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 - ❖ DNA extraction (miniprep)
 - ❖ Diagnostic gel review
 - ❖ Intro to tissue culture
 - ❖ Today in Lab: M1D5

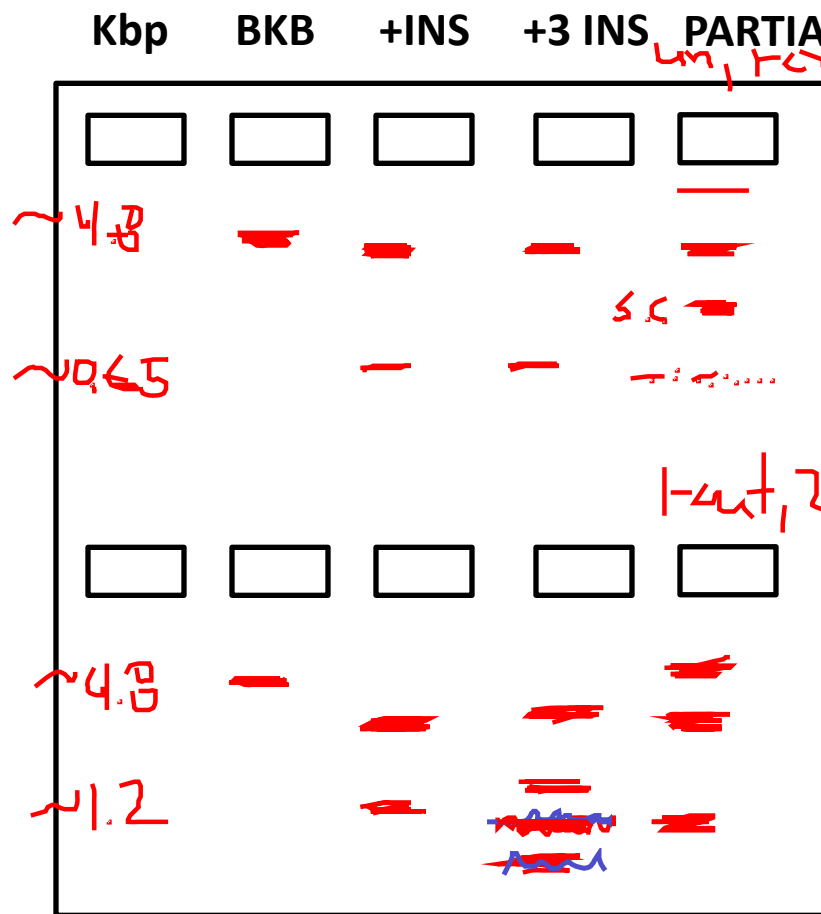
Announcements

- Next time: lab practical (and oral defense for some)
 - can bring laboratory notebook – including Day 1 primer design printout
 - *change*: CAN bring pre-lab notes also
- OH next week: Mon 12:30-1:30, Tue 4-5 pm
- Vacuum aspirators contain bleach for biohazardous waste (i.e., cells)
 - after bleach treatment, these go down the sink
- Chemical waste and sink-safe chemicals (w/out cells) should NOT be aspirated
 - the former is a safety risk, the latter just a hassle

Extracting DNA from XL1-Blue

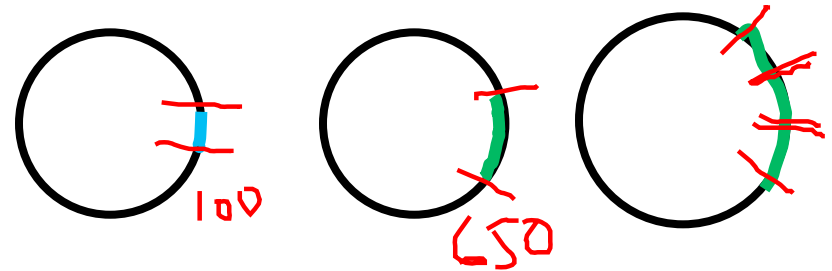
Step	Contains	Purpose
Prepare	EDTA Buffer, glucose	- weakens cell envelope - keep otherwise stable
Lyse	SDS Na^+ NaOH	- disrupt/solubilize lipid membranes, proteins - denatures ds → ss DNA
Neutralize	Acetic acid/KAc	neutralize pH, precipitate SDS <div style="display: flex; align-items: center;"> <div style="margin-right: 20px;"> $\text{O} \rightarrow \text{O}$ mixture </div> <div> SDS genomic DNA washes out </div> </div>
Transfer	N/A	use supernatant
Wash, collect	A) EtOH B) dry, water	- precipitates xDNA - EtOH intensifies w/ digest

Diagnostic DNA gels

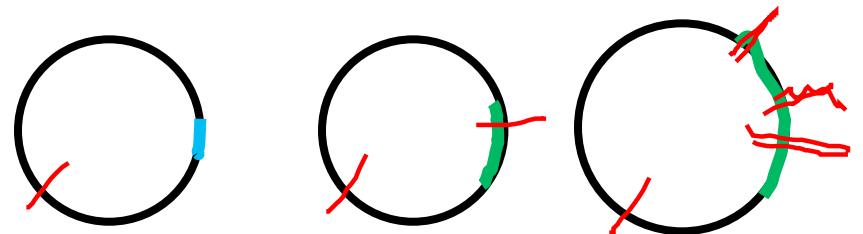


Choosing restriction sites for digest

XbaI
EcoRI



BamHI
XhoI



Practice tissue culture (TC)

- MES = murine embryonic stem cells
- Adherent cells
- Add trypsin to remove from dish
- Re-plate at lower density

→ “passage” cells

- Practice counting
- More about mammalian cells next time (review)

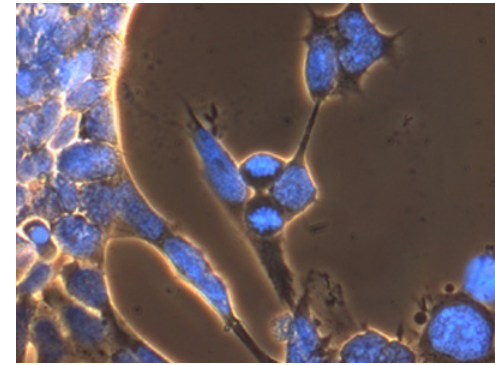


Image from http://www.stemcellresources.org/library_images.html

Today in Lab (M1D5)

- Miniprep three $\Delta 5$ -EGFP candidates, and bacteria transformed with pCX-NNX
 - tip: orient tubes in centrifuge
 - pCX-NNX = control for *technique*
- Set up digests
 - tip: make reaction cocktail \rightarrow efficiency
 - add loading dye before leaving lab
- Count and post colony #s on M1D5 *Talk* page
 - we will discuss briefly before heading to TC
- TC practice session (all together)
 - don't need notebook, just a piece of scrap paper

Interpreting your ligation results

Consider...

- Why might some groups not have gotten many exptl colonies?
- What does the *no ligase* vs. the *no insert* sample control for? Which one do you expect to have more colonies?
- How big is the variation for identical samples, and what are the possible sources of error causing the variation?