

# M2D7: Induce CRISPRi system

11/3/16

1. BE Communication workshop: Journal Club
2. Pre-lab
3. Analyze sequencing results
4. Prep for dCas9 induction and mixed-acid fermentation

## Major assignments for M2

- Research Article (20% of grade)
  - due by 5pm on Sun., November 20<sup>th</sup>

### Research Article content

1. Title
2. Abstract
3. Introduction
4. Materials and Methods
5. Figures and Results
6. Discussion
7. References

- Blog post for M1 due by 10pm on Mon., Nov. 21st

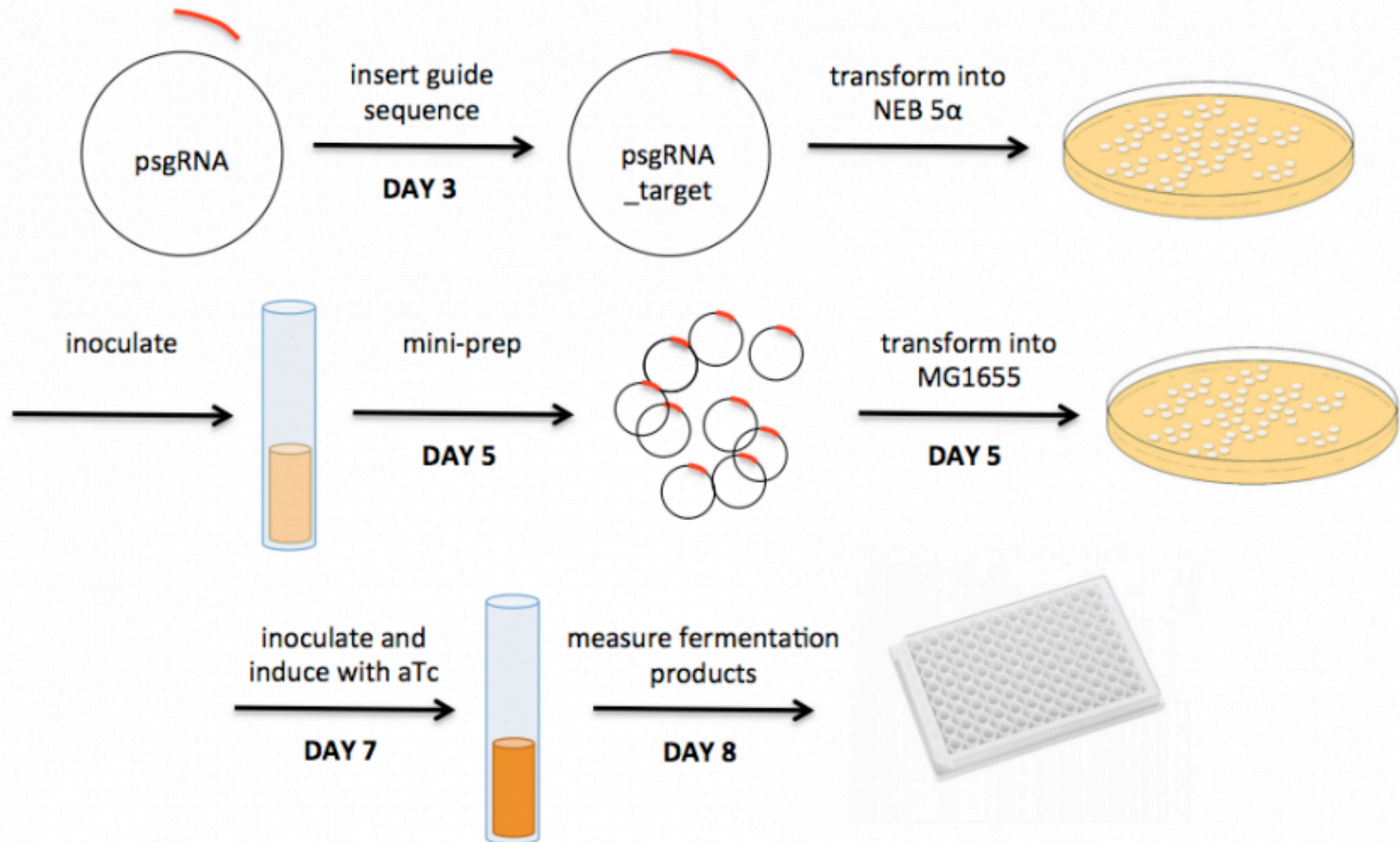
## Extra office hours

- Saturday Nov. 19<sup>th</sup>  
10am-5pm

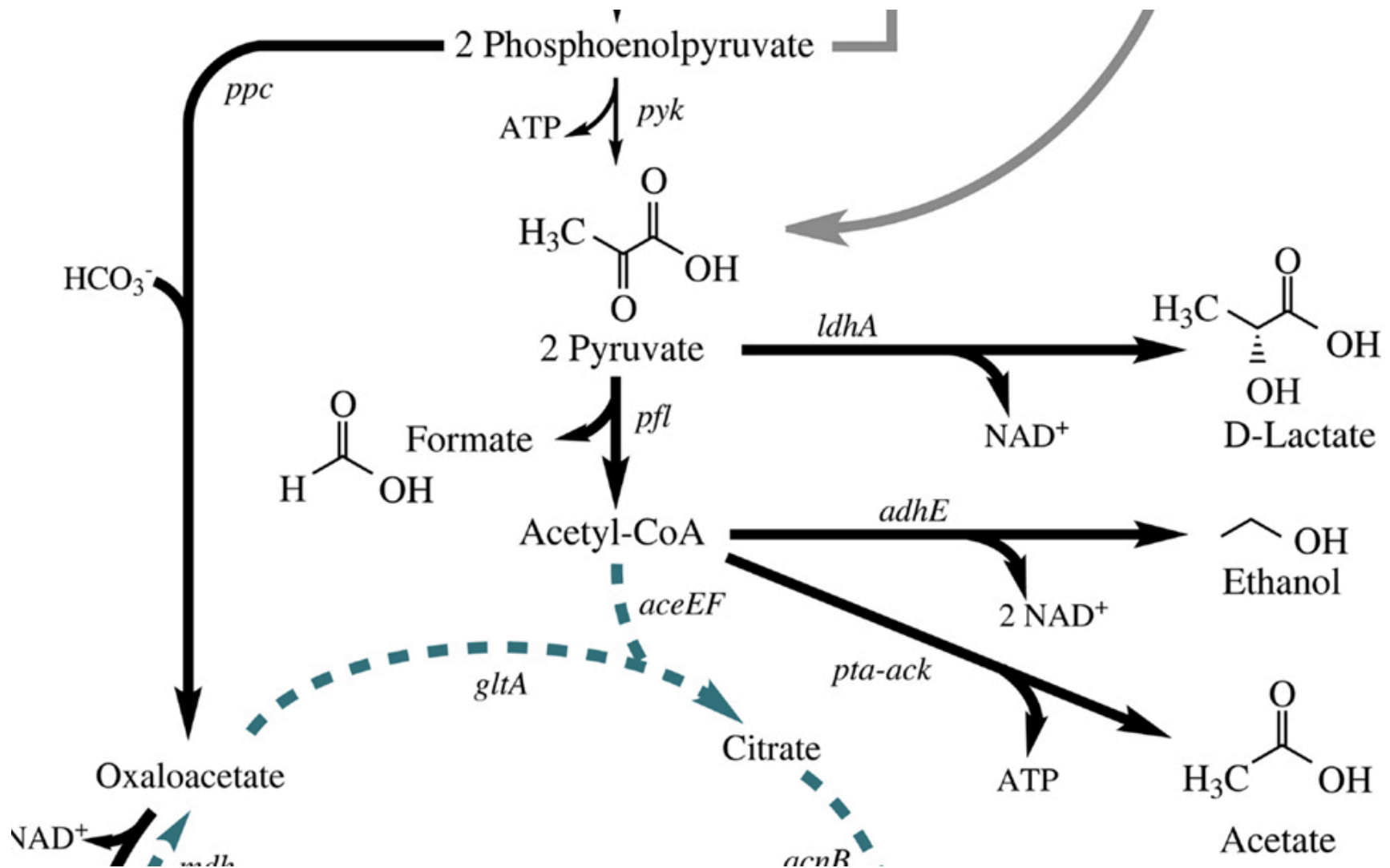
## Regular office hours

- Noreen: Monday 1pm and 5pm in 16-317
- Leslie: Monday 4pm, Wednesday 9am in 16-429b
- Maxine: Monday 2pm, Friday 9am in 16-239

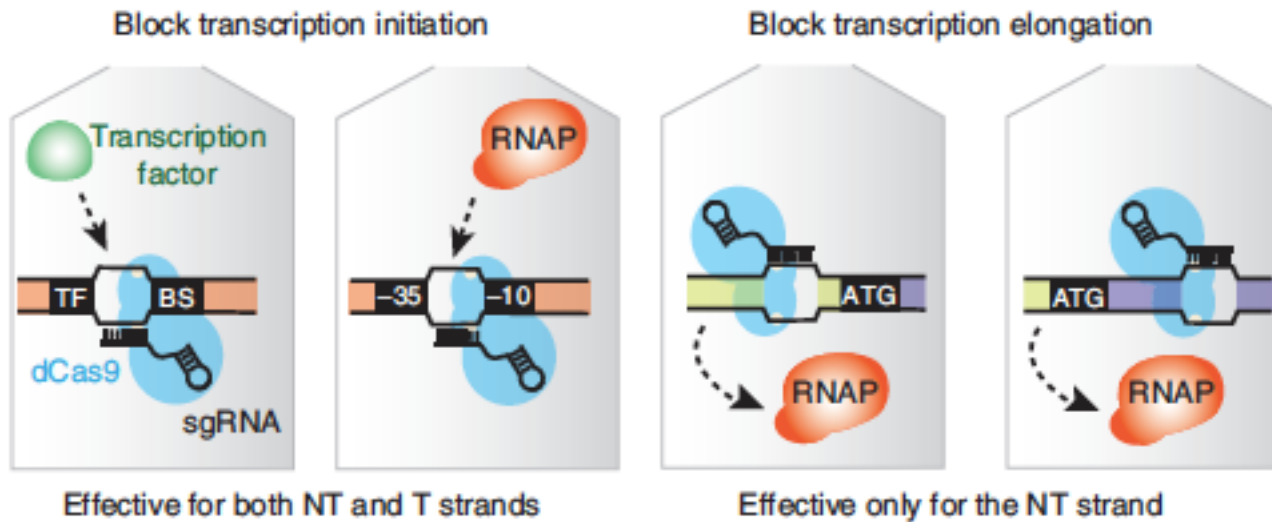
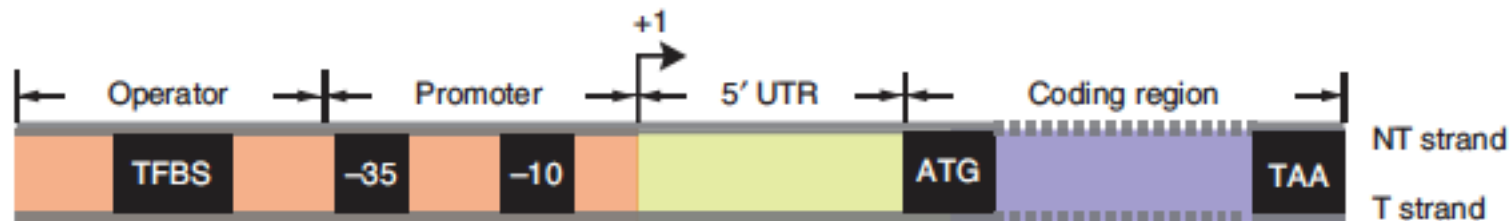
# M2 experimental overview



# Using CRISPRi manipulate the *E. coli* fermentation pathway



# Design of gRNA for CRISPRi system



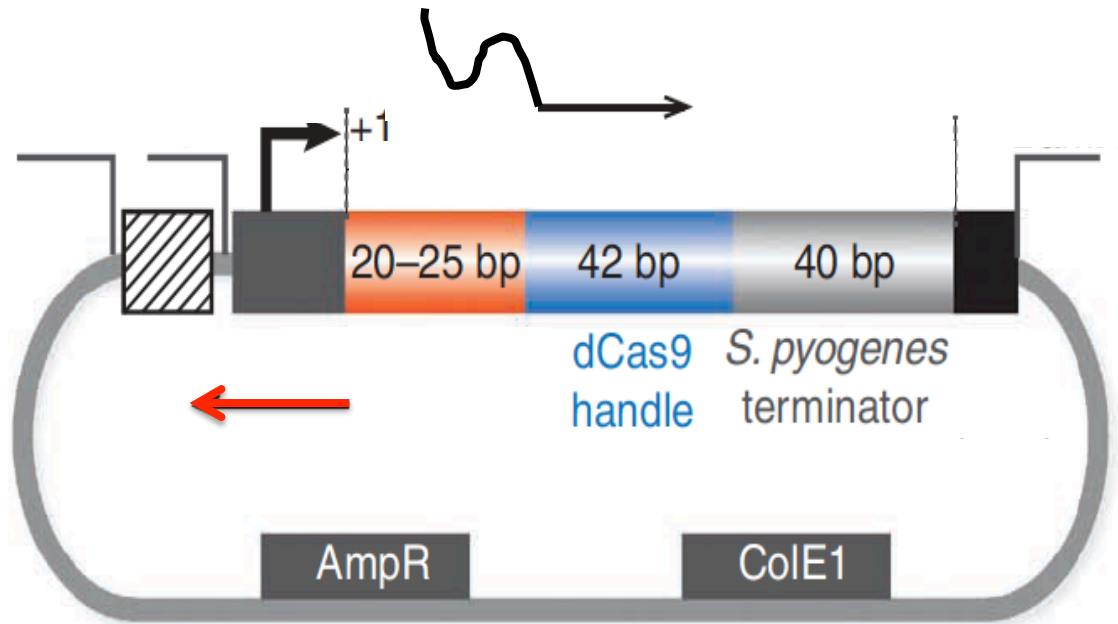
# Please add your targeting info to the wiki today

-> On the Mod2 Overview page, discussion tab

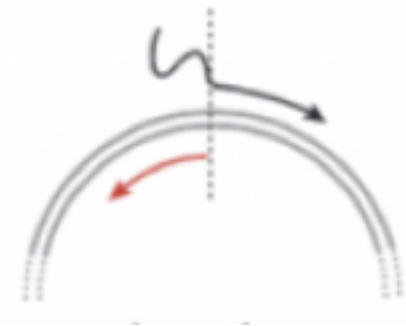
T/R: lactate [\[edit\]](#)

Team	Gene targeted by CRISPRi gRNA	gRNA sequence (without tag at 3' end)	Locus targeted (e.g. beginning of gene, putative promoter, -35 region)
yellow	pflB	TGTCGAAGTACGCAGTAAAT	
green	pflB	ATAAAAAATCCACTTAAGAAGGTA	
blue	pflB	TTCATTAAGCTCGGACATGTAACA	
pink	pflB	AAATAAAAAATCCACTTAAGAAGGT	
purple	pflB	AAATCCACTTAAGAAGGTAGGTGT	

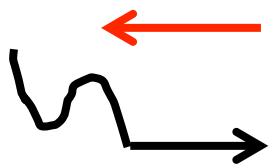
# M2D3: Generated pgRNA\_target by SDM



pgRNA\_template



insertion (NEB5α kit)



CRISPRi universal *amplification* reverse primer

forward primer including



crRNA to be inserted



dCas9 handle

# Analyzing Sequence Information

- Remember to reverse and complement your reverse primer sequence before alignment
- Consider importing your entire gRNA expression vector seq. with your target sequence if alignment is not working well

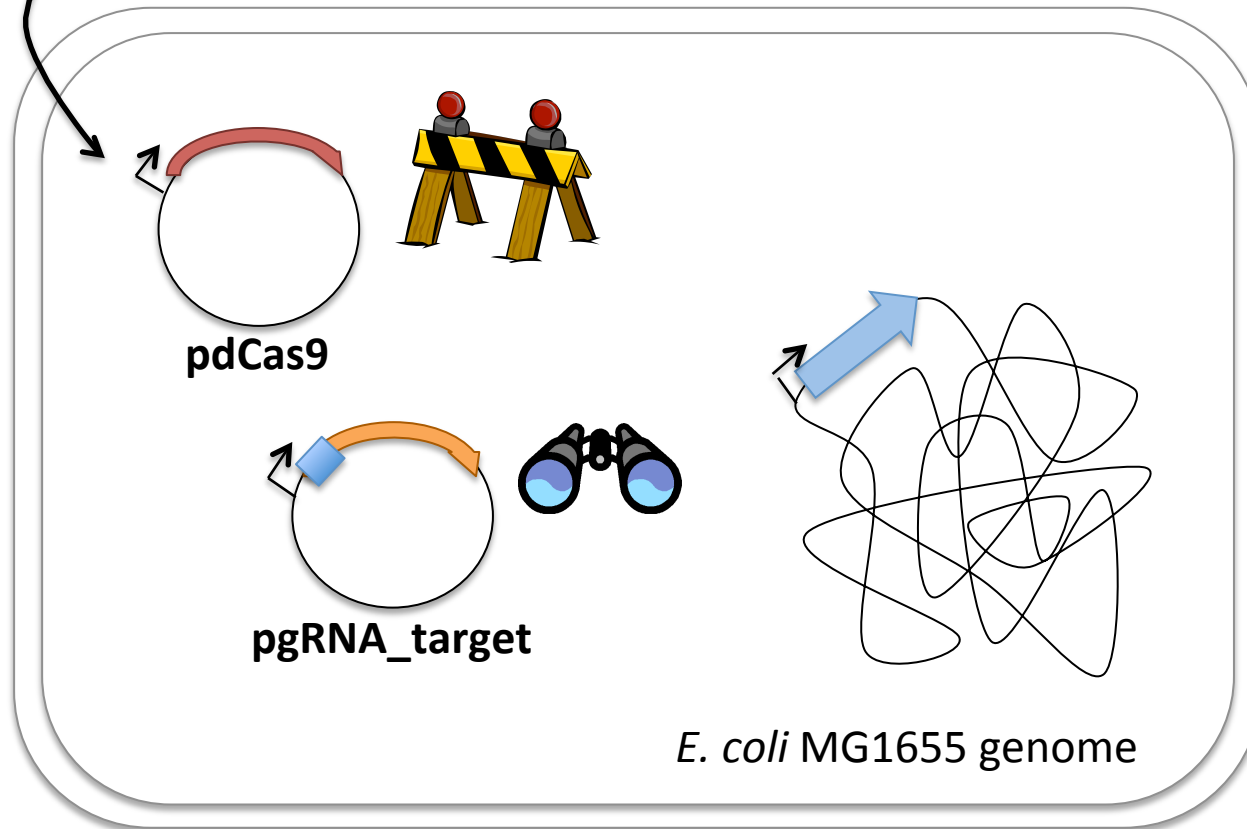
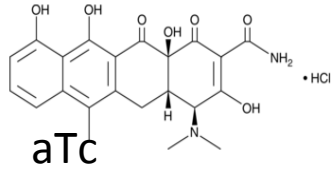
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0>-----*-----*-----*-----*-----*-----*-----*-----*-----*----->0
72>----->172
01>AAGAACCATTATATCATGACATTAACTATAAAAATAGGCGTATCACGAGGCAGAATTTAGATAAAAAAATCCTTAGCTTTGCTAAGGATGATTT>400
0>----->0

1>---gaattctaagatcctttgacagctagctcagtcctaggtataataactagt-----gtttagagctagaaatagcaag>73
73>---GAATTCCTAAGATCTTTGACAGCTAGCTCAGTCCTAGGTATAACTACTAGTAAATCCACTTAAGAAGGTAGGTGTGTTTTAGAGCTAGAAATAGCAAG>269
01>CTGGAATTCCTAAGATCTTTGACAGCTAGCTCAGTCCTAGGTATAACTACTAGTAAATCCACTTAAGAAGGTAGGTGTGTTTTAGAGCTAGAAATAGCAAG>500
1>---gaattctaagatcctttgacagctagctcagtcctaggtataataactagt-----gtttagagctagaaatagcaag>73

74>ttaaaaaaggctagtcctgattatcaacttgaaaaagtggcaccgagtcggtgctttttttgaagcttgggccgaacaaaaactcatctcagaagaggat>173
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01>TTAAAATAAGGCTAGTCCGTTATCAACTTGAAAAAGTGGCACCAGTCGGTCTTTTTTTGAAGCTTGGGCCCGAACAAAACTCATCTCAGAAGAGGAT>600
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```

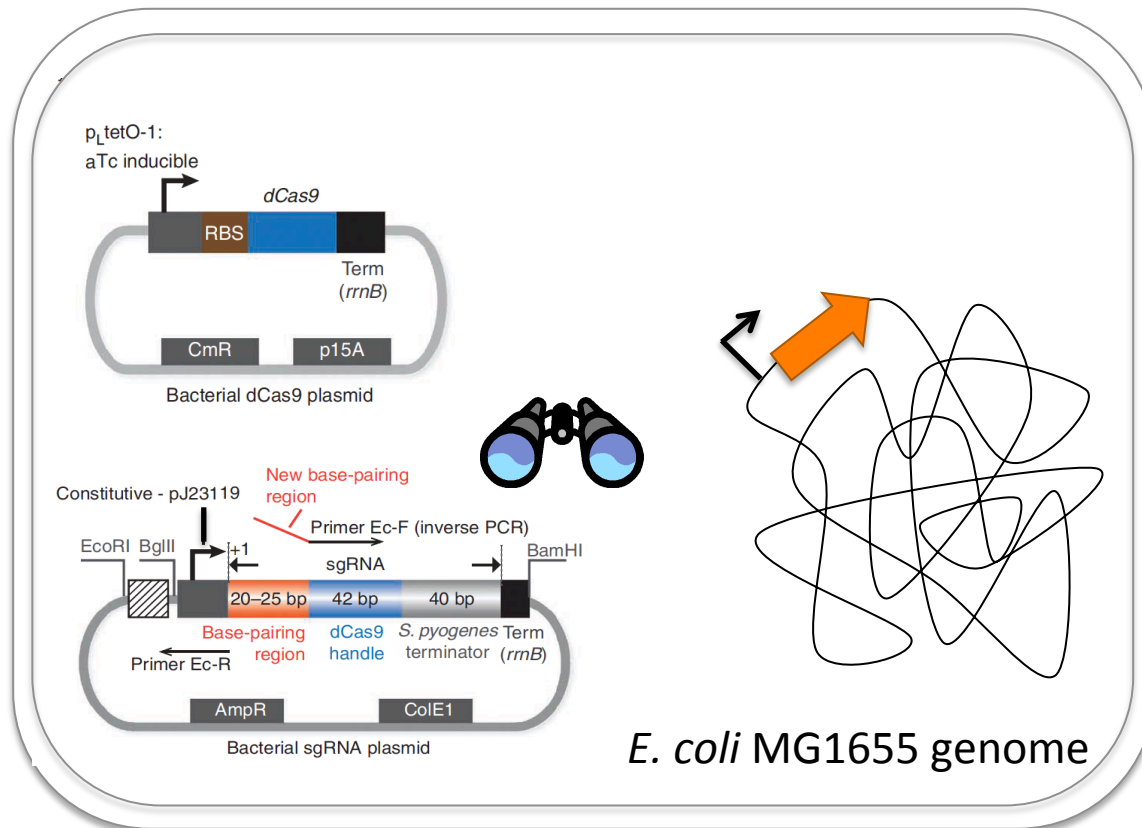


# CRISPRi system overview



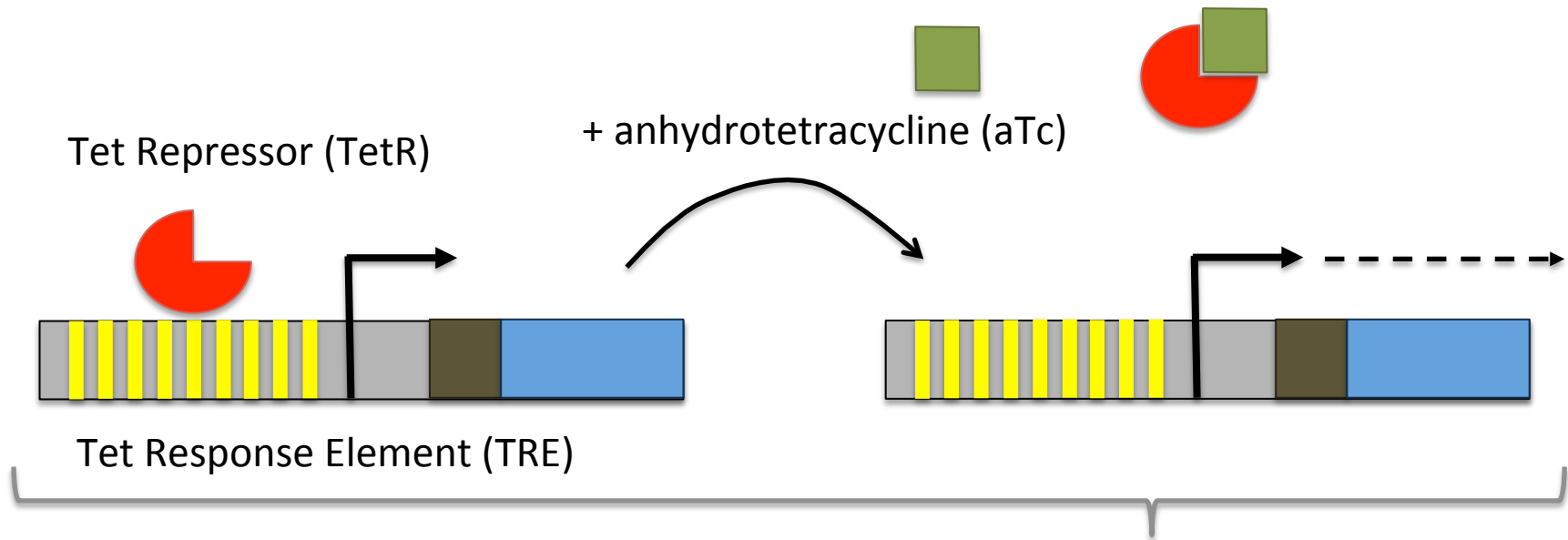
- Target gene
- pgRNA\_target
- pdCas9

# CRISPRi 'inactive' in absence of inducer

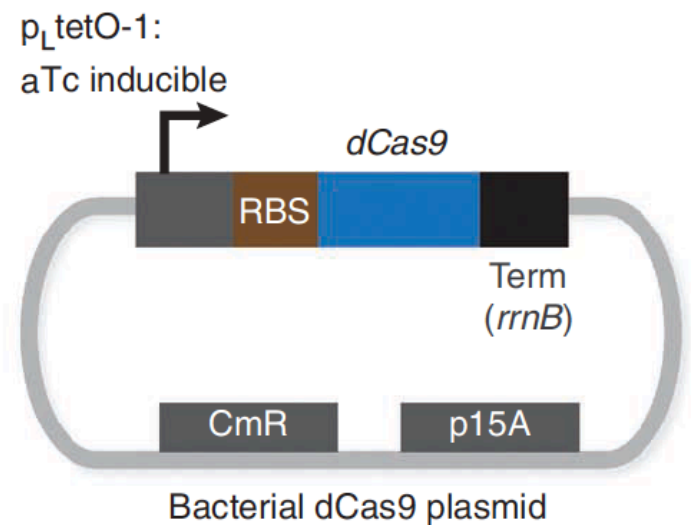


- pgRNA\_target expressed constitutively
  - Always transcribed and binding to target gene

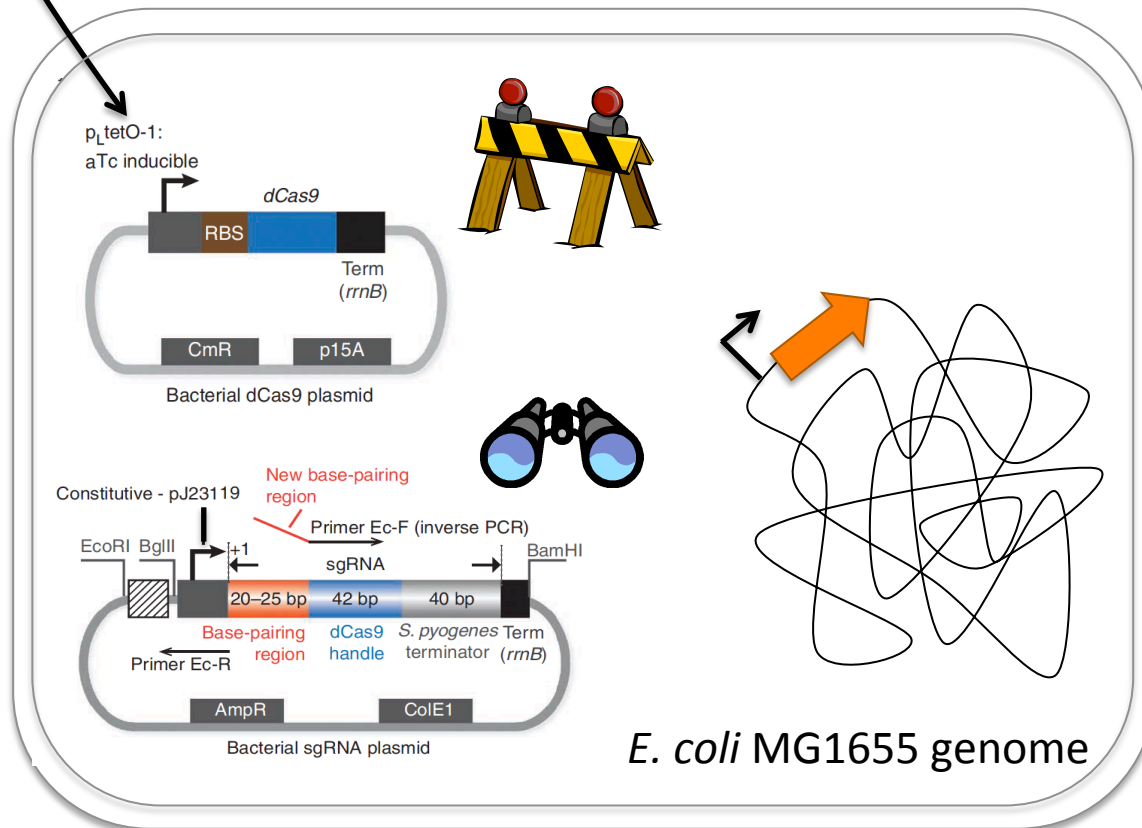
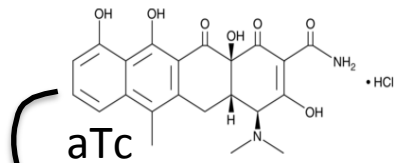
# aTc induction of pdCas9



- Tet promoter regulates expression of dCas9 gene



# CRISPRi 'blocks' gene expression in presence of inducer



- pdCas9 expressed when aTc added
  - When transcribed associates with pgRNA\_target / target gene

# Media for mixed-acid fermentation and pdCas9 induction

- What are the necessary components?

LB (E coli media)

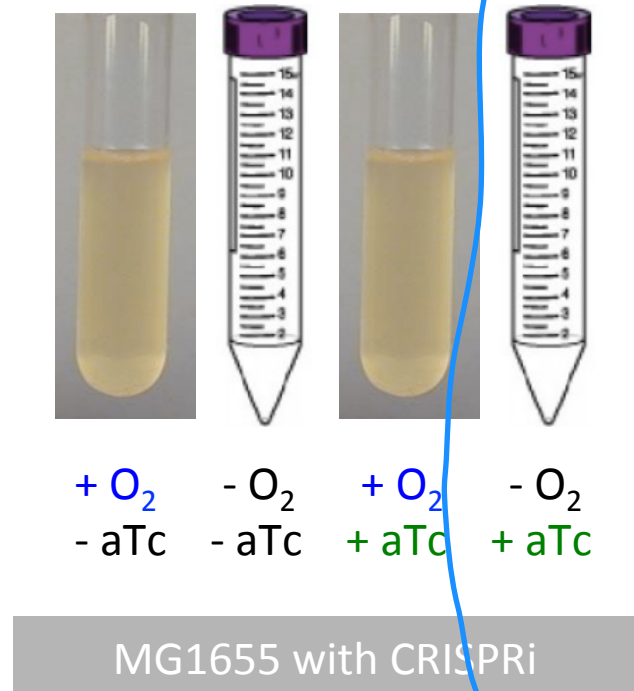
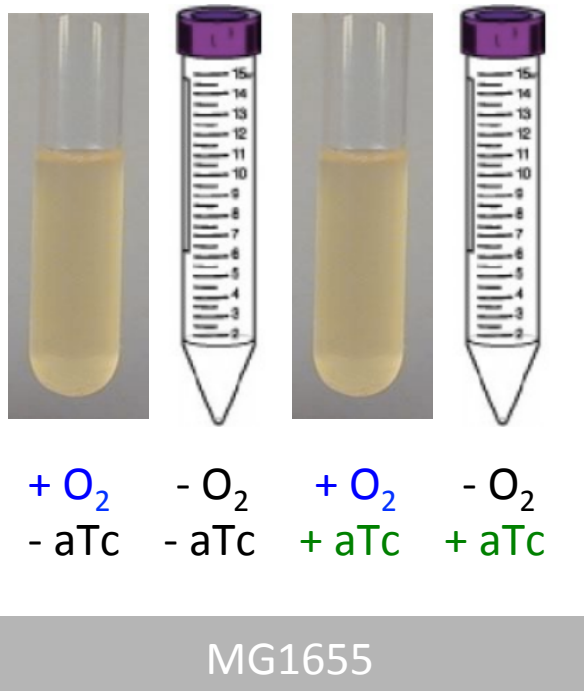
AMP ← antibiotics  
CML ←

ATC ← induce pdCas9 expression

# Media for mixed-acid fermentation and pdCas9 induction

*lactate*

- Where do we expect most ethanol if hypothesis confirmed?



# Today in lab...

- Download your sequencing data from discussion tab and align using ApE software
- Prepare media for mixed-acid fermentation inoculations