

# Consider coming to lab today/Monday if you think you'll work with mammalian cells

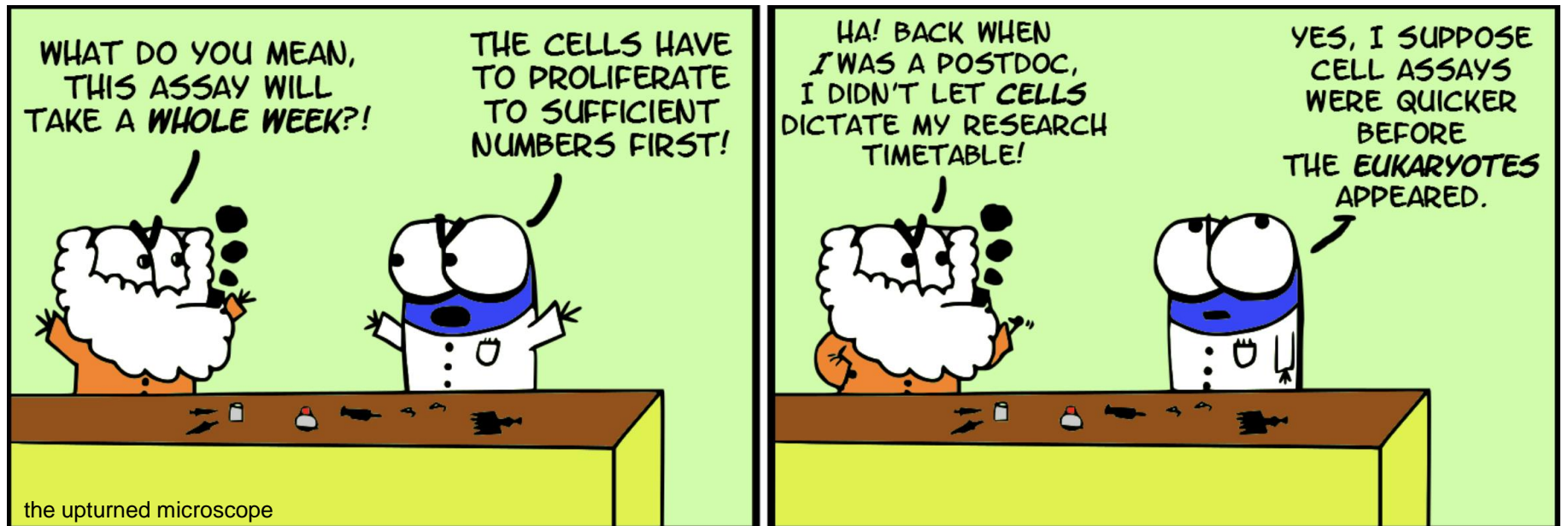
Lab is a safe space for learning what can be pretty anxiety-inducing techniques

- Sterile mammalian cell culture technique
- Working with a benchtop microscope
- Cell viability assays



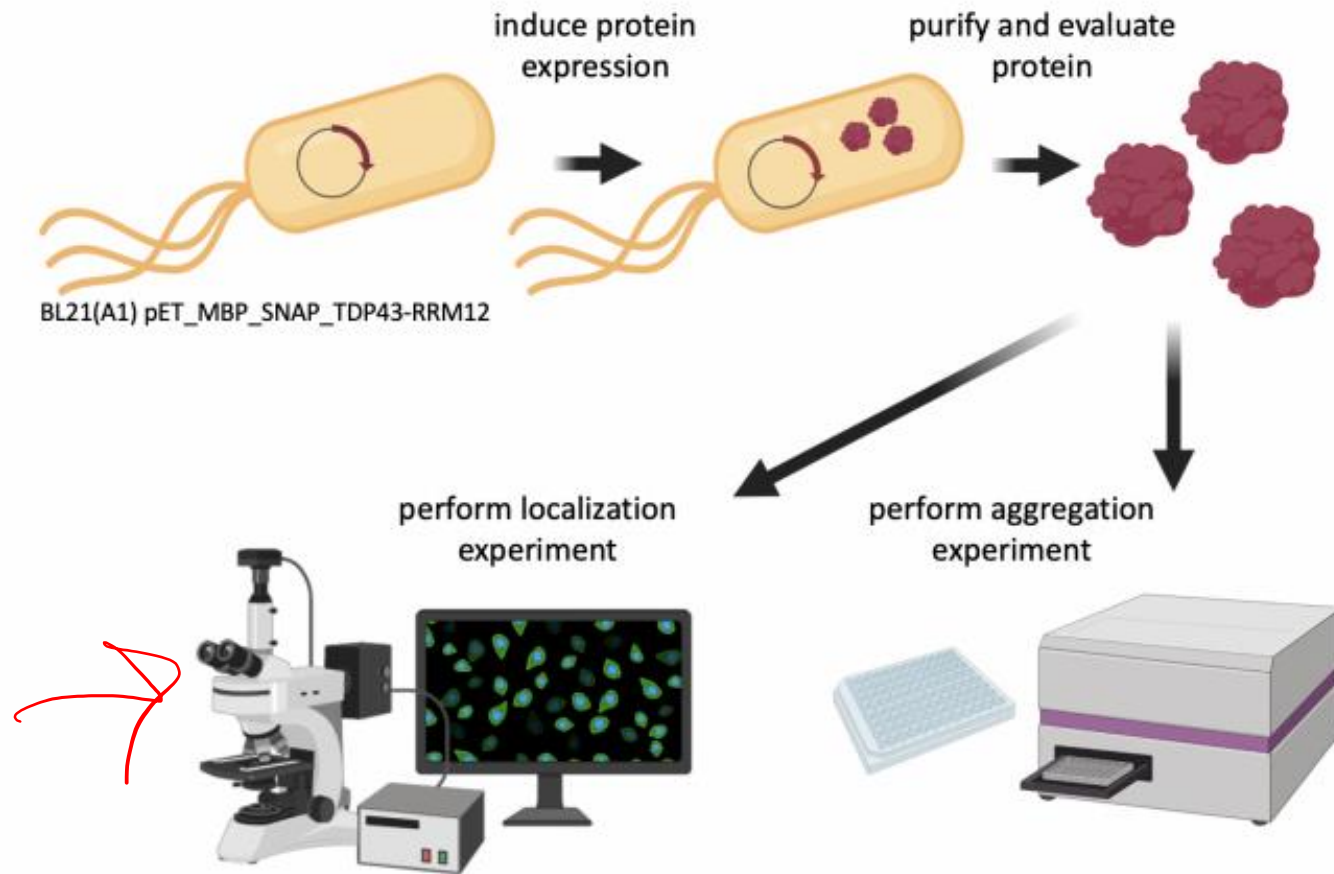
# M1D6: Learn best practices for mammalian cell culture and seed CAD cells for TDP43-localization experiment

1. Prelab discussion
2. Learn about cell culture in the lab
3. Research CAD cell line



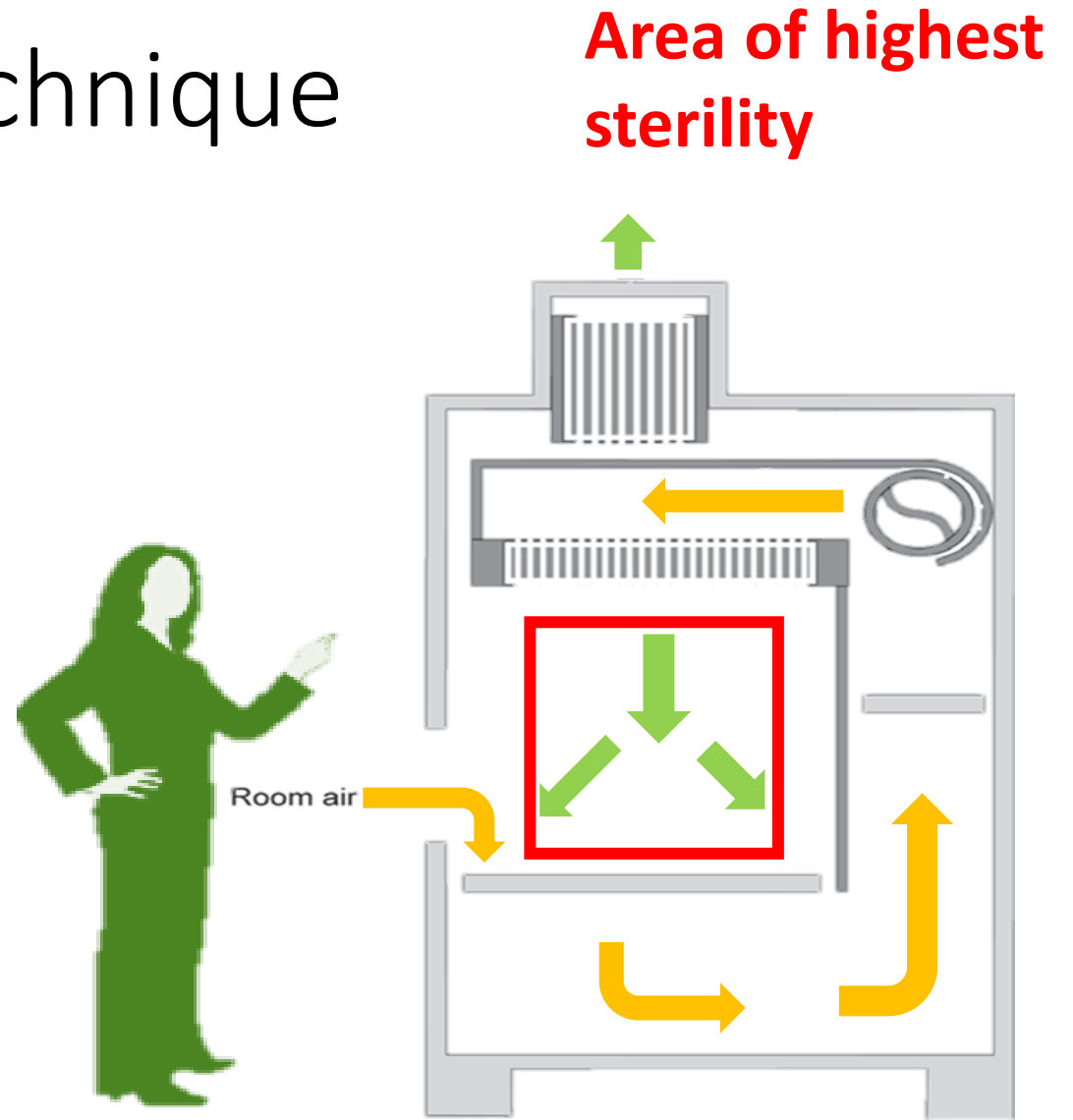
# Overview of Mod 1 experiments

**Research goal: Use functional assays to characterize ligands identified as binders to TDP43 from SMM technology**



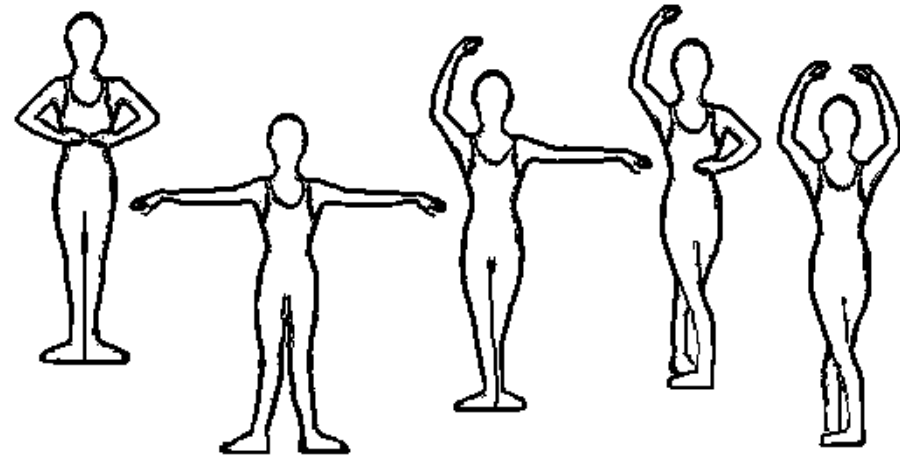
# Tissue culture sterile technique

- **70% ethanol** everything:
  - Wipe cabinet before and after use
  - Wipe everything that enters the cabinet
  - Do not spray cells with EtOH
- **Do not disturb air flow:**
  - Do not block grille or slots
  - Minimize side-to-side arm movements
  - Work > 6" away from sash
  - Leave blower *on always*
- Do not talk into incubator!
- Only open sterile media and pipettes in hood



# Sterile Technique, condensed

- 1) Ethanol everything (minus anything containing cells)
  - 1) Gloved hands, unopened plastic pipettes, autoclaved containers, media bottles.... EVERYTHING
- 2) Move *slowly*
- 3) Don't block the grate



# Mammalian Cell Culture Medium – CAD formulation



## Food:

- DMEM/F12 (Dulbecco's Modified Eagle Medium/Nutrient Mixture F-12)
  - **Defined**
    - Salts, H<sub>2</sub>O, Sugar, Amino Acids
    - Phenol Red (Purple – Old, Orangey-Red – OK, Yellow – Bad)
- FBS (fetal bovine serum)
  - **Undefined**
    - Lot Dependent
    - Lipids, cytokines, growth factors, nucleic acids



## Non-food:

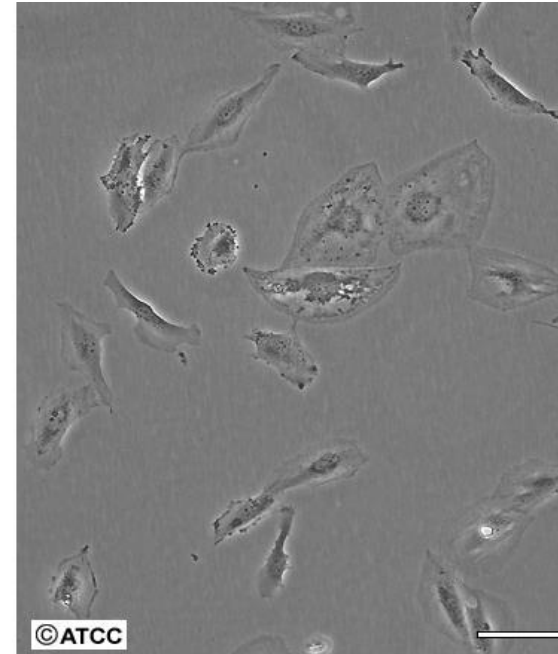
- antibiotics:
  - penicillin
  - streptomycin



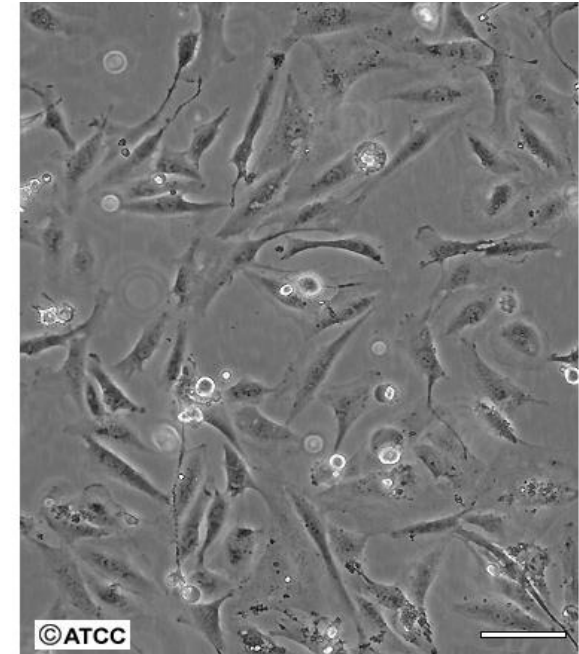
# Mammalian Cell Culture Terminology

- Adherent vs Suspension
  - Adherent – Cells that like to stick to flask/coverslip surface. Flattened.
  - Suspension – Spheroid cells that float in solution
- Confluence
  - % surface covered in cells
- Splitting
  - Taking one culture's cells and passing to more flasks to either expand or maintain cell culture
- Seeding
  - Splitting with a known number of cells to achieve a desired confluence/density

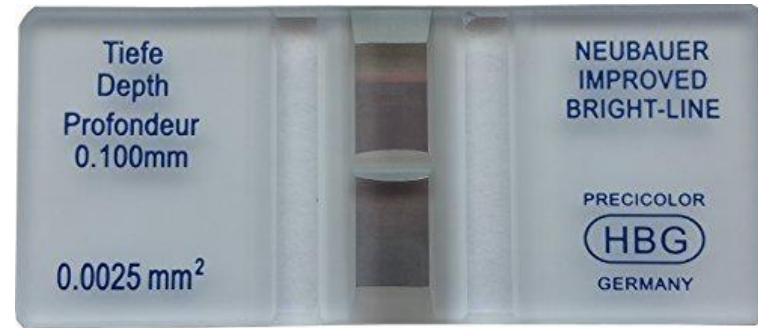
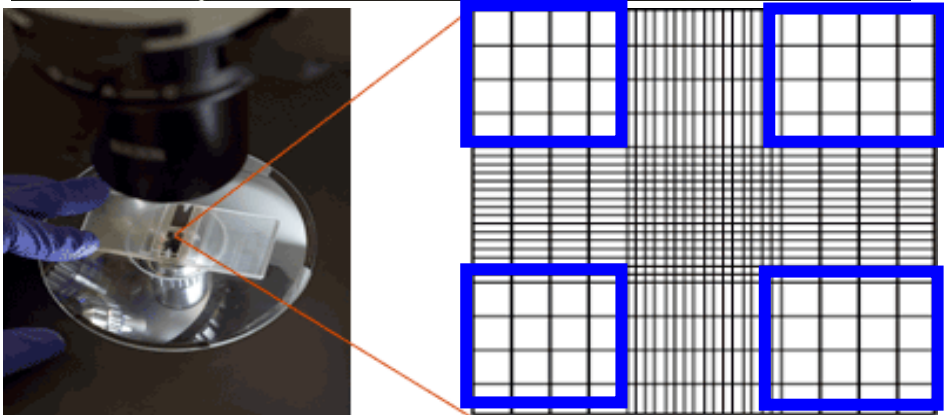
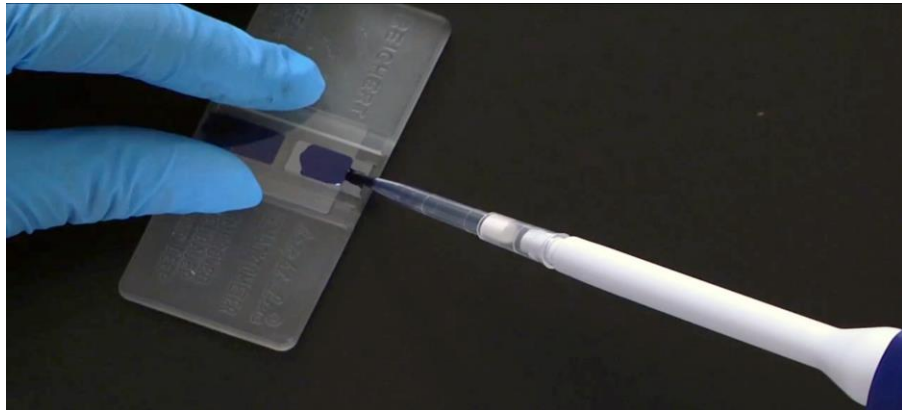
Low Density



High Density



# Counting cells



- Hemocytometer
- Trypan blue

# cells / mL = 10,000 x  
average of 4 corners



# Trypan Blue is an Exclusion Dye

- 1) TB is toxic, viable cells pump the dye out
- 2) Count clear cells with visible light halos

