

# Module 1 overview

## *lecture*

1. Introduction to the module
2. Rational protein design
3. Fluorescence and sensors

## *lab*

1. Start-up protein eng.
2. Site-directed mutagenesis
3. DNA amplification

## **PRESIDENT'S DAY**

4. Review & gene analysis
5. Protein expression
6. Purification and protein analysis
7. Binding & affinity measurements
8. High throughput engineering

4. Prepare expression system
5. Gene analysis & induction
6. Characterize expression
7. Assay protein behavior
8. Data analysis

## **Lecture 4: Review & gene analysis**

- I. Review of the project
  - A. Project aims and rationale
  - B. Methods, work completed so far
  
- II. Analysis of mutant genes
  - A. Restriction digests
  - B. DNA sequencing

## **Module 1 assignment**

Protein engineering research article

1. Abstract
2. Introduction
3. Materials and Methods
4. Results
5. Discussion
6. References
7. Figures

## Module 1 assignment

Protein engineering research article

1. Abstract

2. Introduction

Why are calcium sensors important?

What is protein engineering; how does it relate?

What is inverse pericam?

Why is it useful/interesting to tune pericam?

Why did you choose your mutations?

3. Materials and Methods

4. Results

5. Discussion

6. References

7. Figures

## Module 1 assignment

### Protein engineering research article

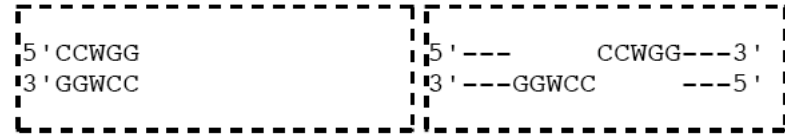
1. Abstract
2. Introduction
  - Why are calcium sensors important?
  - What is protein engineering; how does it relate?
  - What is inverse pericam?
  - Why is it useful/interesting to tune pericam?
  - Why did you choose your mutations?
3. **Materials and Methods**
4. Results
5. Discussion
6. References
7. Figures

## Restriction enzymes digest specific DNA sequences

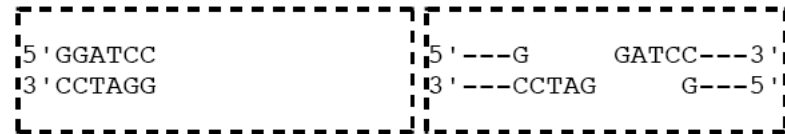
*EcoRI*      *Escherichia coli*



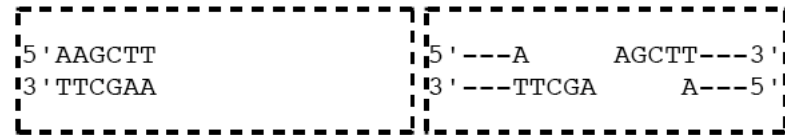
*EcoRII*      *Escherichia coli*



*BamHI*      *Bacillus amyloliquefaciens*



*HindIII*      *Haemophilus influenzae*



[www.wikipedia.com](http://www.wikipedia.com)

you designed mutations that can be assessed by restriction mapping:

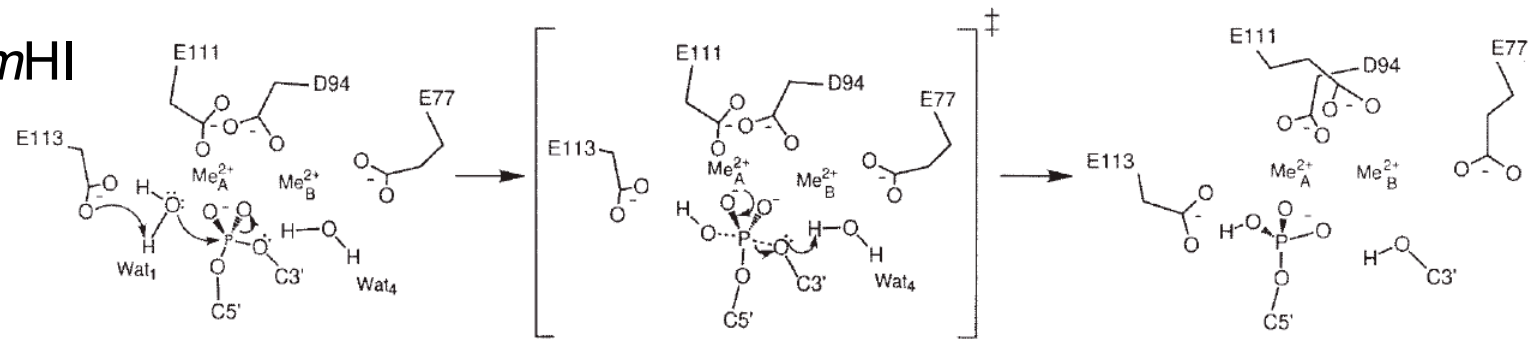
...TACATCAGCGCTGCTCAG...  
 ...ATGTAGTCGCGACGAGTC...  
 Y I S A A Q

...TACATCCTCGCTGCGCAG...  
 ...ATGTAGGAGCGACGCGTC...  
 Y I L A A Q



# How do restriction endonucleases work?

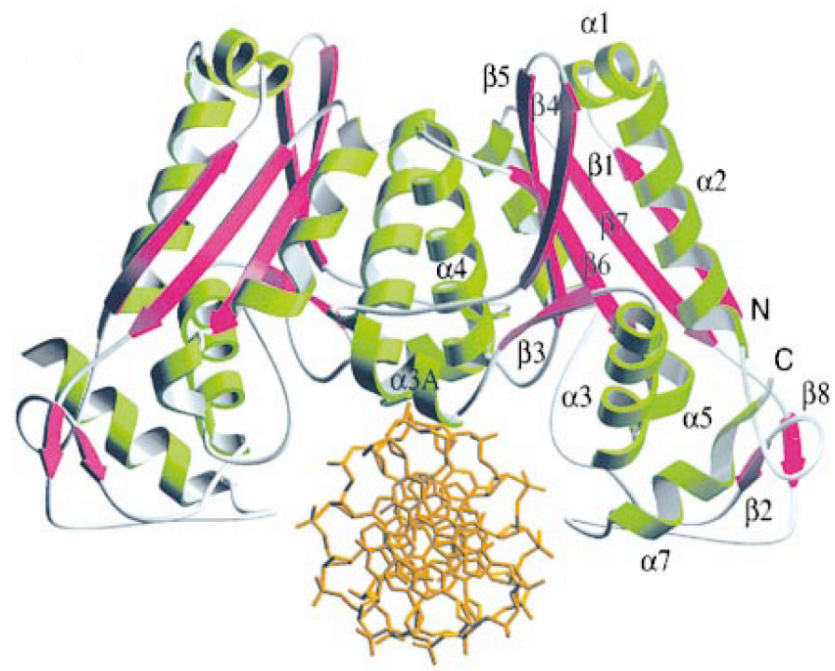
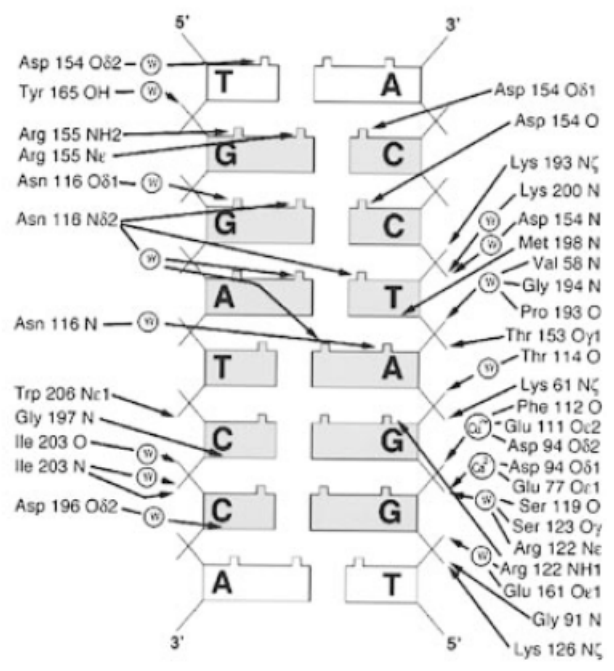
*Bam*HI

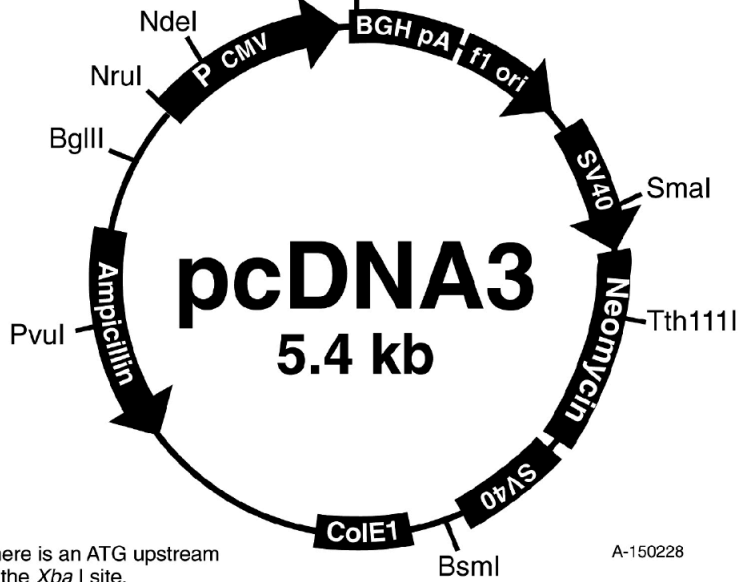
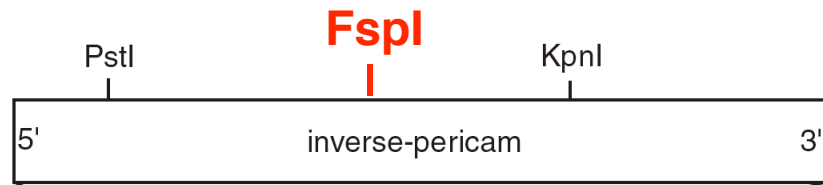


**Pre-reactive state**

**Transition state**

**Post-reactive state**

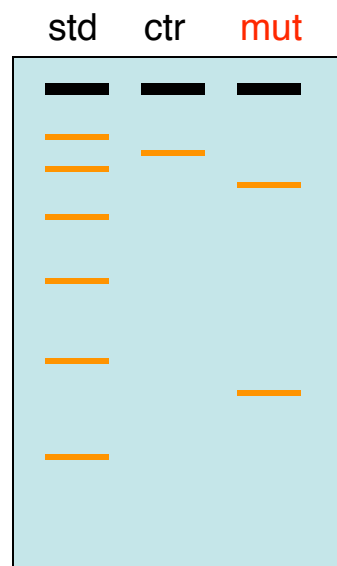




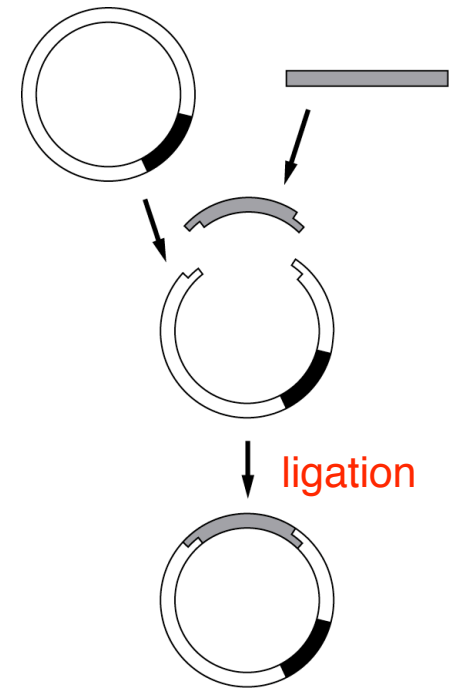
\* There is an ATG upstream of the Xba I site.

A-150228

diagnostic digest

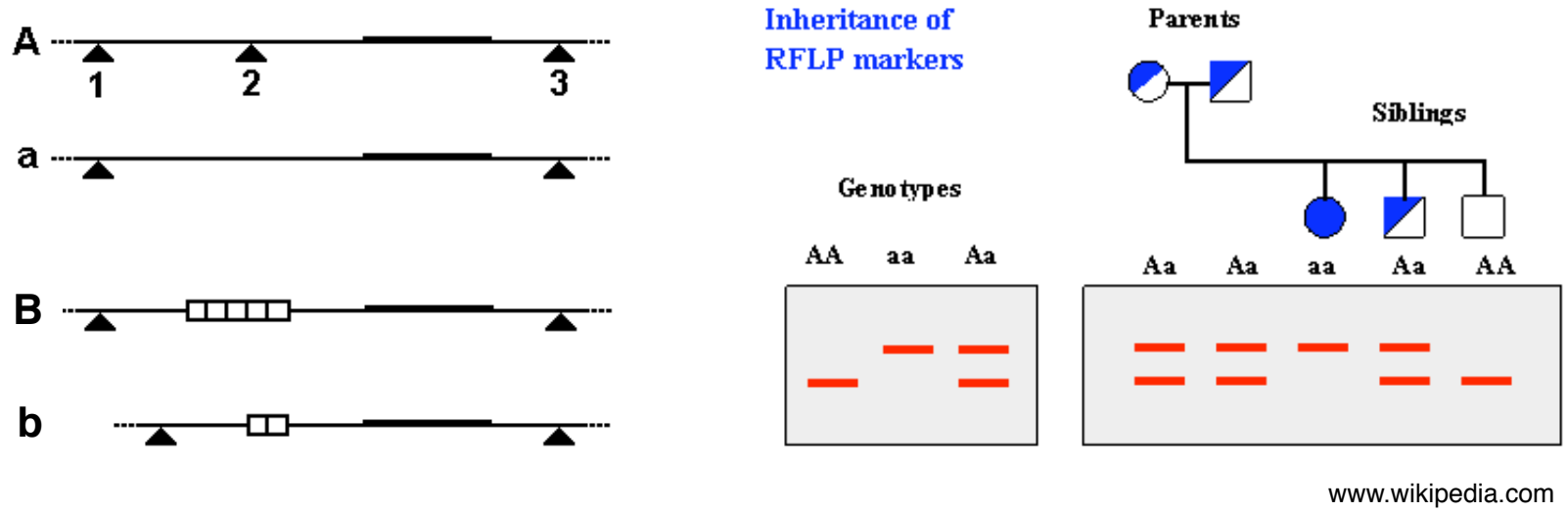


restriction endonucleases in cloning





Genetic polymorphisms can be associated with different distributions of restriction sites—restriction fragment length polymorphisms (**RFLPs**) used for genotyping



Suppose alleles A and B each occur in 50% of the population and segregated independently, what are the chances that a randomly chosen individual displays the AB phenotype?

How many biallelic polymorphisms would have to be considered for each genotype to have a 1:1,000,000 chance of occurring, assuming equal prevalence of each?

(This slide not covered in lecture, just for your info)

