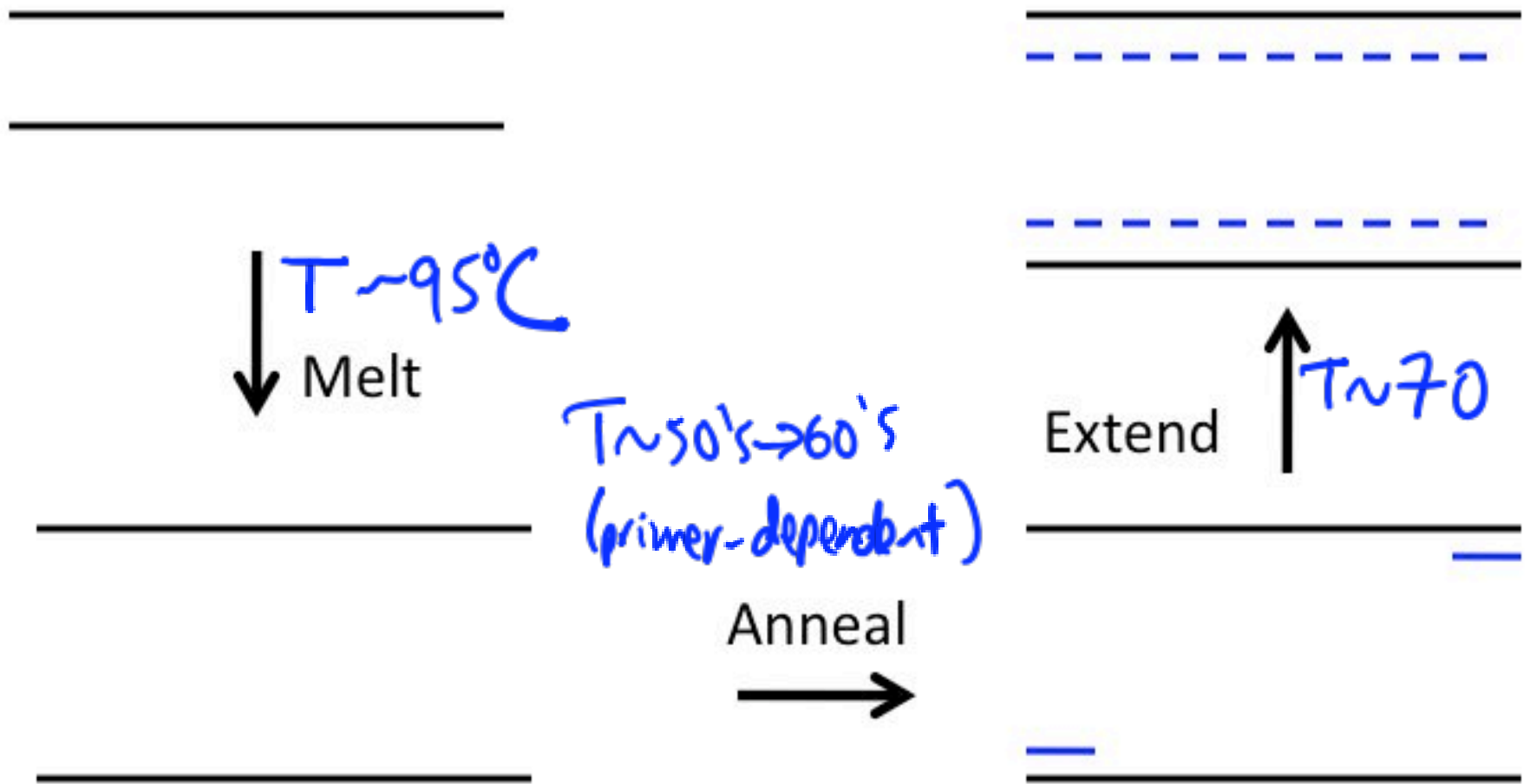


- Announcements
- Quiz
- Pre-lab Lecture
 - ❖ Writing a Methods Section
 - ❖ Gel Electrophoresis
 - ❖ Bacterial Transformation
 - ❖ Today in Lab

Announcements

- Friday: ChemE seminar about biomaterials
 - 66-110, 3 pm
- Next Thursday: BE seminar about biogels
 - 32-141, 4:05 pm

Quick note: what drives PCR?



Methods section tips

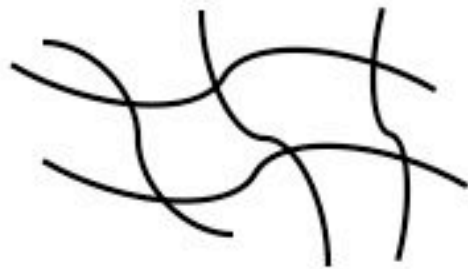
- Organizing sub-sections
Start w/ an overview sentence giving the purpose of that exp.
- Methods should be concise and complete
 - Space-wise, avoid tables/list when a sentence will do
 - Sentence-wise, avoid extra/confusing words
 - Content-wise, cover what's needed and only that needed to understand and replicate your exp.
- Concentrations are more useful than volumes; or you can state amounts, plus total volume

Methods section exercises

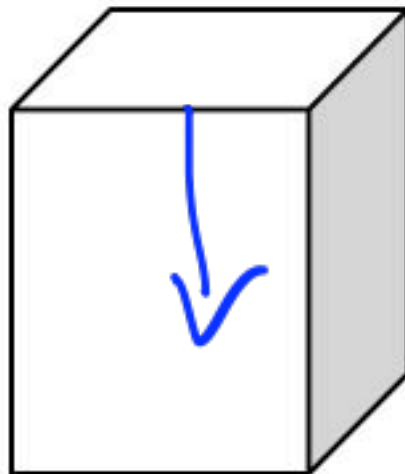
- Which is more readable: “To the Y were added the X” or “The X were added to the Y”?
- How can I more quickly express “1 g of protein in 45 mL of water and 5 mL of 5X buffer B”?
2% protein in (1X) ~~water~~ aqueous buffer B
- Which parts of an SDM are unique to a given experiment, versus standard protocol?
cycles; T annealing; t extension → 1 min / 1 kbp DNA composition / concentration primers, template

DNA Electrophoresis (EP): Principle

Agarose gel



-



+

DNA



Agarose and DNA are both

bio polymers → entanglements (molec.)

Driving force for separation: charge
* mass; charge

DNA moves - to + because of phosphate groups

Separation is according to: size

smaller

DNA moves faster because

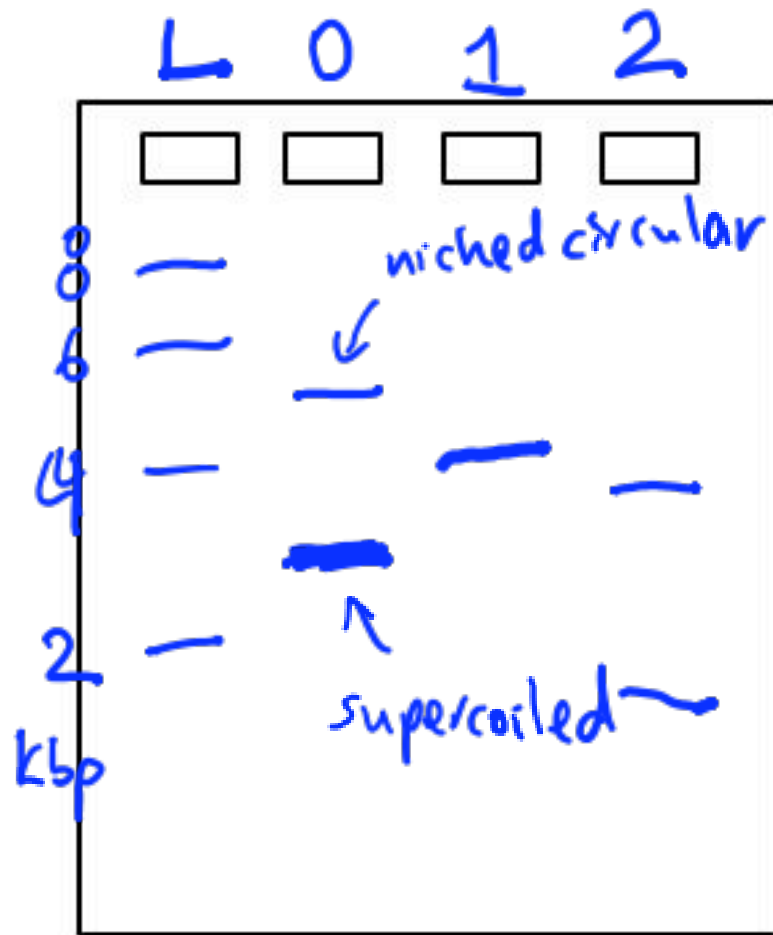
entanglements ↑ w/ size

DNA EP: Visualization

Loading dye: glycerol → sink into wells
xylene cyanol → visual tracking dye

Ethidium bromide: fluoresces under UV,
if bound to DNA

DNA EP: Analysis



DNA ladder: Standards of known size, conc.

Relationship: distance $\propto \frac{1}{\log(MW)}$

Samples:
 uncut plasmid \rightarrow 2 forms
 single-cut \rightarrow 4Kbp - linear
 multi-cut \rightarrow multiple bands

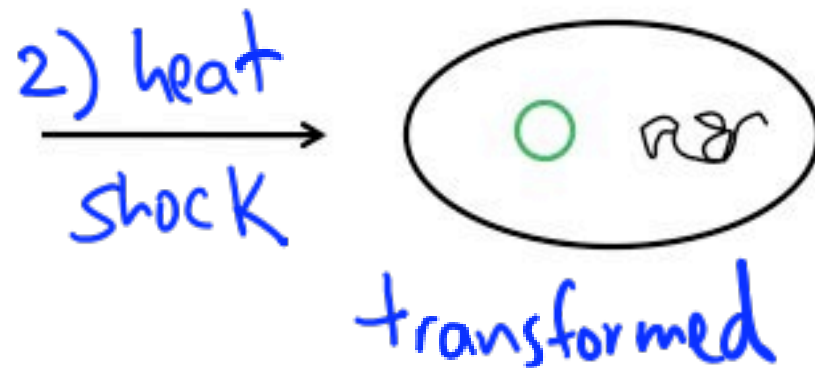
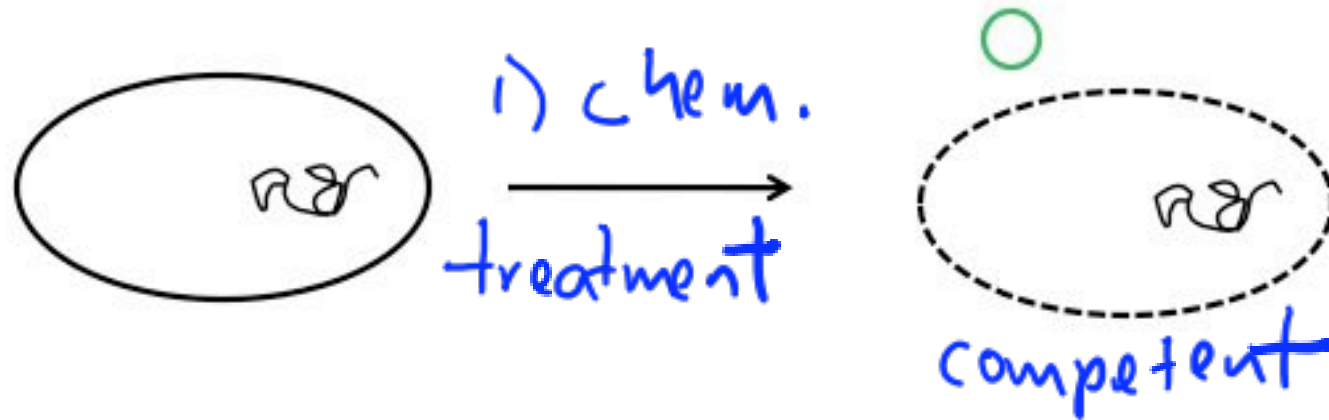
DNA EP: Clean-up and Safety

- Use **nitrile gloves** when handling DNA gels and all equipment used for gels.

EtBr → MUTAGEN

- Gels and gel-contaminated papers are disposed of in solid chemical waste.
- Wear **eye protection/face shields** if cutting DNA bands out of a gel.

Bacterial transformation

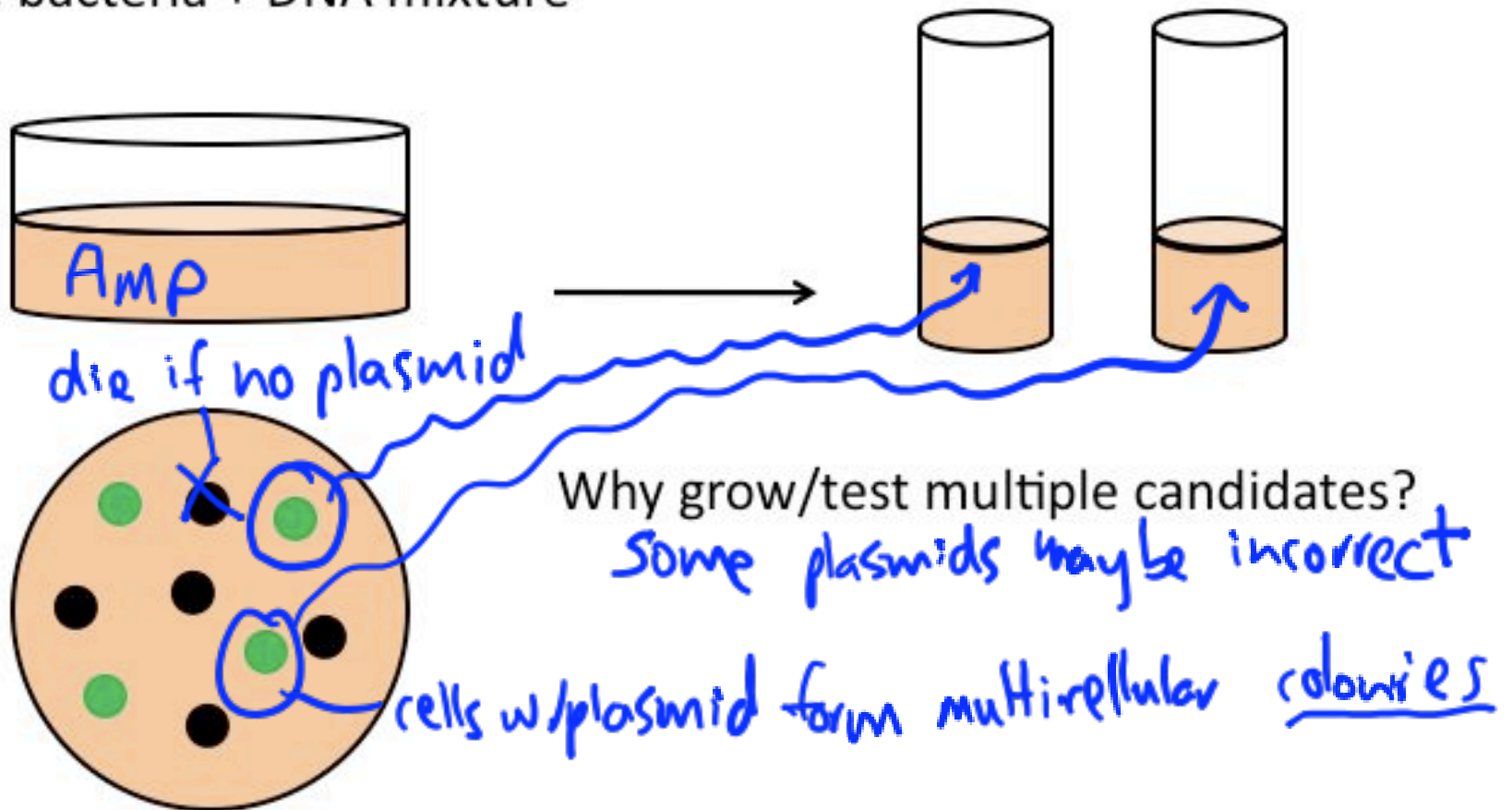


other methods (1-step)

- 1) electroporation
- 2) ballistics

DNA Amplification in Bacteria

Plate bacteria + DNA mixture



Today in Lab

- Set up gel: runs 45 min, we will photograph it.
 - Mark your area with a coloured sticker
 - Bring your USB key up front
 - Update: in BLANK wells, teaching faculty will put concentrated parent plasmid instead (reference)
- Meanwhile, discussion w/Neal and Atissa.
- Finally, bacterial transformation – be gentle!