# Creating a bioremediation model system target selection and engineering approach



## Module Outline

- M2D1: Environmental heavy metal contamination
- M2D2: Model system target selection and engineering approach
- M2D3: Model system choosing and modifying a chassis
- M2D4: Screening a system—high throughput vs functional screens
- M2D5: Analysis of elemental metals laboratory and field approaches
- M2D6: Applying remediation strategies—advantages and pitfalls
- M2D7: Engineering a problem-specific bioremediation solution
- M2D8: Comm Lab



## Overview of today's lecture

- Choosing a target
  - Conceptual pipeline for target selection
  - Fet4 as a target
  - Other options to consider in our model system
- Modifying a target
  - Genetic engineering as an approach
  - Exogenous expression vs genomic integration
  - Rational design vs random mutagenesis



# Considerations for choosing a target



## How well characterized is the potential target?

- Is the sequence known?
- Has the structure been solved?
- Is the interactome known?
  - Are there known ligands or protein-protein interactions?
- Is the biological function(s) known?



- Fet4: low affinity iron permease
- Sequence and subcellular localization known
- Structure is predicted
- Non-essential
- Expressed mostly under anaerobic conditions

# What do we know about target interaction with our substrates?

- Fet4 takes up multiple types of divalent metals
- Fet4 knockout *S. cerevisiae* take up less cadmium than wild type
  - Endogenous Fet4 expression can be downregulated by cadmium
- The SMF1 transporter will take up more cadmium in response to a single point mutation
- Fet4 will change affinity for iron in response to a single point mutation



## What is the subcellular distribution of the target?

- Cytosolic vs. membrane bound
  - Transmembrane domains
- Cell surface vs. intracellular organelle
  - Localization sequences
- Localization is a key part of function
  - Provides insight into binding partners
- Localization-specific features can be points for manipulation or necessary to avoid



# How abundant is the target and under what conditions is it expressed?

- Protein expression can be tightly regulated
  - Location specificity
  - Condition specificity
- Endogenous Fet4 expression is regulated by ROX1
- Rox1 is a transcriptional repressor of hypoxia induced genes





## Does the target interact with other proteins?

- Proteins form complex interactions with other proteins
- Homo / heterodimers /tetramers
- Obligate vs non-obligate
- Stable vs transient



## How would modulating this target affect its native function?

- Change binding affinity or specificity?
- Change access to or preference for binding partners?
- Modulate function vs destroy function



## What are the downstream effects of modulating the target?



Cusabio

## Why was Fet4 chosen as a target for this project?

- 1. Previous functional observations
  - Transporter does not show strong specificity for a particular metal
  - Wild type Fet4 can take up cadmium
- 2. Expressed at the cell surface
- 3. Low affinity iron transporter
  - Iron uptake is redundant under normal conditions
- 4. Low basal expression in aerobic cultures
  - Overexpress our mutant without as much wild type noise
- 5. Not a highly conserved, ubiquitous protein
  - Fewer considerations for off target effects

What are other potentially effective S. cerevisiae cell surface metal transporters to target?



Genetic engineering to modify a target

# Genetic engineering encompasses a broad range of tools to manipulate targets

- Repression
  - Inhibit promoter
  - Inhibit RNA Polymerase
- Overexpression
  - Change endogenous promoter
  - Exogenous expression
- Addition
  - Integrate new gene into the genome
  - Exogenous expression of new gene
- Mutation
  - Alter gene sequence
  - Single or multiple changes



#### **Novel sequence insertion**



# Exogenous expression and genomic integration can both be utilized for genetic engineering



Random mutagenesis approaches for library development

 $H_2N$ 

Α

N N

Nvv

.....



## Using rational design for selective mutations

- Examine what is known about target:
  - Function
  - Structure
  - Orthologs



## Pros and Cons of mutagenesis strategies

### **Random mutagenesis**

### Pros

- Don't need as much information about the gene
- Unbiased screen casts a wider net

## Cons

- Lots of "junk" to sort through
  - truncations, silent mutations, etc...
- Multiple mutations in a gene

## Site directed mutagenesis

### Pros

- Able to selectively create single mutations
- Smaller population to screen for effects

### Cons

- Requires more background planning
- Easy to miss a potentially valuable mutant

## What are you doing in lab?

- Follow through with the actual mutagenesis based on your rational design
- Transform your mutagenesis mixture into *E. coli* 
  - Will purify colonies from this transformation to identify plasmid that has your mutation

