

- Announcements, Review HW
- Lab Quiz
- Pre-lab Lecture
  - ❖ Writing a Figure caption
  - ❖ DNA Extraction (Miniprep)
  - ❖ Diagnostic Gel Review
  - ❖ Safety + Technical Tips

# Announcements, old HW

- Can one group come to Tue Oct. 28 class? yellow
- Today at 3 pm, presentation on biosafety
- Your lab report is due in one week
  - OH (room TBA) on: Sun 2:30-9 pm  
Tue 4-5 pm
- Please post your colony counts in *talk* page table before leaving – we'll discuss them next time
  - \* show me your process, from raw data  
ladder = standards = known DNA lengths (bp)

# Figures: Style and Scope

- **Title:** concise, informative give overall result/goal.  
↳ often similar to that Results sub-section heading
- Caption: give context for result, from big to small.
  - Introduce what we are looking at.
  - Include only as much Methods as needed to understand result.
  - Define all elements (e.g., DNA ladder)
  - Cover primarily facts (results or expectations), limiting more complex interpretations → go in Discussion
- Aesthetics simplicity, clarity  
→ at-a-glance labeling when possible

# Figures: Example

\* tell a story  
title gives result  
(w/out over-interpreting)

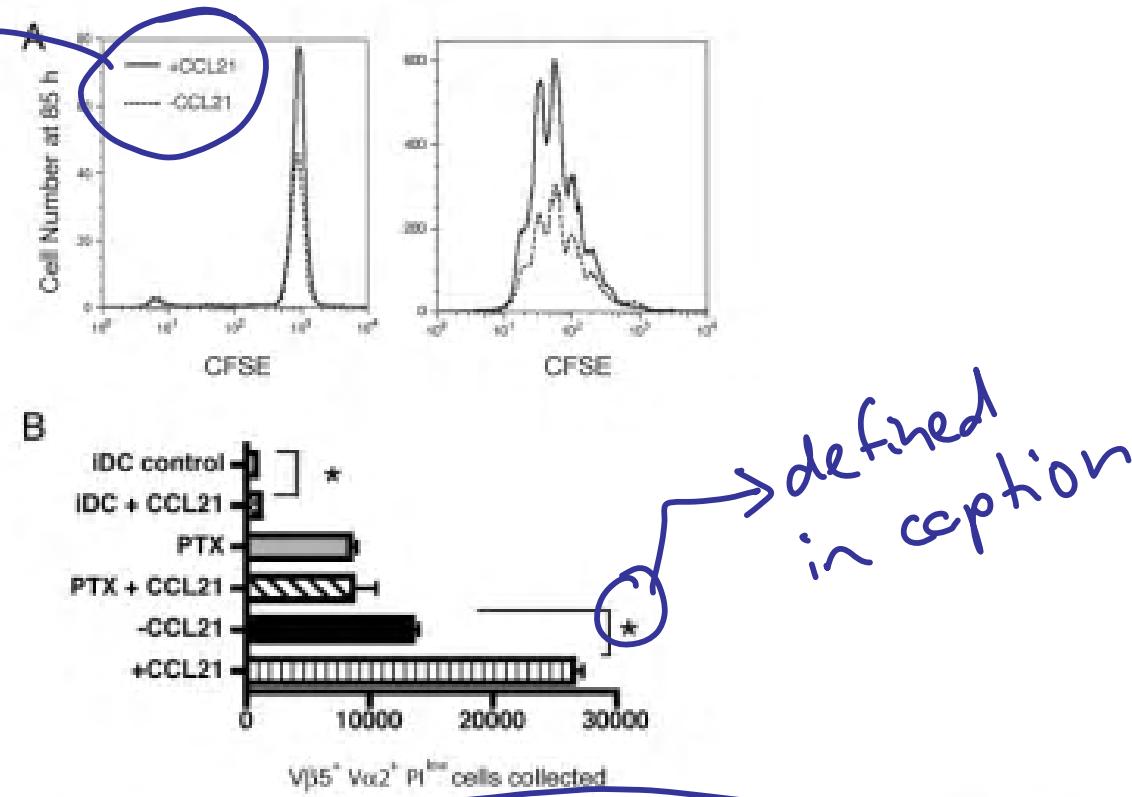


Figure 3 CCL21 impacts naïve T cell proliferation under conditions of rare Ag-specific T-DC encounters. Co-cultures comprising 9% OVA-specific OT-II CD4<sup>+</sup> T cells, 81% C57Bl/6 CD4<sup>+</sup> T cells, 5% OVA-mDC and 5% iDC with/without CCL21 were analyzed by flow cytometry at 85 h. (A) Sample CFSE histograms are shown for control (left, iDC only) and experimental (right, with OVA-mDC) conditions. (B) OTII cell recovery for all conditions is shown. Ave  $\pm$  std. dev. for 3 wells per condition. [\* indicates bracketed conditions statistically different ( $p \leq 0.05$ )]. (A-B) are from 1 representative of 5 experiments.

Overview  
{ of exp.  
} details of  
} results

# Figure Captions: Practice

Fig. 3

## Fig. N functional title descriptive caption

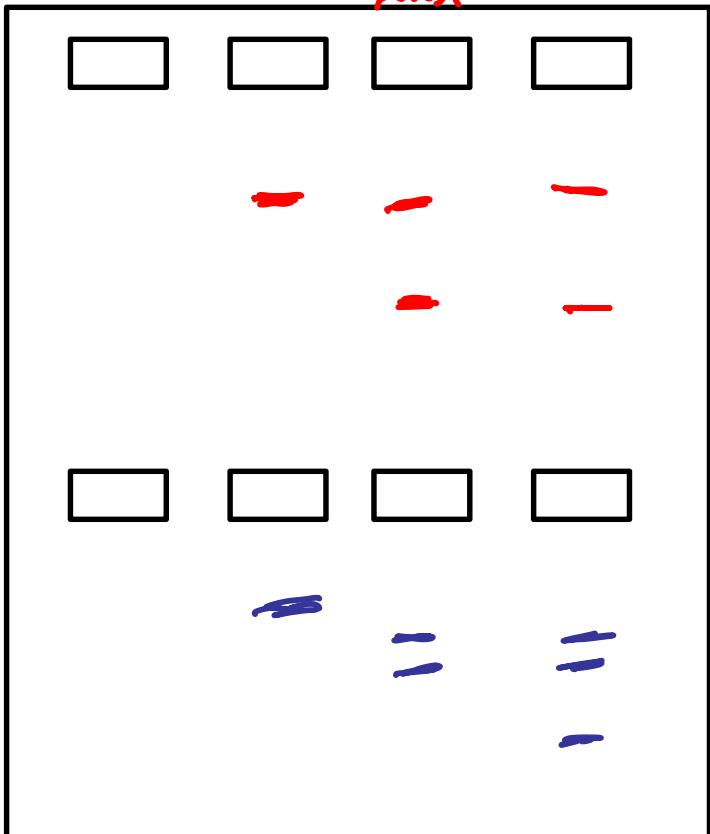
- Agarose gel title ideas: restriction digest,  
restriction digest to { confirm and extract }  
DNA fragments for cloning
- Overview sentence topics: define pCX-NNX, D32GFP  
(actual/expected sizes)
- Supporting detail topics: all observed sizes = results  
and what they mean, label reference bands in ladder
- Methods to include or not:  
~~4% agarose~~ 4% agarose V  
~~volume of loading dye~~ (1% agarose)

# DNA Extraction from Bacteria

Step	Contains	Purpose
Soln. I	EDTA buffer, glucose	→ weaken cell envelope → otherwise stable
Soln. II	SDS $\text{mNa}^+$ NaOH	→ disrupt membrane, solubilize lipids/proteins → ds DNA → ss DNA $\odot \odot$
Soln. III	acetic acid/ KAc.	→ neutralize pH, precipitates SDS $\downarrow$ plasmid
Transfer	N/A purpose : isolate plasmid	genomic DNA "trash" $\odot \odot$ renature $\odot$
Final steps	EtOH, $\text{H}_2\text{O}_1$ , drying	→ precipitating DNA, washing away salts, etc. → ethanol would interfere w/digest

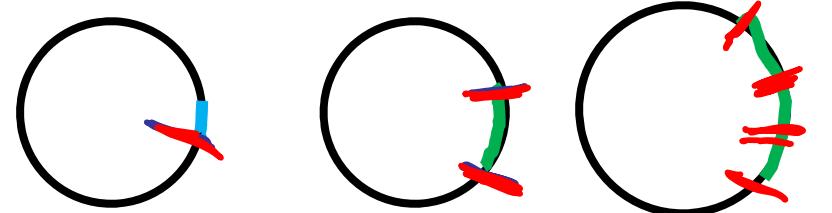
# Diagnostic DNA Gels

(bk<sub>b</sub>) (bk<sub>b+</sub>) ↗(multiple)  
 NNX ΔS- NNX M ins.

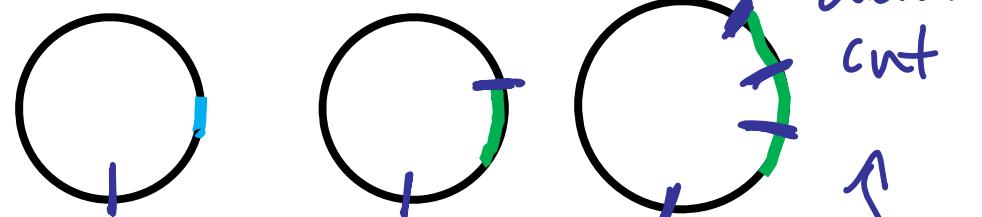


Choosing restriction sites for digest

BamH I  
E(CR)



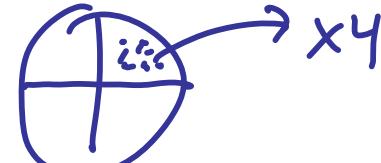
BamH I  
XbaI



one enzyme didn't cut

- \* choose one site uniquely on insert, one on backbone
- \* consider what data would look like if

# Today in Lab

- Miniprep three  $\Delta 5$ -EGFP candidates, and bacteria transformed with pCX-NNX  
*control for your technique*
- Count and post colony #s  

- Visit from Rhonda O'Keefe, EHS *3pm*
- Set up digests
  - We will add loading dye if lab runs late