M2D3: Assess purity and concentration of purified protein

- 1. Prelab discussion
- 2. Visualize protein purity with SDS-PAGE
- Measure protein
 concentration with BCA assay



Homework

Outline the Introduction for your Research Article

Due M2D4: Tuesday, Oct 31!

Structure of the Introduction

• Looks suspiciously like the Background and Motivation from the Data Summary...



- Broad context for your work
- Why is this work **important**?
- What information from the **current literature** is needed to understand the work?
- What **gap** in the current literature will your project address?
- What is your research goal/hypothesis?
- Here we show...
- Make sure transitions from one topic to the next are clear

Broad outline example

• Impact statement:

- Spinal cord injury (SCI) is bad
- Specific background:
 - <u>Concepts</u>
 - Why is this an important problem to solve?
 - What do I need to understand about SCI to follow this project?
 - <u>Components</u>
 - Introduce what we're working with (receptor and drug)
 - Techniques (if relevant)
 - Implant drug delivery system
- Knowledge gap:
 - Currently no effective treatments, perhaps because none are able to be given rapidly
- Research goal/Hypothesis:
 - Receptor-drug combination paired with the delivery system will encourage both axon growth from the injury site and functional recovery following injury

Needs transitions!

• Here we show...

- We see axon growth through the tissue scar following injury
- Functional range of motion is recovered

Lab work

SDS-PAGE gels and BCA assays

Notes on plasmid DNA on an agarose gel





Protein purification review

• Why this step?



- What's on the resin?
- What's in the expelled liquid?

Concentrate protein before testing

- Filter device sits within centrifuge tube...add protein to filter device for centrifugation
- Filter device has MW cutoff of 10 kDa ...**protein is retained in the filter device** during centrifugation
 - PfFKBP35 = 35kDa
 - His-tag = 2kDa
- How does this concentrate the protein?
- How does this remove excess imidazole?



How will you assess purity and concentration?

- Check **purity** using **SDS-PAGE**
 - Visual detection of other proteins in sample
 - Identifies purity of sample at multiple stages of purification
- Measure concentration using BCA assay
 - Colorimetric assay
 - Calculate concentration from standard curve





Purity: Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE)



- Laemmli sample buffer / loading dye:
 - SDS
 - β-mercaptoethanol (BME)
 - bromophenol blue
 - glycerol
- Boiling:

How are proteins separated?

- Laemmli buffer and boiling results in _____ and _____ charged proteins
- SDS-PAGE separates proteins by

- Electrophoresis completed in TGS buffer
 - Tris-HCl
 - SDS
 - Glycine



How are proteins visualized?

Coomassie brilliant blue G-250 dye used to stain gel after electrophoresis

- Hydrophobic and electrostatic interactions with basic residues
 - Arg, His, Lys, Phe, Tyr, Trp
- Complex between dye and amino acids is blue
 - Useful to visualize protein on a gel





Be mindful when assessing SDS-PAGE protein samples

Consider the order of your samples:

- 1. molecular weight ladder
- 2. pellet
- 3. lysate
- 4. flow-through
- 5. wash
- 6. elution
- 7. resin
- 8. concentrated protein.
- Figure will be included in your Research Article!



Concentration: Bicinchoninic acid (BCA) protein assay

Step 1: Chelation of copper with protein, reduction of copper sulfate to copper ion



BCA/Cu¹⁺ absorbance proportional to protein concentration

Standard curve generated using serial dilutions of bovine serum albumin (BSA)

- Use fresh tips between tubes
- Mix well between dilutions
- Be mindful of volumes



Protein concentration (µg/mL)

For today...

- Complete the purity and concentration assessments
 - Divide the work load here!
 - Start immediately by putting your Elution into the concentration column to spin