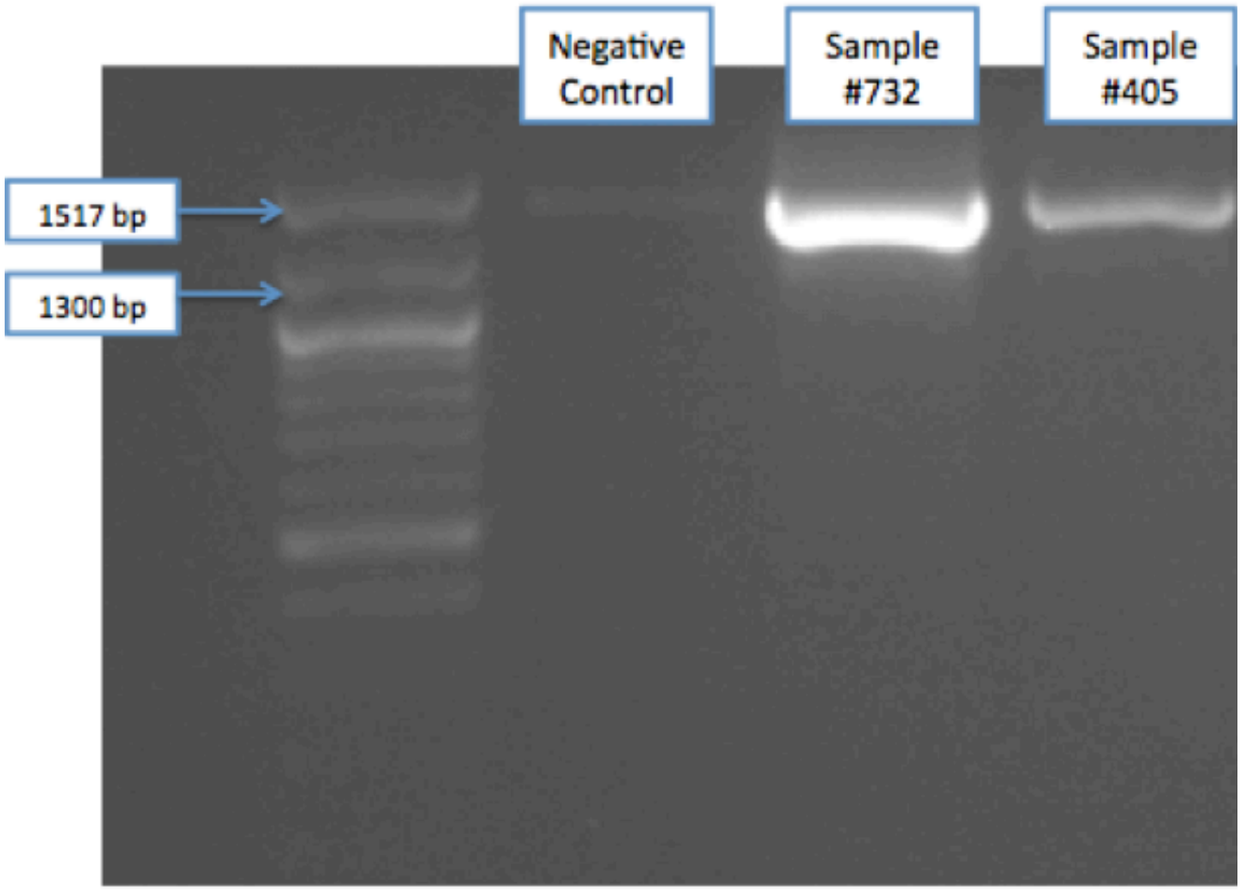


Figure 2: Agarose Gel of 16s rRNA PCR product amplified from gull samples.

- Data Summary – Mod 1 11am – due Thurs, Oct 10 Stellar
- Concise statements (think poster)
  - Options – order of importance
  - figure friendly
- No band in 'Neg Control' suggests ...
- Band @ approx 1400bp suggests positive PCR amp of 16s RNA

Current title: Agarose gel of 16s rRNA PCR product amplified from gull samples.

Review M1D3 FNT: Figures & Data Interpretation

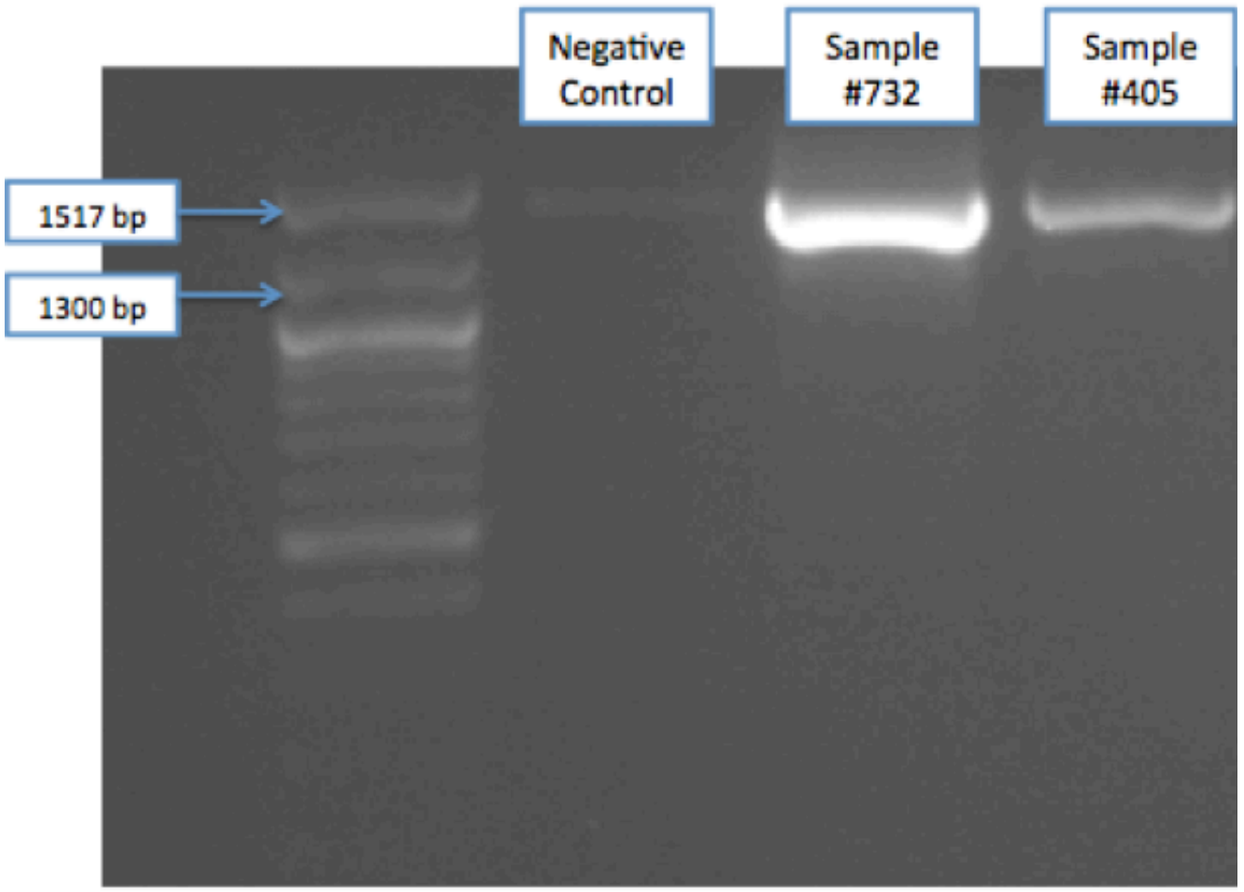


Figure 2: Agarose Gel of 16s rRNA PCR product amplified from gull samples.

Alternate titles:

Amplification of bacterial 16s rRNA gene from Alaskan gull stool samples.

Electrophoresis of amplified bacterial 16s rRNA genes.

Successful amplification of bacterial 16s rRNA gene from two Alaskan gulls.

Revisit Methods section: What experiments fit together?

PCR

Transformation

XbaI/EcoR1 Digest

PCR product purification

Diagnostic Digest GE

XbaI/EcoR1 Digest
Purification (GE)

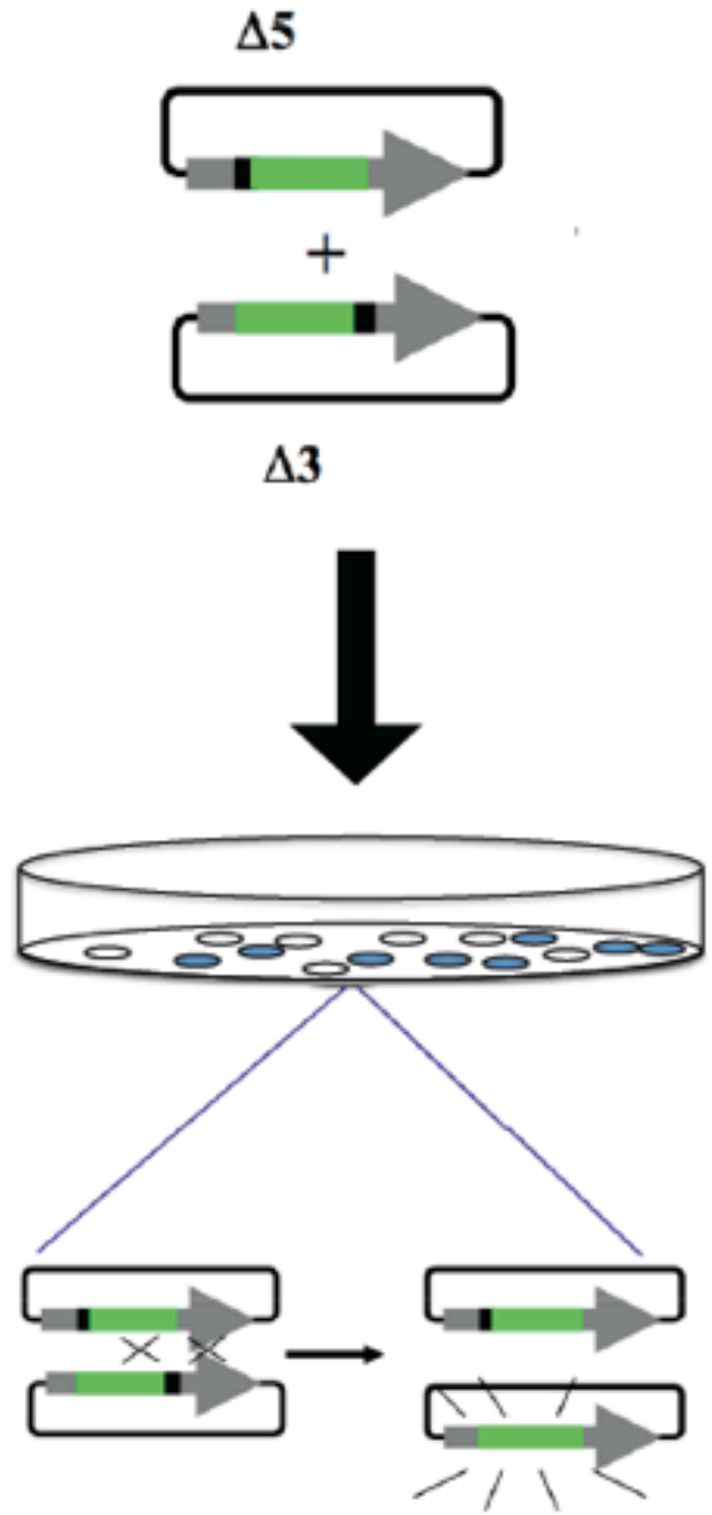
O/N e.coli cultures

Plasmid purification

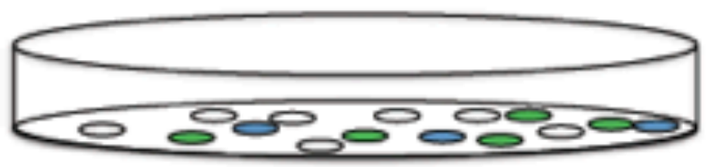
Ligation/Precipitation

Diagnostic Digest

Step 2: Test the system!



Julian Voss-Andreae
Steel Jellyfish (Green Fluorescent Protein), 2006
Stainless steel, 4' x 3' x 3' (1.20 x 0.90 x 0.90 m)
Location: Friday Harbor Laboratories (San Juan Island, WA)



Step 2: Test the system!

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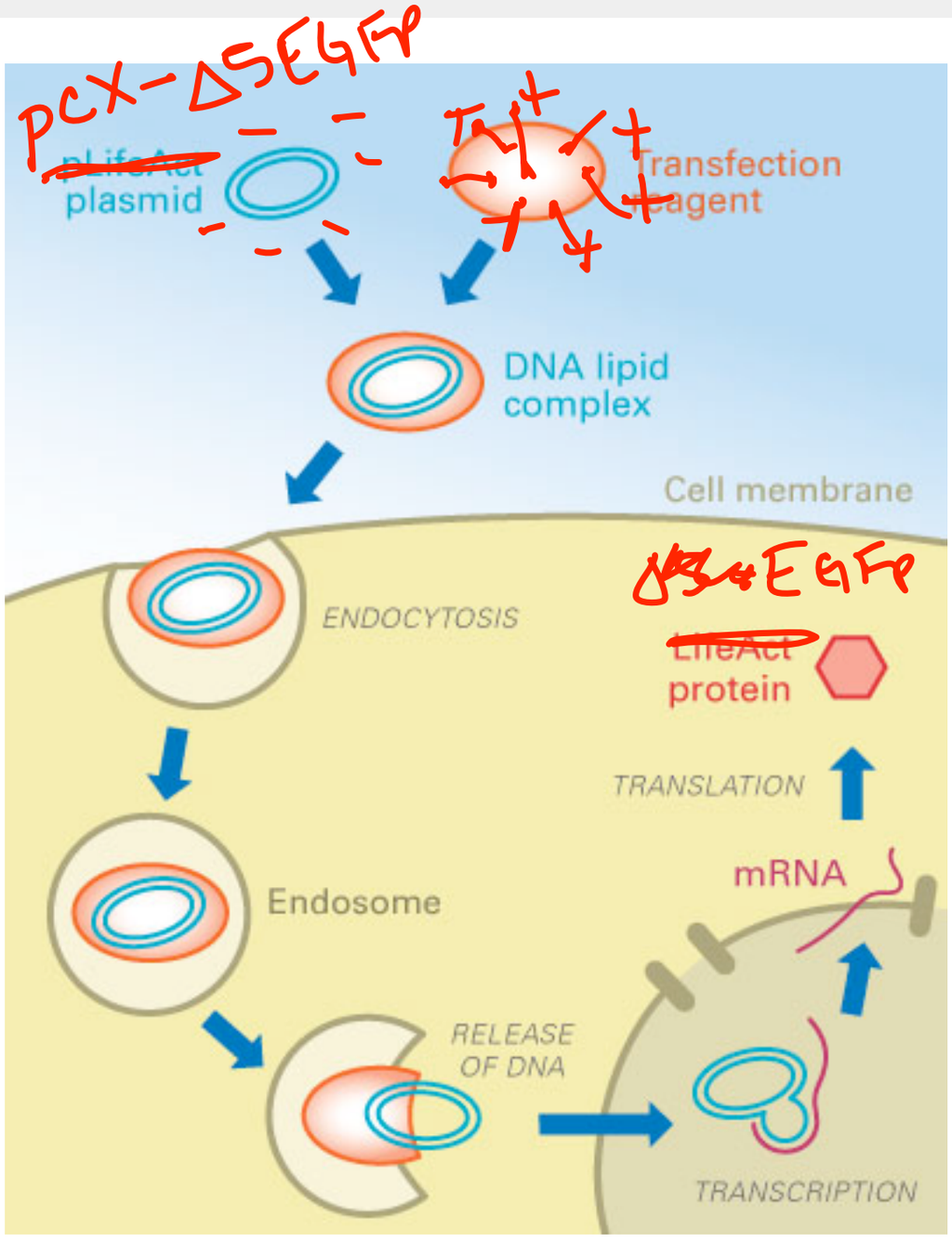
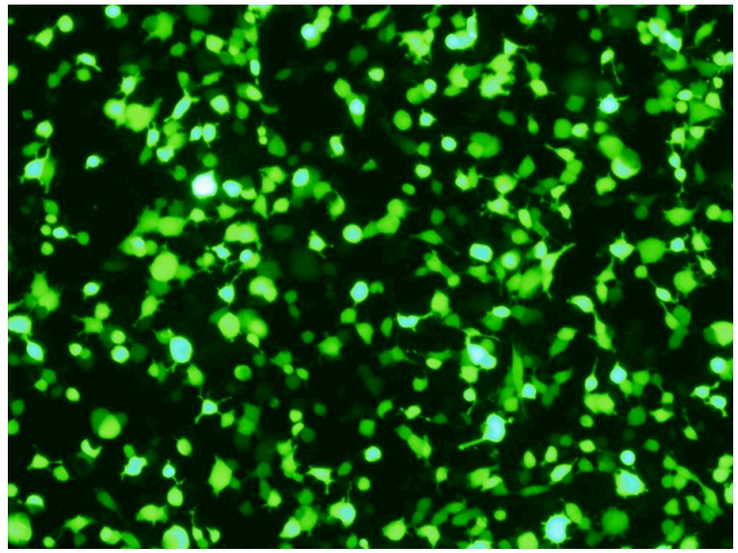
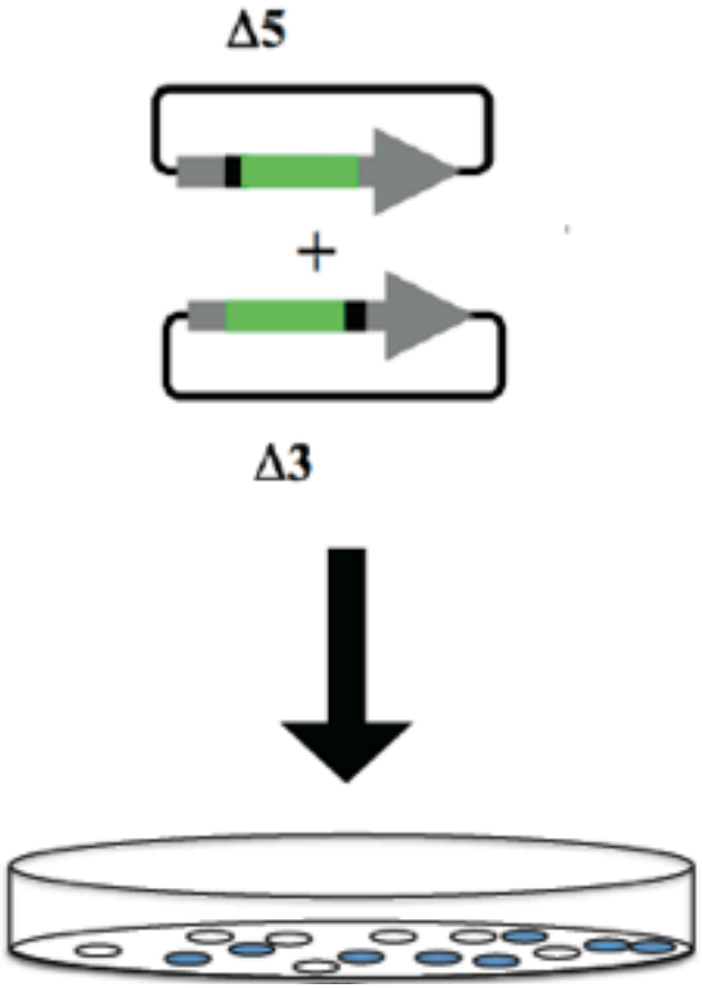
20-YEAR ANNIVERSARY

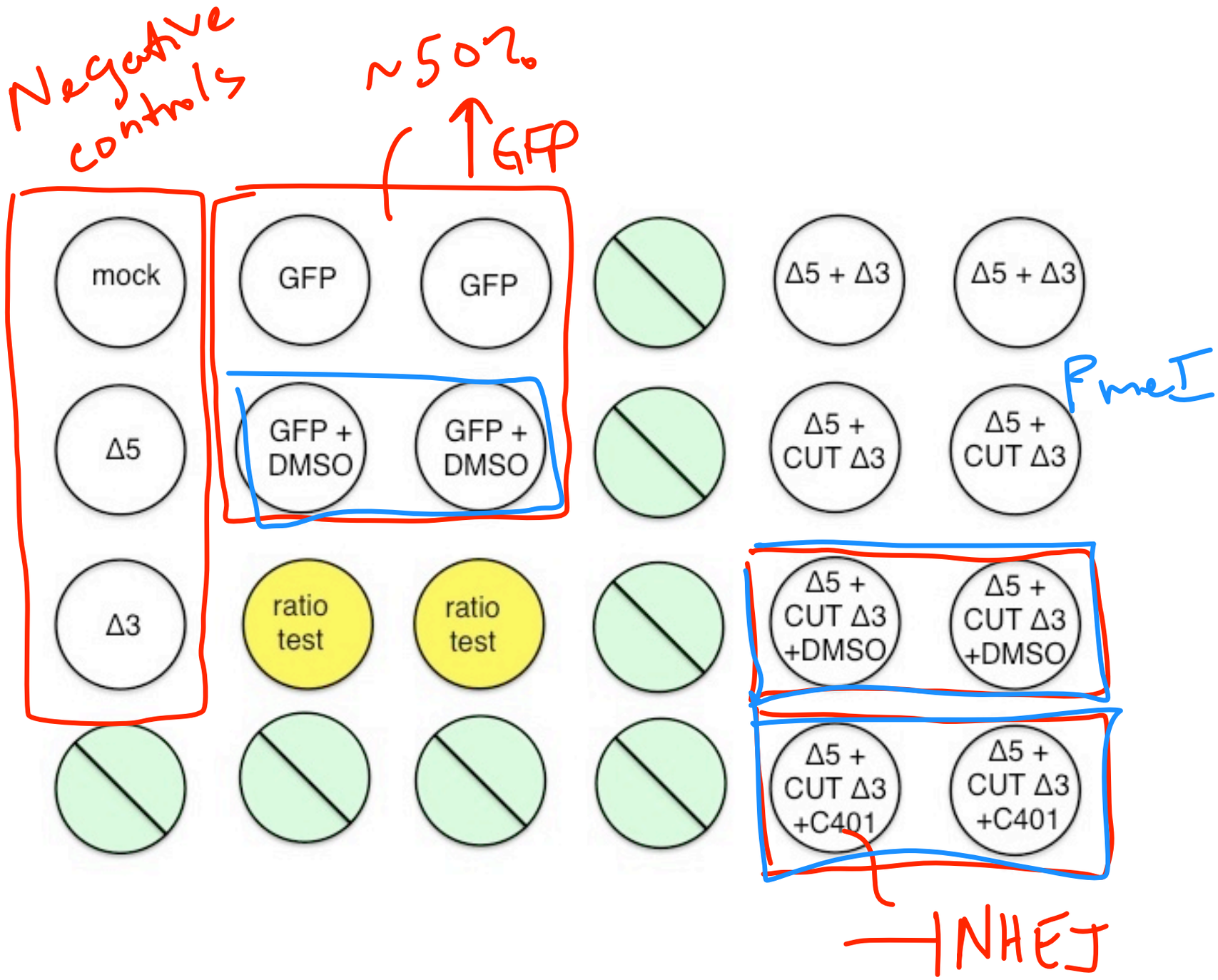




MES ~ max 70%

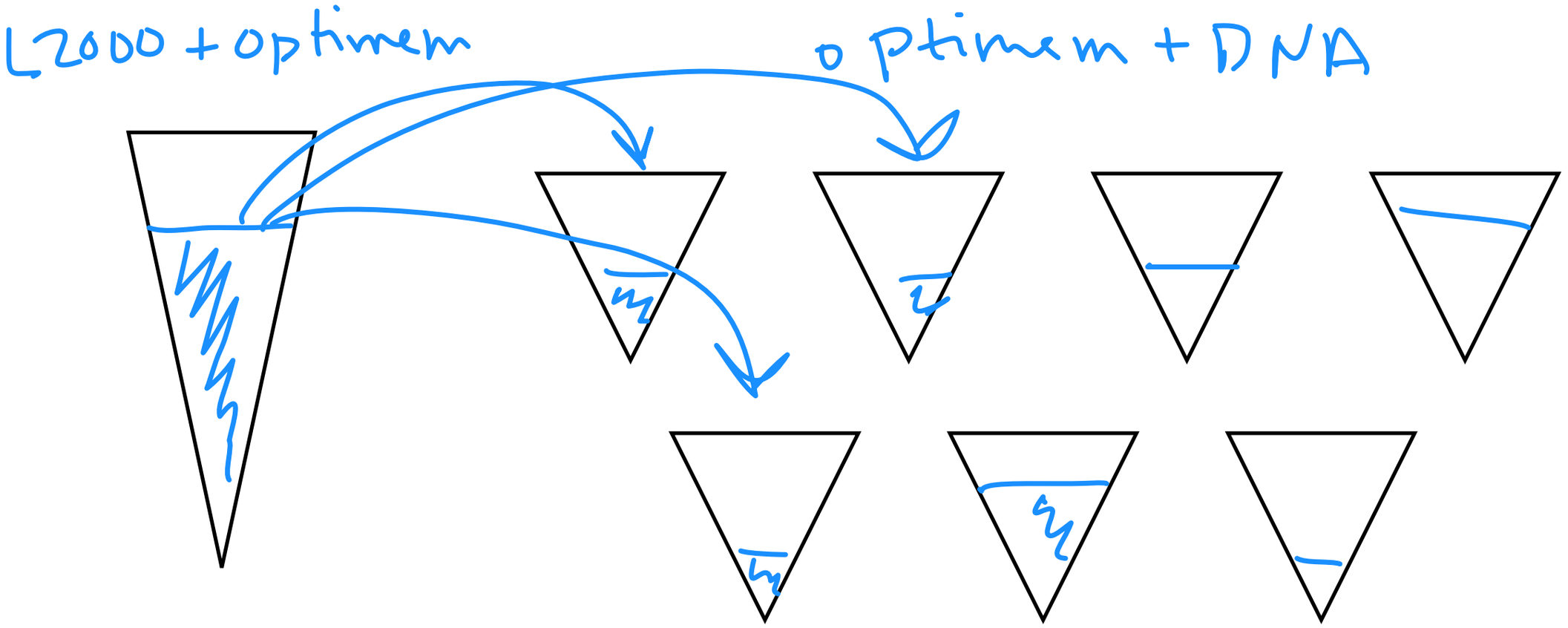
Step 2: Test the system!

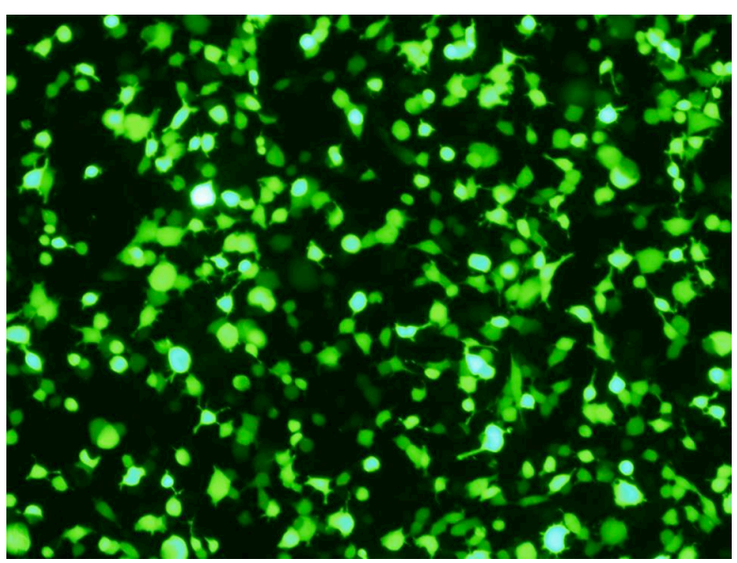


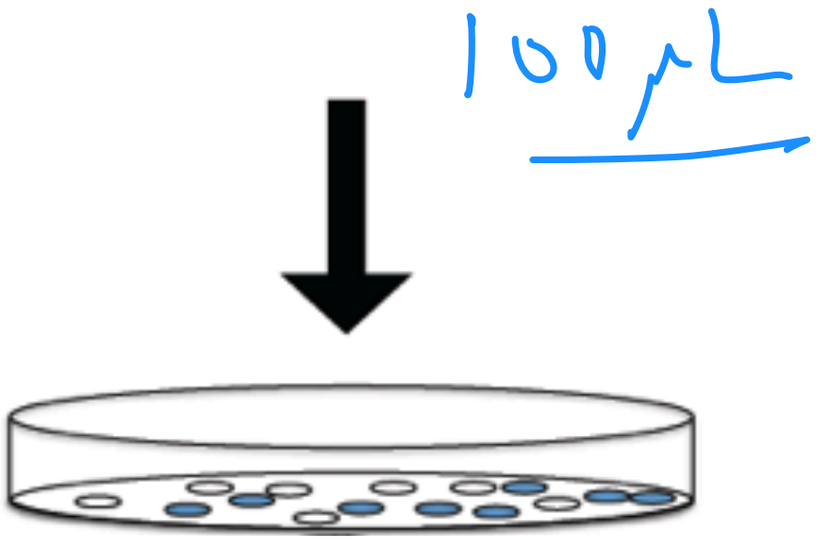



% EGFP Expression
mock
Δ5
Δ3
GFP

Step 2: Test the system!







Tissue culture tips

- Set up a few inches *behind* the barrier/grate
- Minimize opportunities to bump or expose sterile equipment or your samples
  - Uncap bottles *before* opening pipet
  - Keep tips and dishes *closed* when not in use
  - Avoid passing your hands/arms over open dishes
  - Don't try to hold > 2 things at once! ☺
- Take care not to clog the pipet-aids

Today in the lab:

- Set-up lipofections -- pick your bonus condition
- Discuss paper with Prof. Engelward
- Look for feedback on Methods ASAP

Next time in the lab:

- Sign-up on M1D7 Talk Page
- FACS analysis to measure HR efficiency
- Post FACS data on M1D7 Talk page promptly!!!