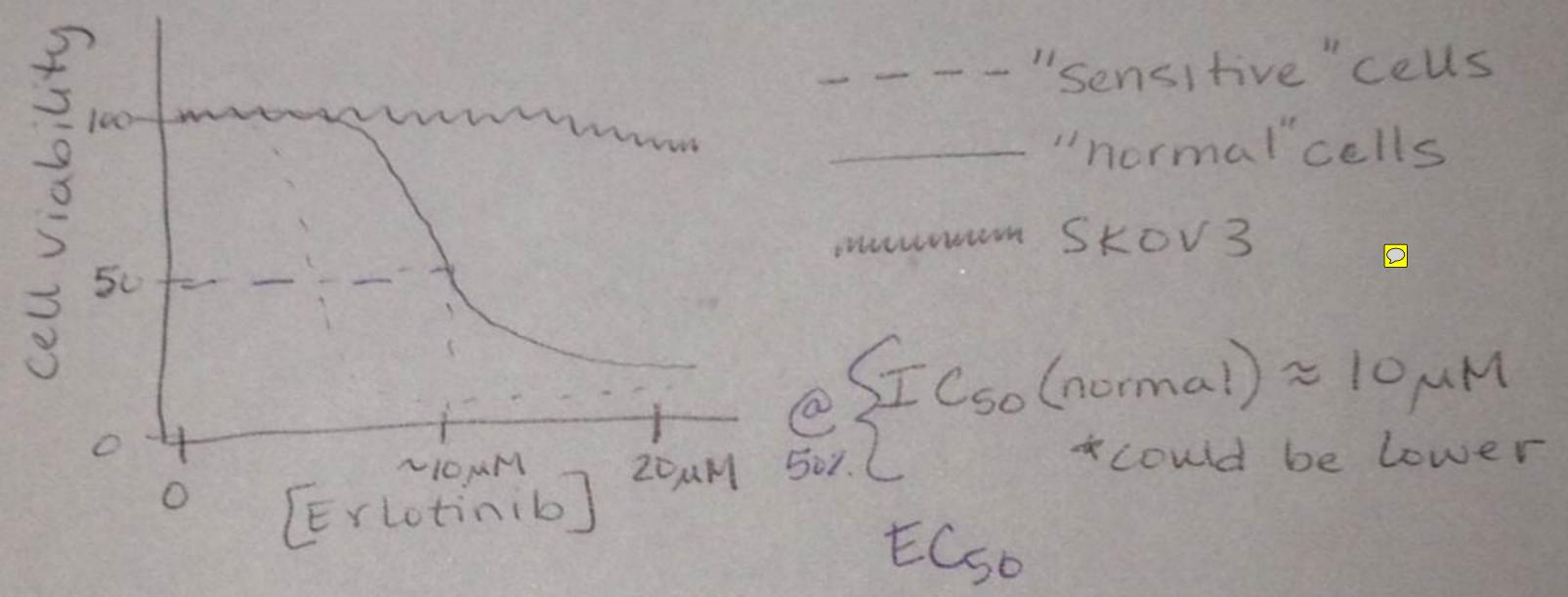
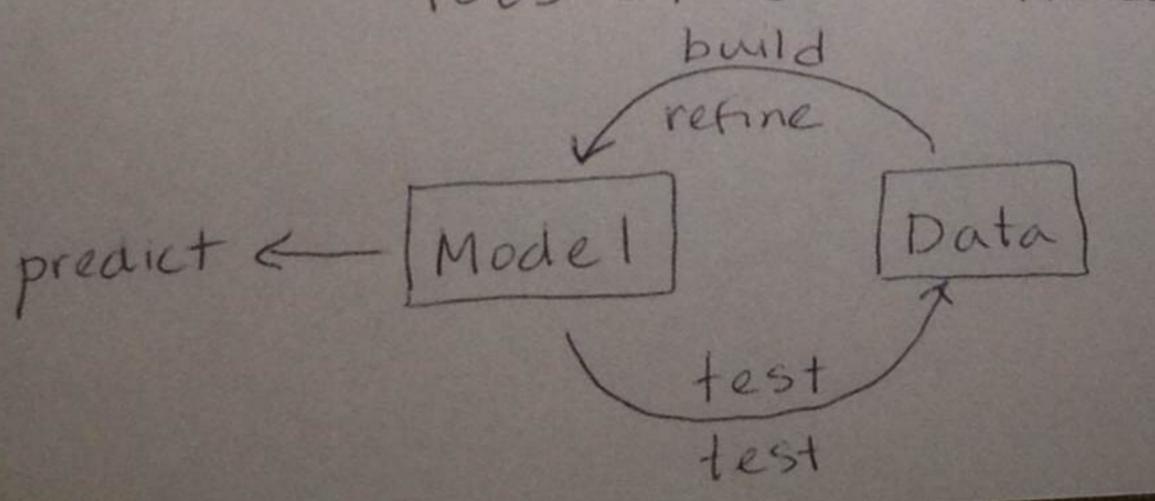
- 1) System -> SKOV3 ovarian cancer - Why? 60% o.e EGFR
 - EEFR inhibitors are unsuccessful in the
 - The mutation rate in ovarian cancer is low < 5% = "easy" to check
- (2) Biological Question/Design challenge



(3) How can we use a "systems biology" approach to increase effectiveness of EAFR inhibitors?

What is "systems biology"?

- Network view vs. favonte protein/gene
- quantification/analysis => modeling
- lots of data needed build



- (9) We use a "simple" ODE model to make a prediction => we will test our prediction on M2D7
- (5) In the meantime: Let's arm ourselves
 With more data?

What can facilitate resistance?

- Mutation V (we tested for highest
 probability)

 Laconclusion -> no mutation
 in EGFR
- 2) Alternative signaling pathways

EGFR: 2260K

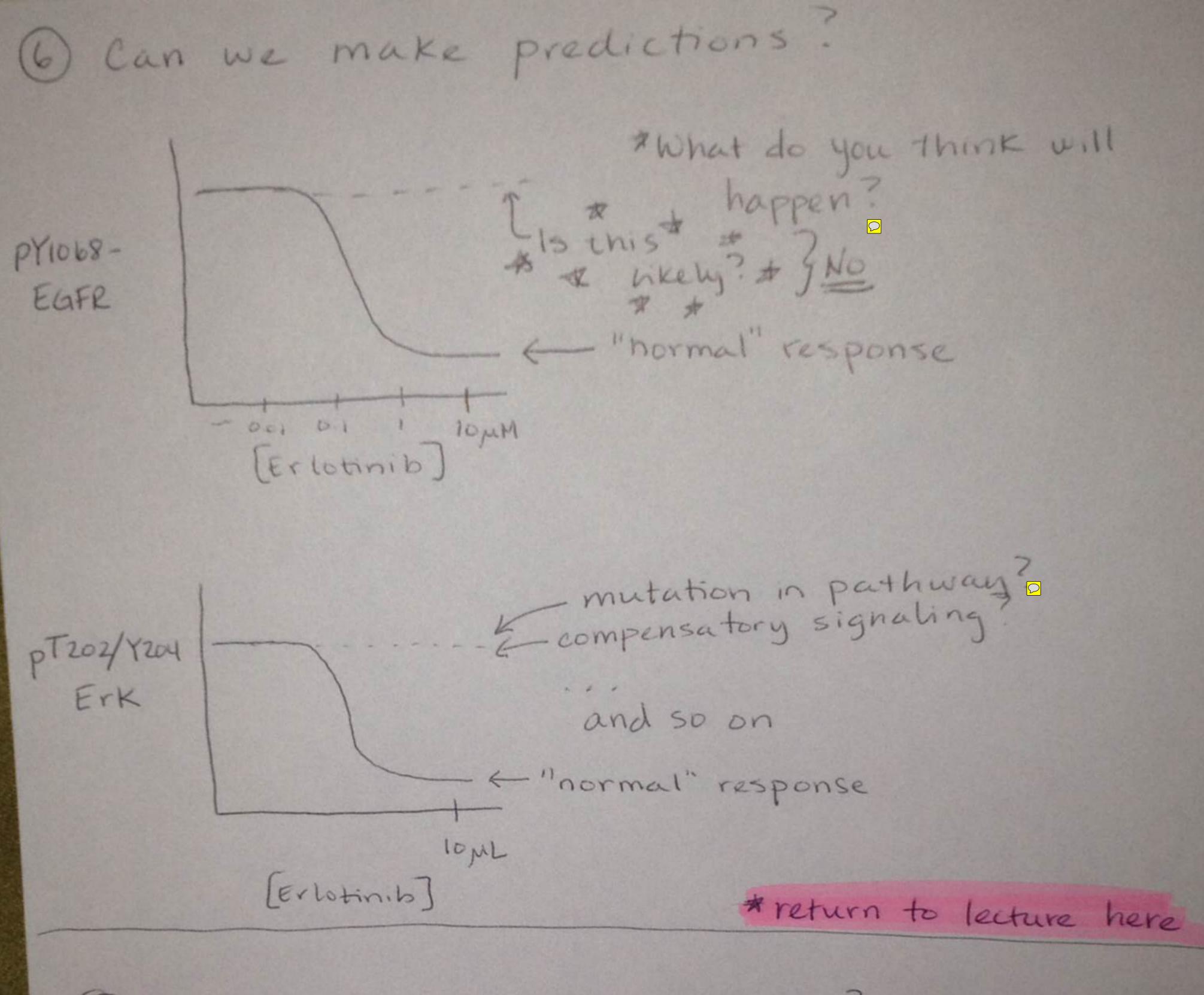
HERZ: 1.4M

HER3: 114K

What do we Know about this?

Look at PAKt, PErk & PSTAT3 after Erlotinib treatment

* go to stide



(7) What if they are all the same? = Next time

Module 2: Systems Engineering (M2D5)

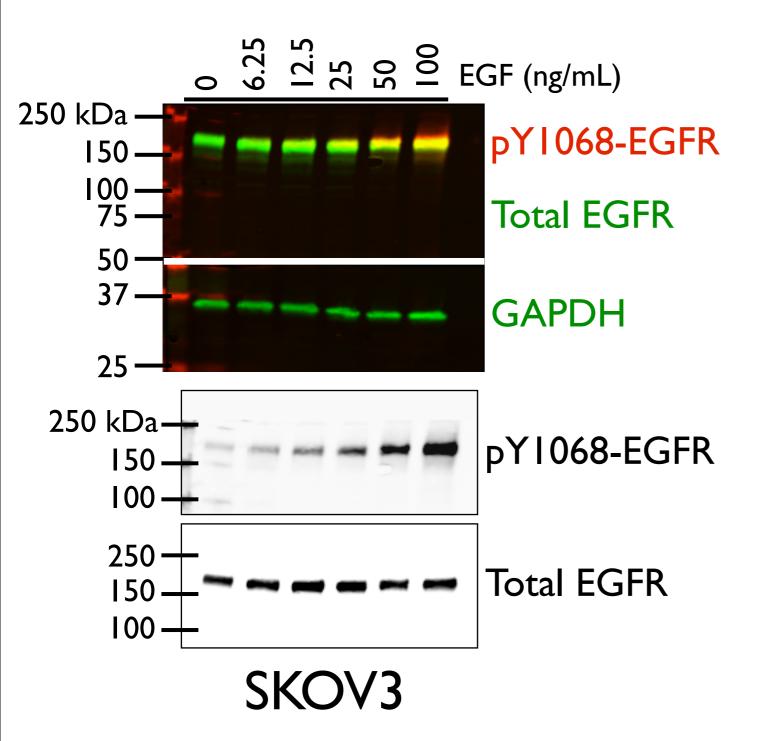
- In-depth about goals of Module 2 -- how does it all fit together?
- Scale it up!
- Talk about next time in lab: wiki pages done Saturday.
- Journal club!





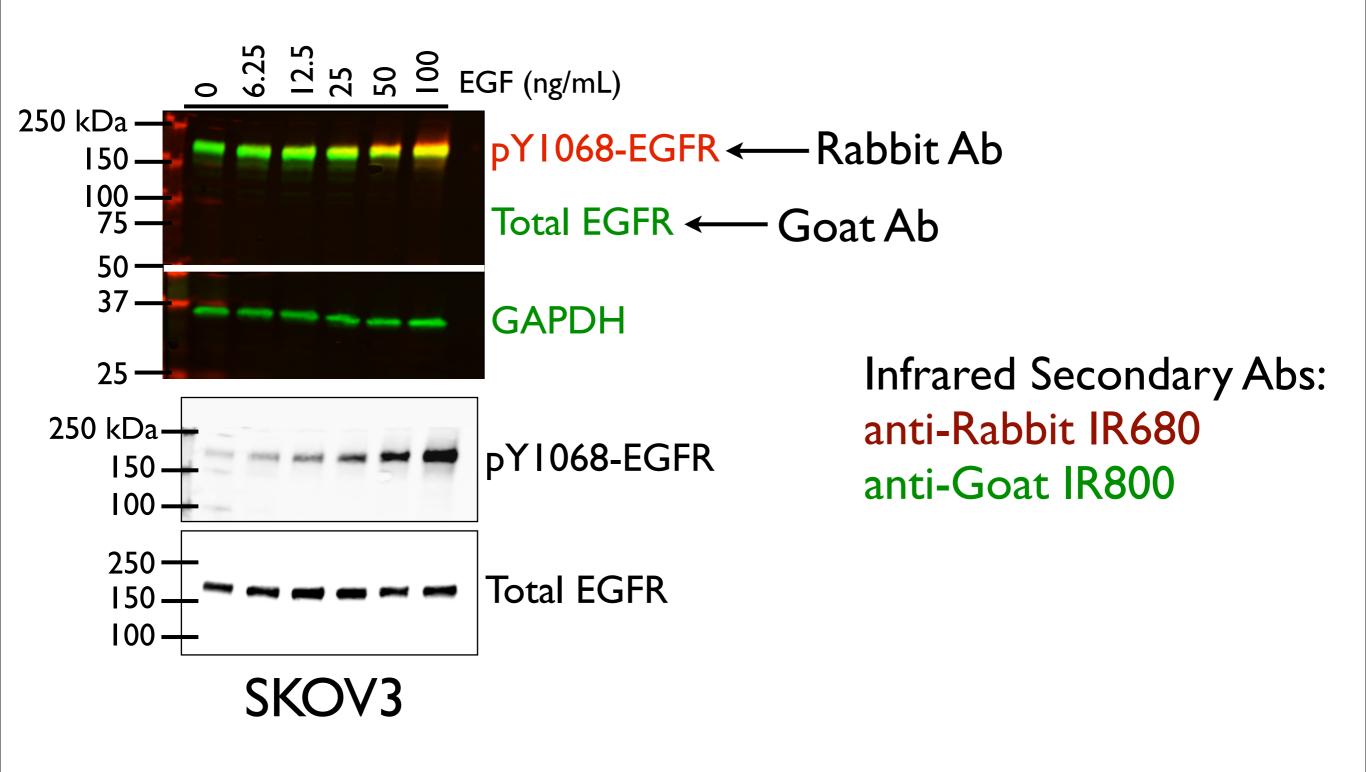


Semi-quantitative analysis: Western blot



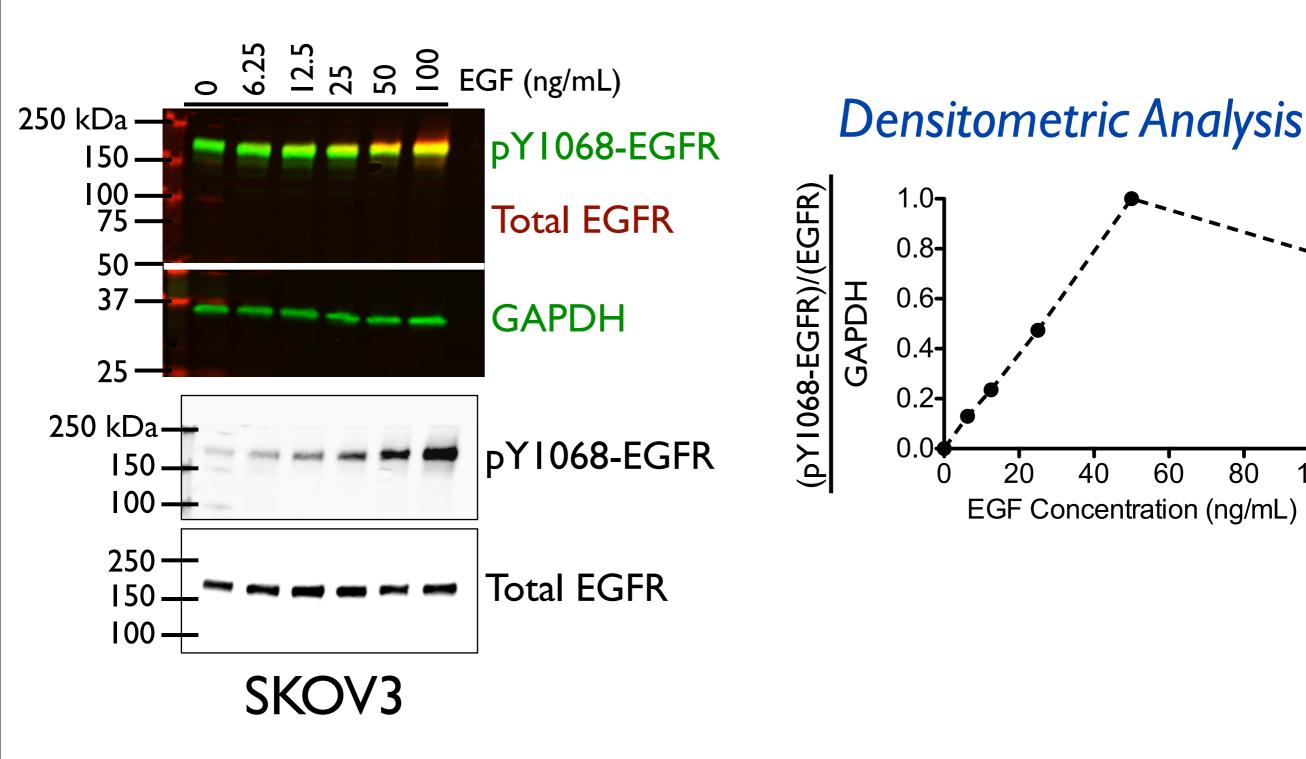
A few signals per lane -- up to ~17 lanes per mini gel.

Semi-quantitative analysis: Western blot



A few signals per lane -- up to ~17 lanes per mini gel.

Semi-quantitative analysis: Western blot



A few signals per lane -- up to ~17 lanes per mini gel.

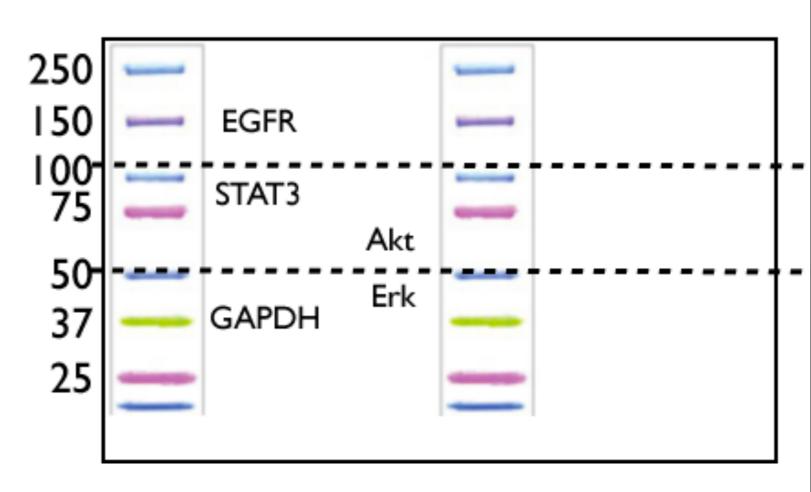
Semi-quantitative analysis: Your experiment

I.All: pY1068-EGFR & Total EGFR

2. Pathway specific:

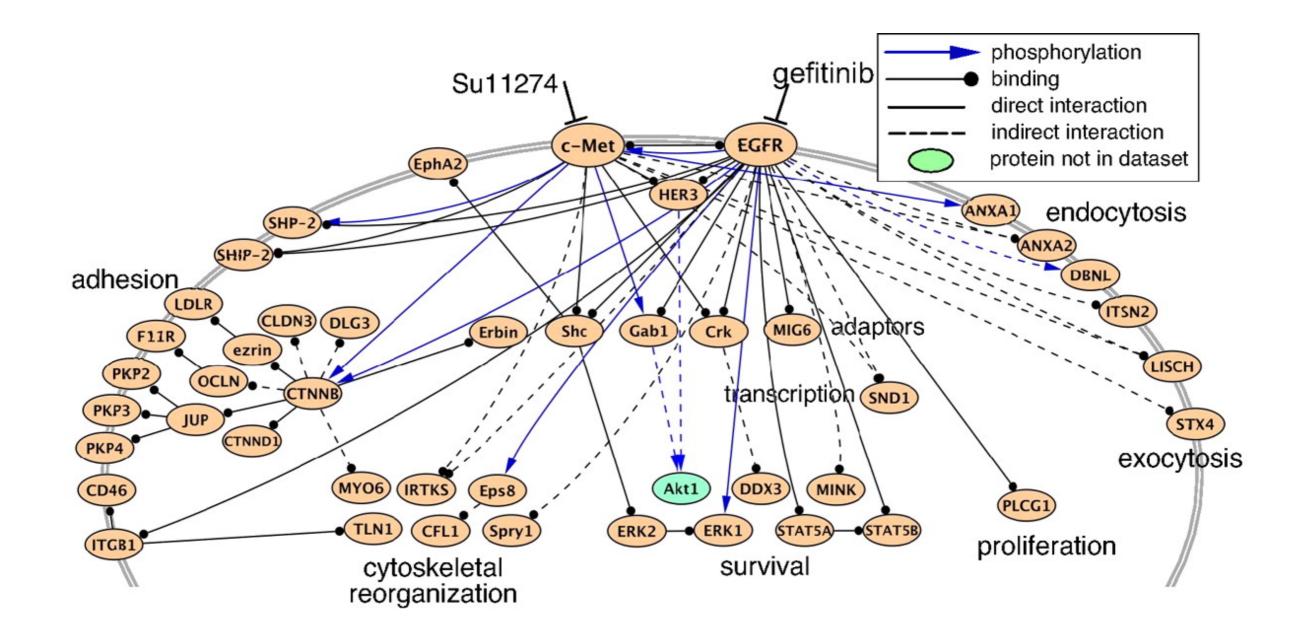
pERK & Total ERK
pAkt & Total Akt
pSTAT3 & Total STAT3

*Last two + GAPDH





What if we wanted a broader network view?



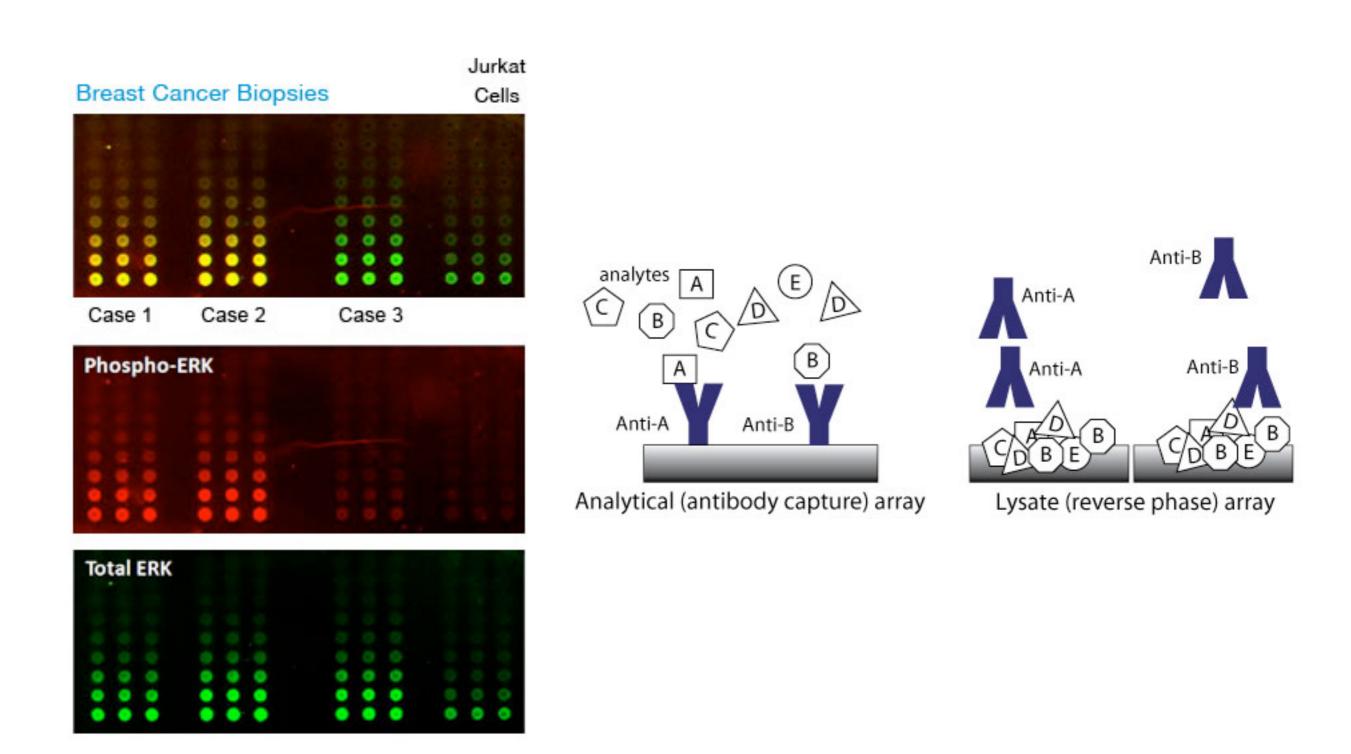
Regulatory networks sensitive to tyrosine kinase inhibitors in H3255 and MKN45 cells revealed by PhosphpScan-SILAC study.

Guo A et al. PNAS 2008;105:692-697

PNAS

©2008 by National Academy of Sciences

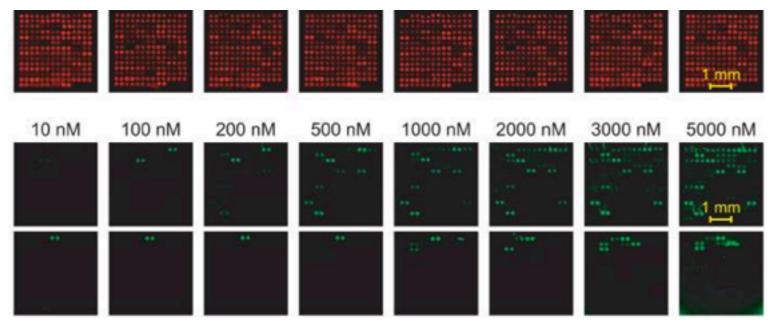
Semi-quantitative analysis: Protein Microarray



A few signals per spot -- up to hundreds of spots per slide.

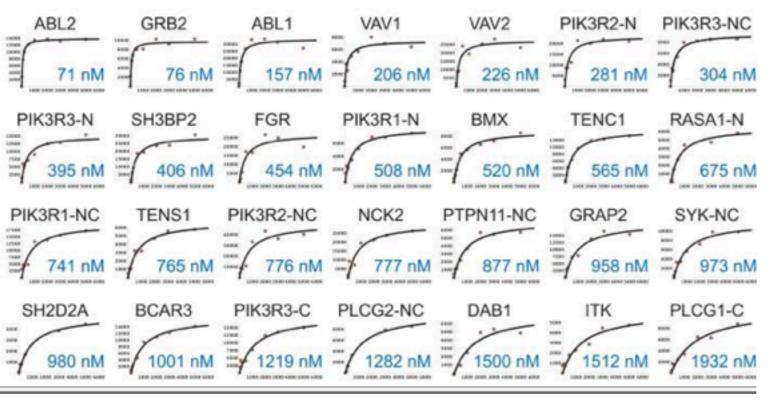
http://www.licor.com/bio/applications/odyssey_applications/rpa.jsp

Semi-quantitative analysis: Protein Microarray





Multiplex -- many conditions can be screened



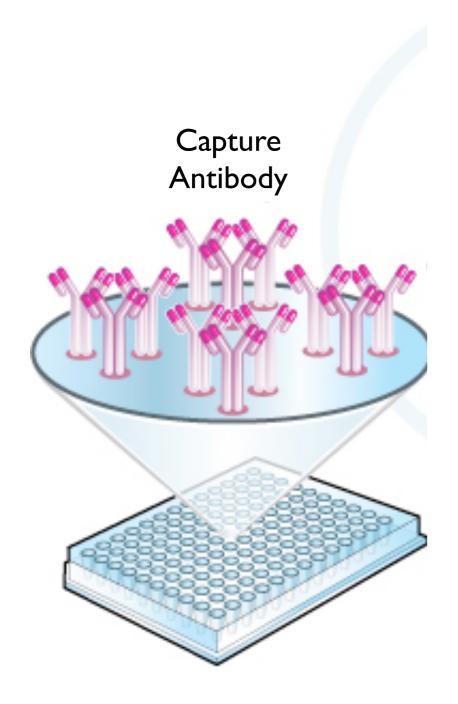
Weakness:

Non-specific interactions (also huge problem in WB)

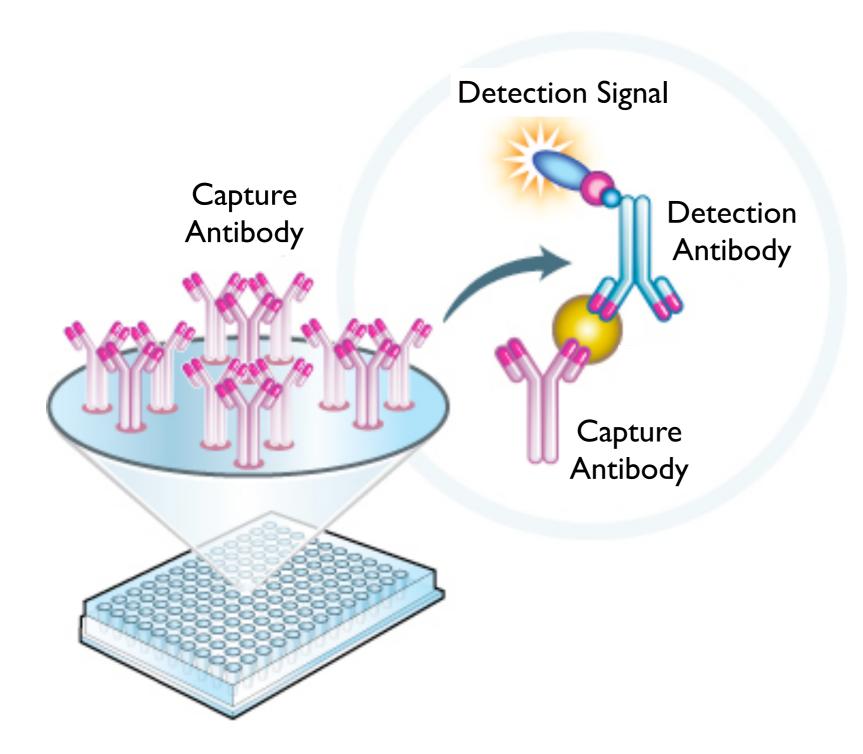
Dissecting Protein Function and Signaling Using Protein Microarrays

Curr Opin Chem Biol. 2009 October; 13(4): 398–405.

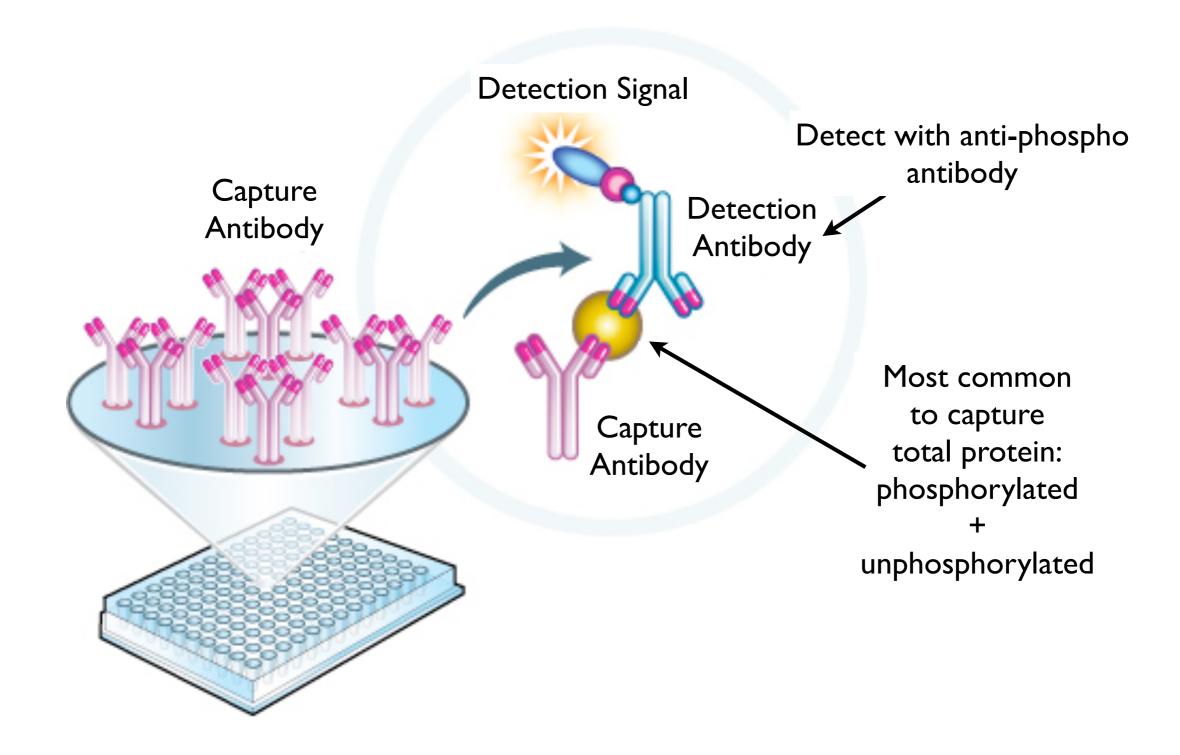
A few signals per spot -- up to hundreds of spots per slide.



One signal per well - but up to 384 wells/experiment.



One signal per well - but up to 384 wells/experiment.

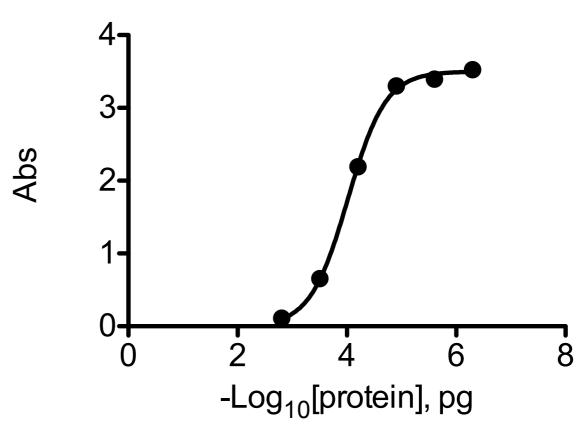


One signal per well - but up to 384 wells/experiment.

How to quantify:

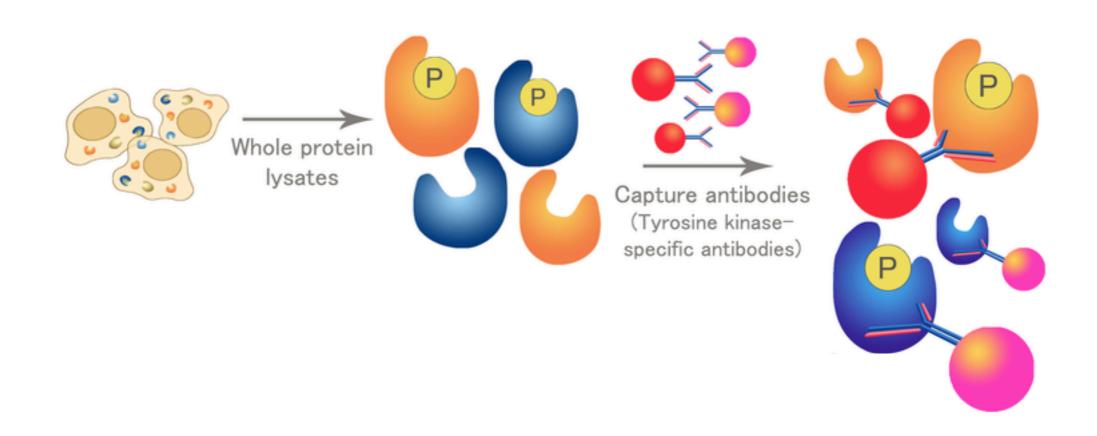
Very often not linear relationships

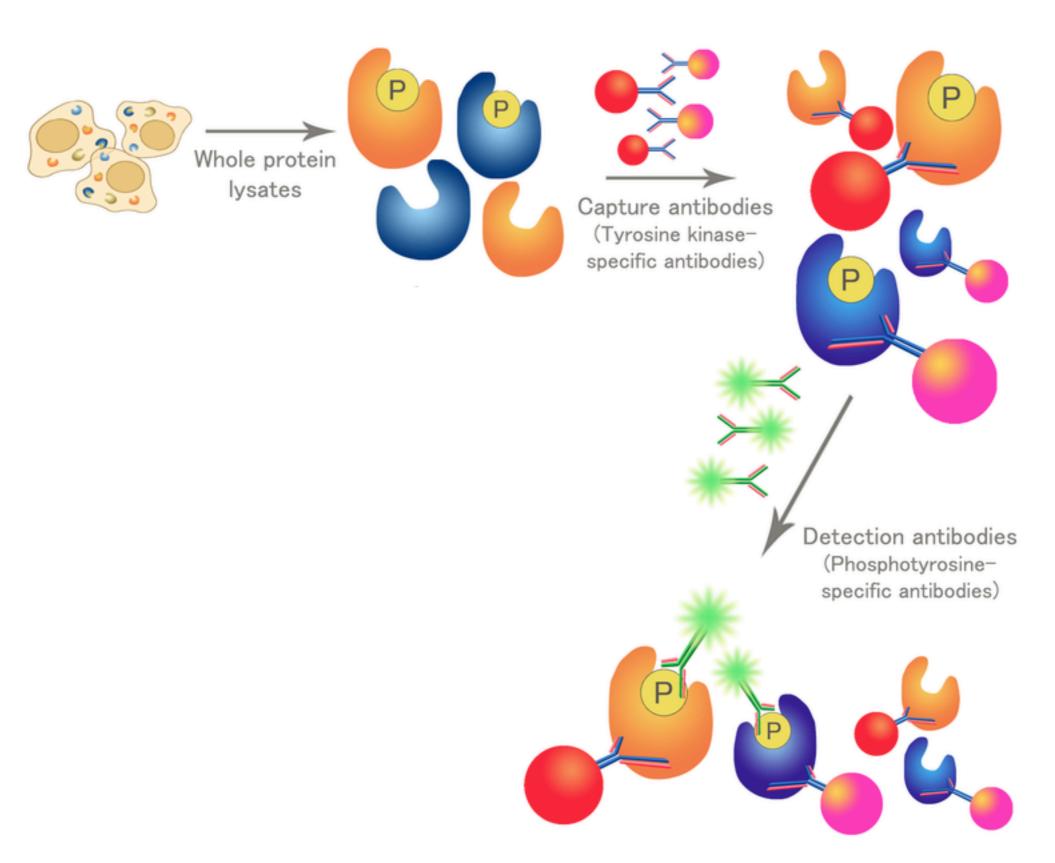
ELISA Standard Curve #2



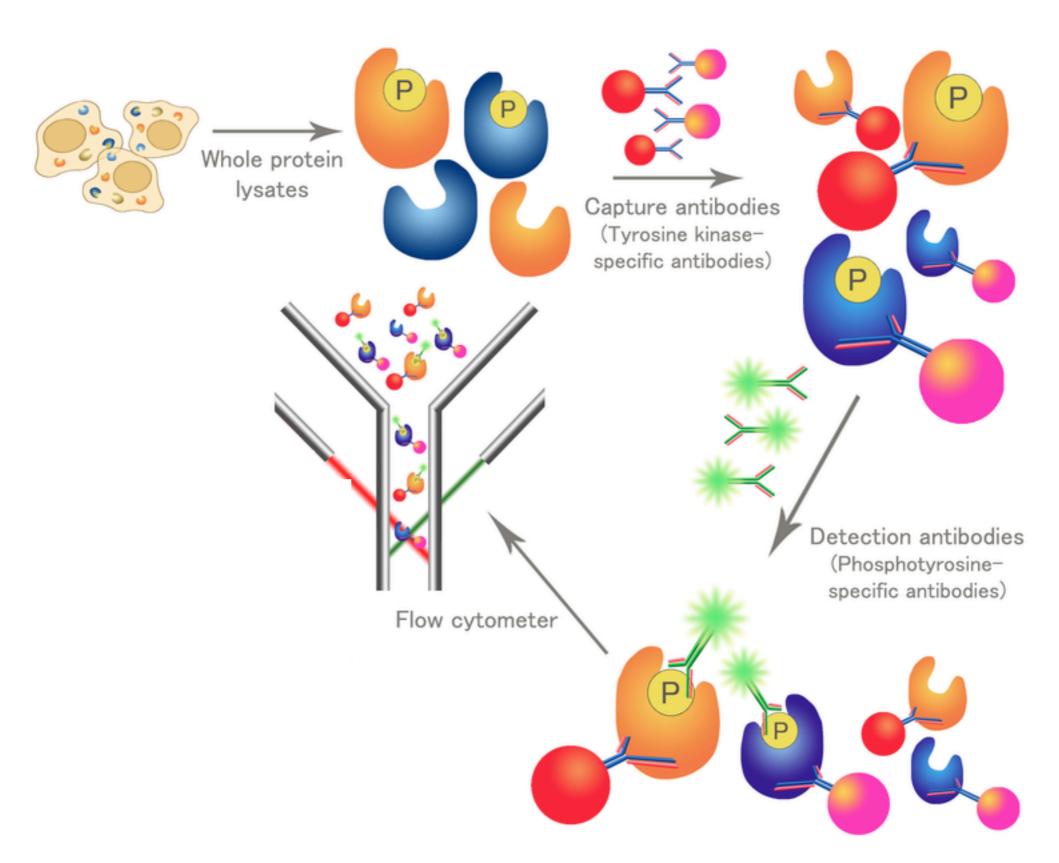
```
Y=Bottom + (Top-Bottom)/(1+10^((LogEC50-X)*HillSlope))
```

General protocol: Create a standard curve on each plate (usually supplied with kits or make your own)

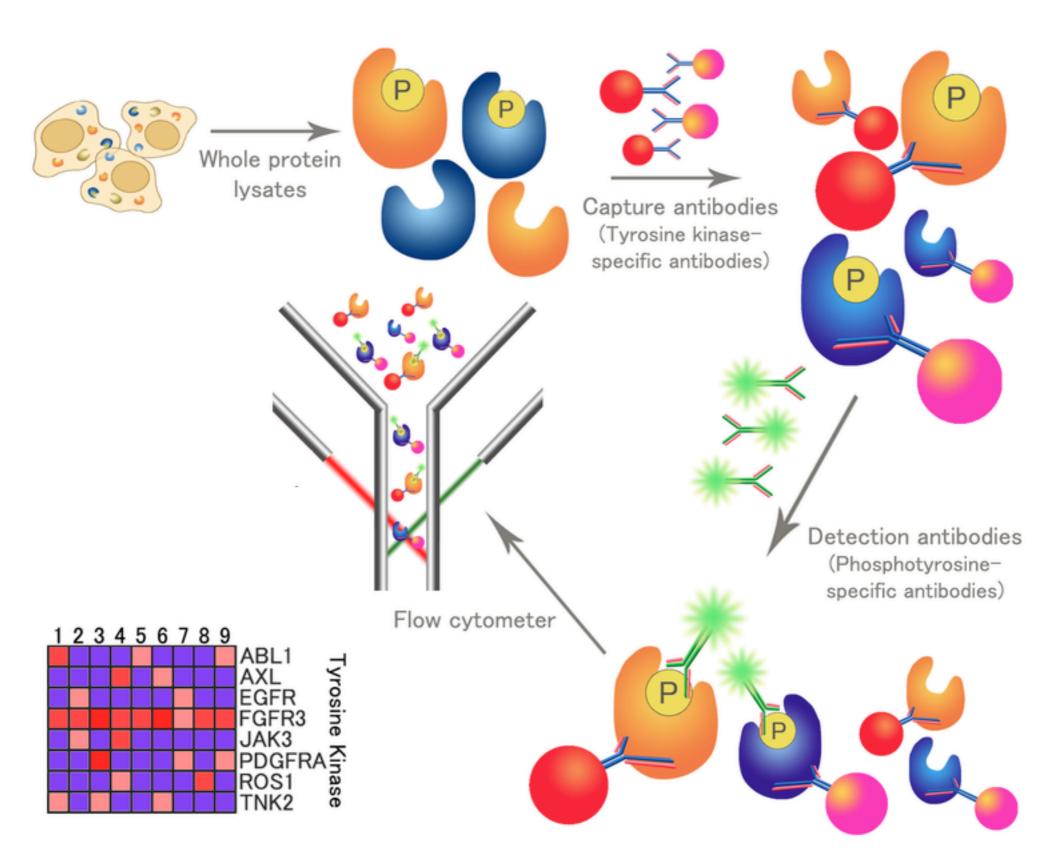




http://commons.wikimedia.org/wiki/File:Workflow_IA.png



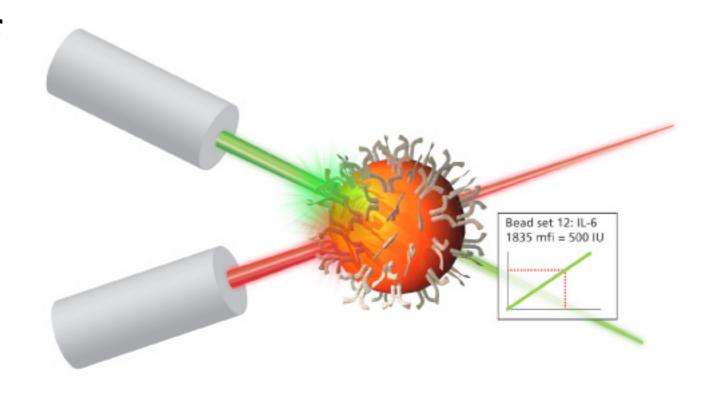
http://commons.wikimedia.org/wiki/File:Workflow_IA.png

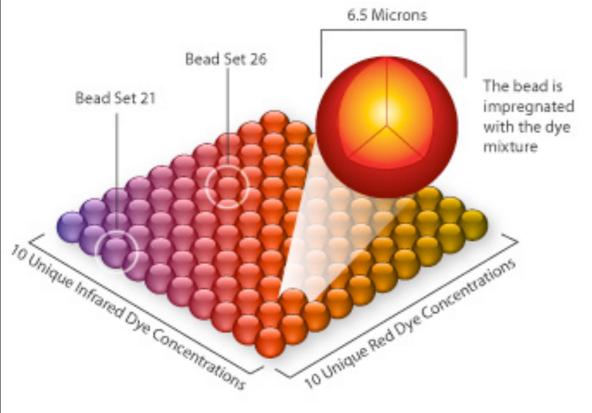


http://commons.wikimedia.org/wiki/File:Workflow_IA.png

Theoretically up to hundreds of conditions per well -- 384 (sometimes > 1500!) wells per experiment

In reality: 20-30 different phosphoproteins / well max







http://www.ebioscience.com/knowledge-center/product-line/procartaplex/technology.htm

Module 2: Systems Engineering (M2D5)

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