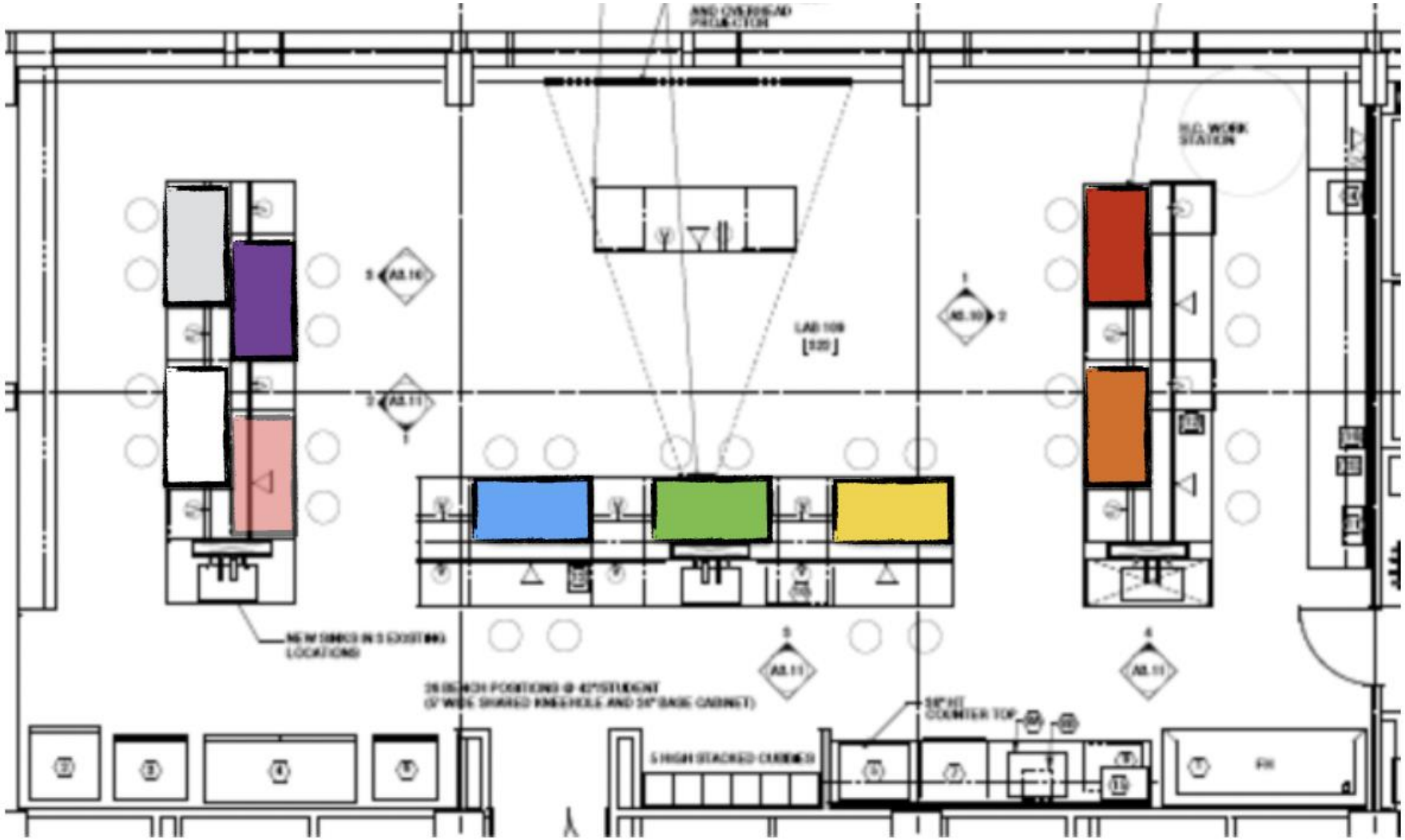
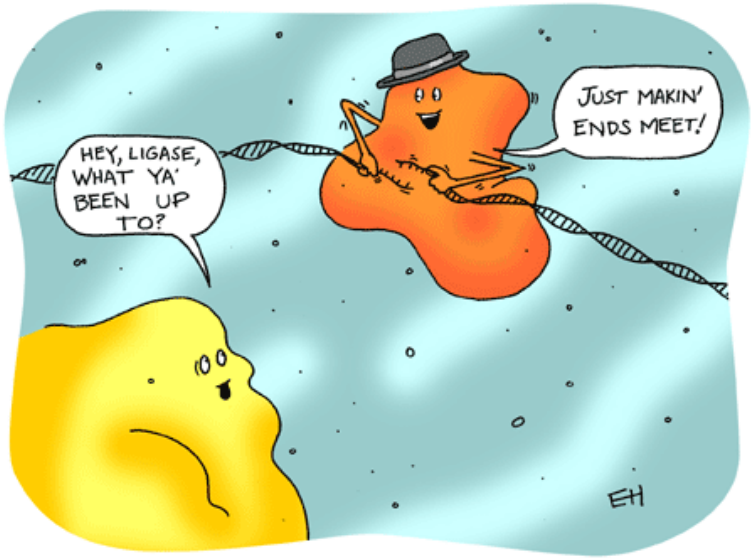
Partner and bench assignments



Red Team:
Tilly & Arbri
Orange Team:
Jake & Alex
Yellow Team:
Katia & Logan
Green Team:
Tim & Andrew
Blue Team:
Vivian, Josh, Charles
Pink Team:
Kim & Jalen
Purple Team:
Nicole & Anika

M1D1:
Complete *in silico* cloning of protein expression plasmid

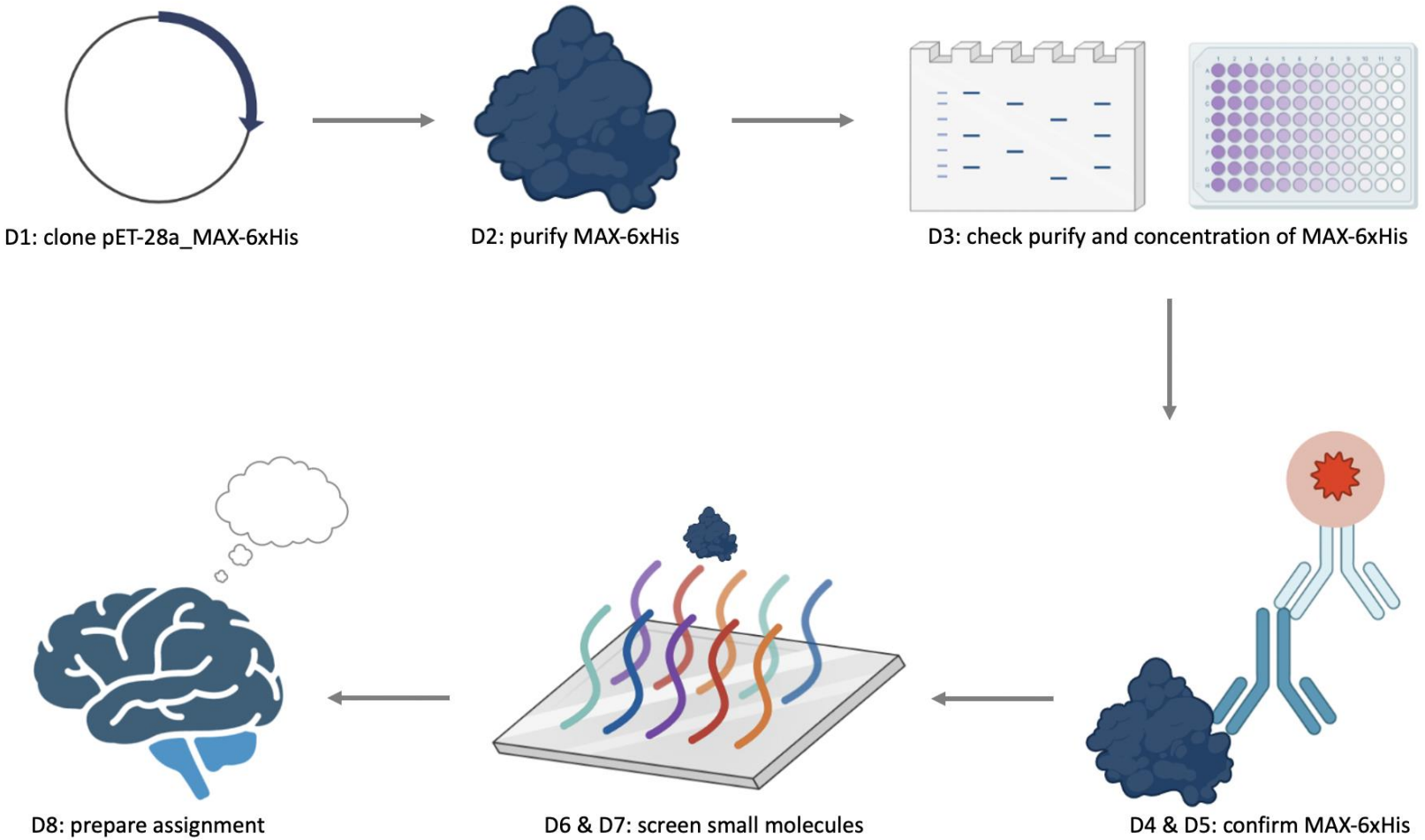
1. Laboratory Orientation quiz
2. Prelab discussion
3. Build protein expression plasmid - virtually
4. Confirm protein expression plasmid - actually



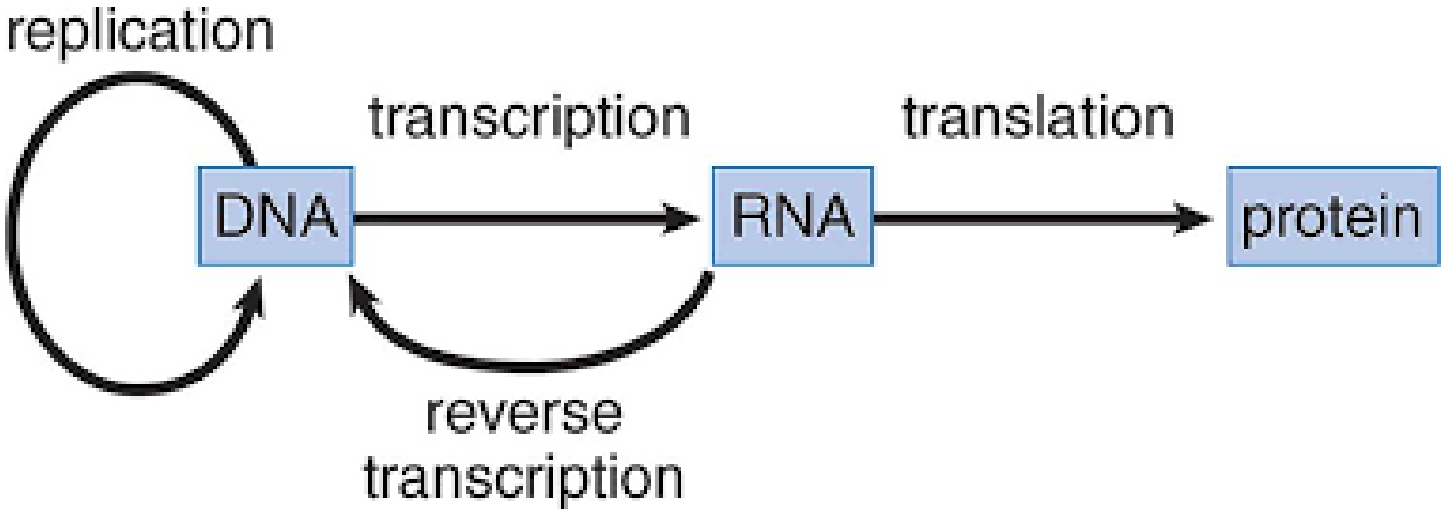
Mark your calendars!

- Data summary (15%)
  - completed in teams and submitted via Canvas
  - draft due 3/18, final revision due 3/27
- Research talk (5%)
  - completed individually and submitted via Gmail
  - due 3/4
- Laboratory quizzes
  - scheduled for M1D4 and M1D8
- Notebook (5% and part of 5% Participation score)
  - one entry submitted via Canvas 24 hr after M1D8
- Blogpost (part of 5% Participation score)
  - due 3/20 via Slack

Overview of Mod 1 experiments:

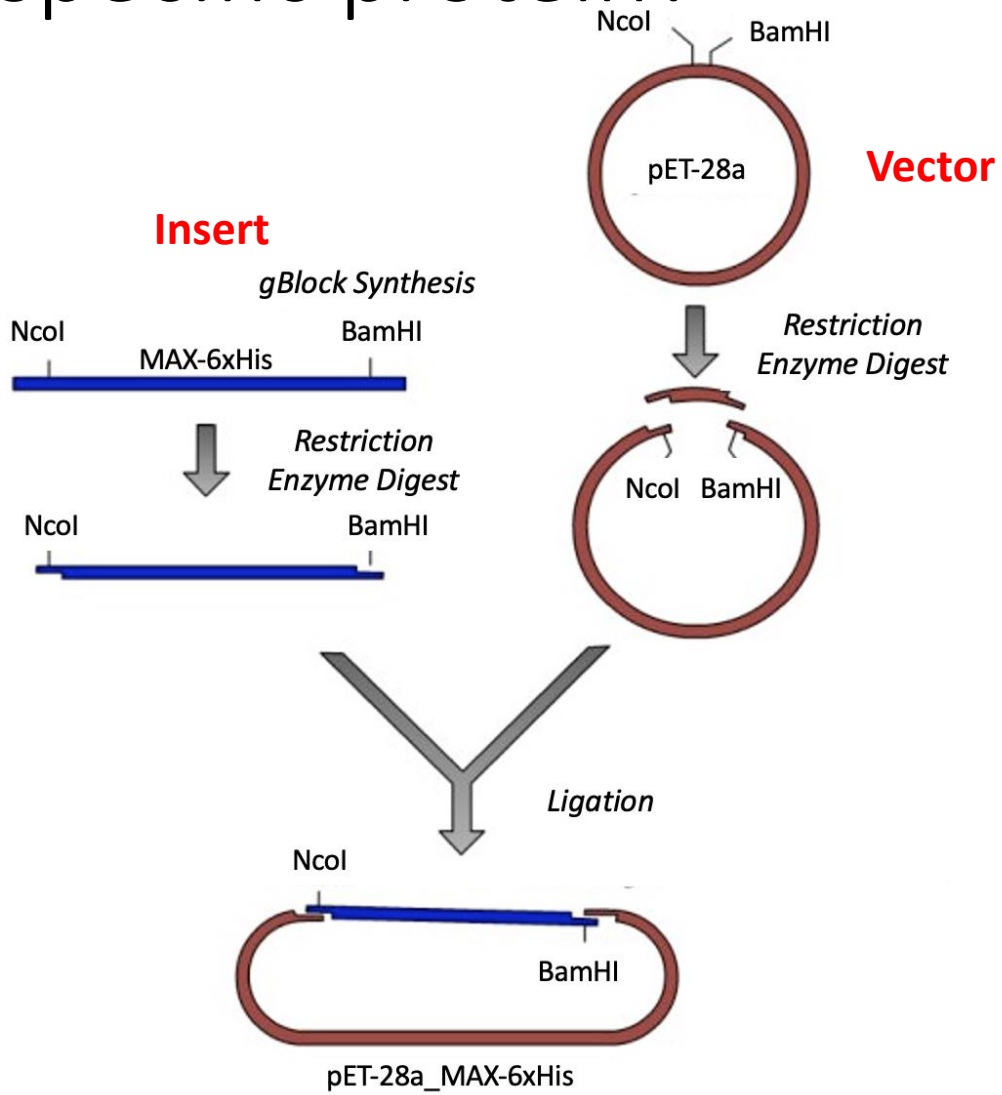


It all starts with DNA…



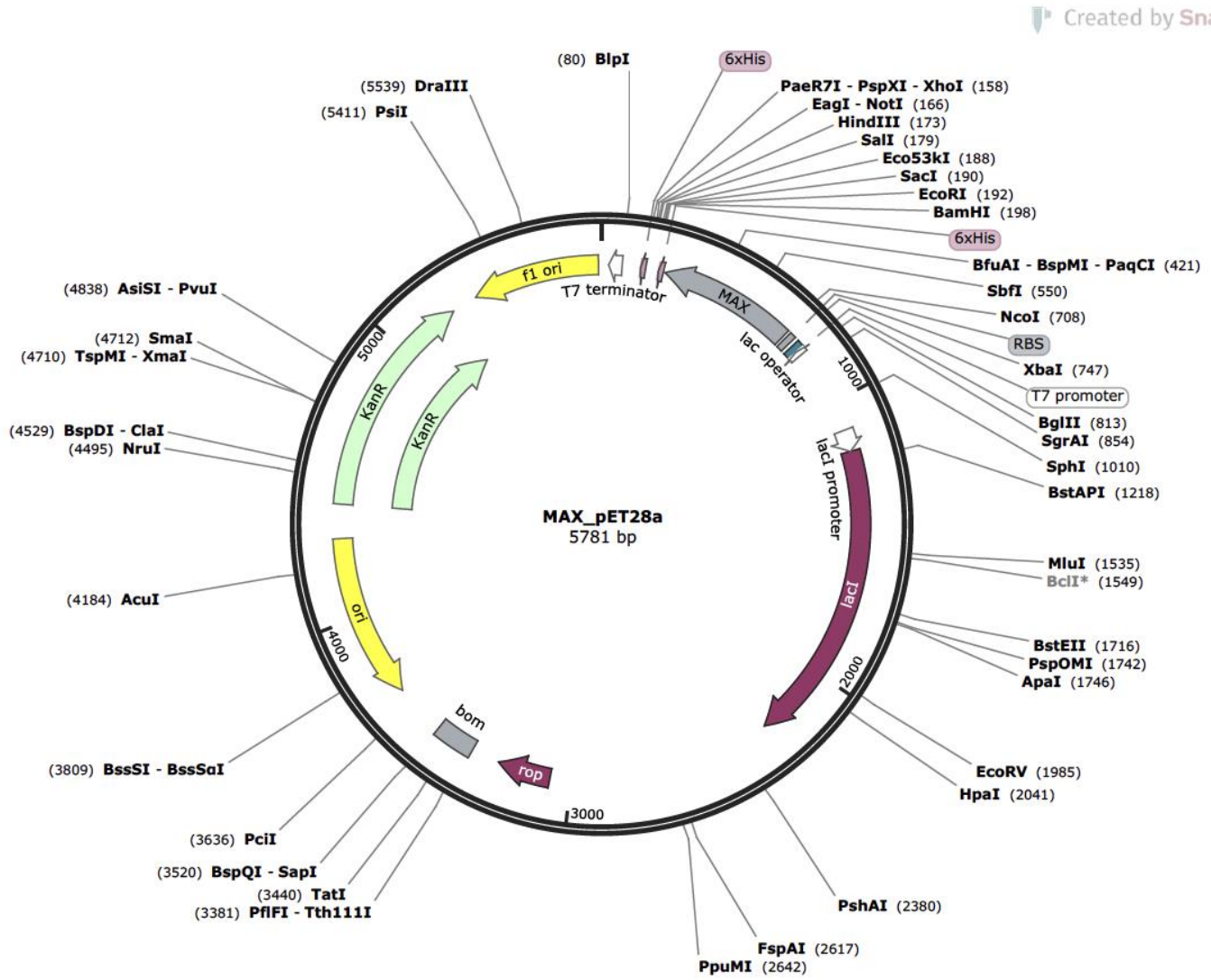
What if we want to make a specific protein?

- Who are the players?
  - Insert GOI (Gene of interest)– **MAX-6xHis**
  - Vector Other essential stuff
- What is the process?
  - Digestion Cut DNA with Restriction Enzymes
  - Ligation Staples the insert DNA into the open plasmid

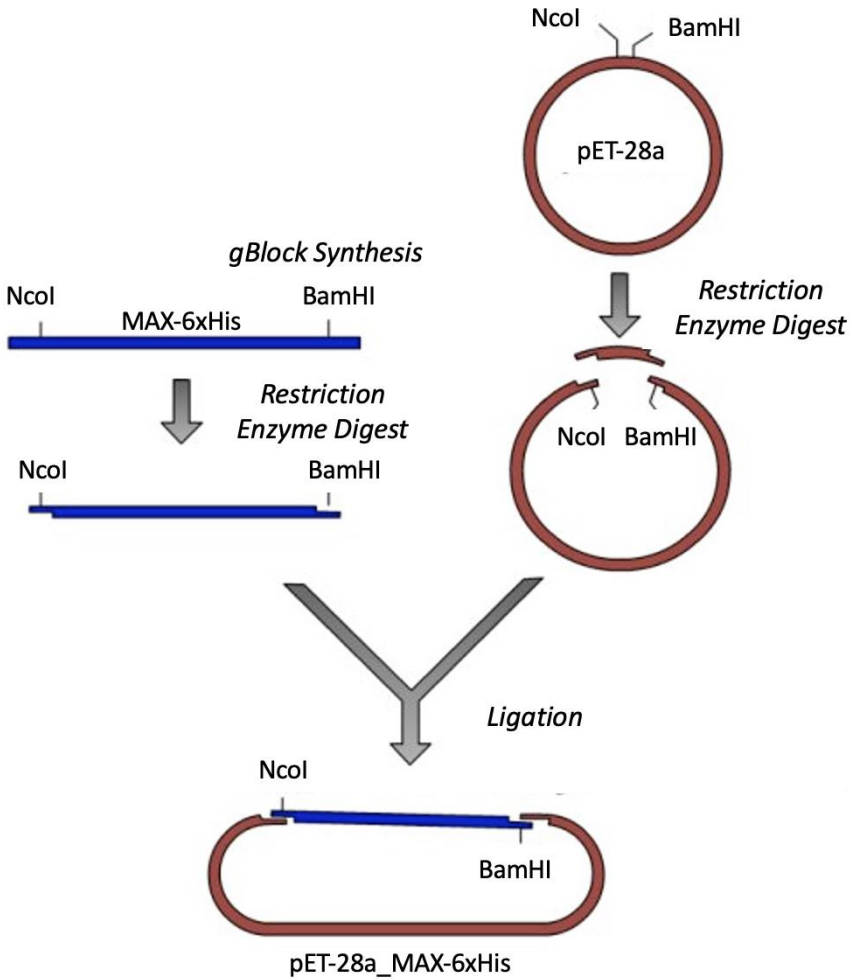


What's in a vector?

1) ORI – (Origin of Replication)
2) Promotors
3) RE sites
4) Antibiotic Resistance Gene

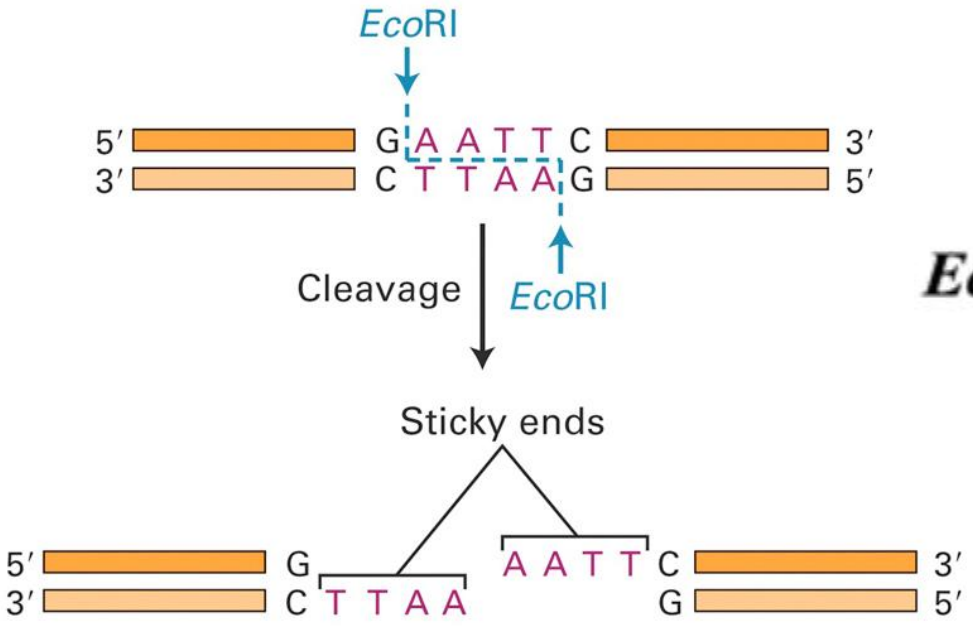


1. Prepare insert by designing gBlock



2. Digest insert and vector using restriction enzymes

Why are most restriction sites palindromes?

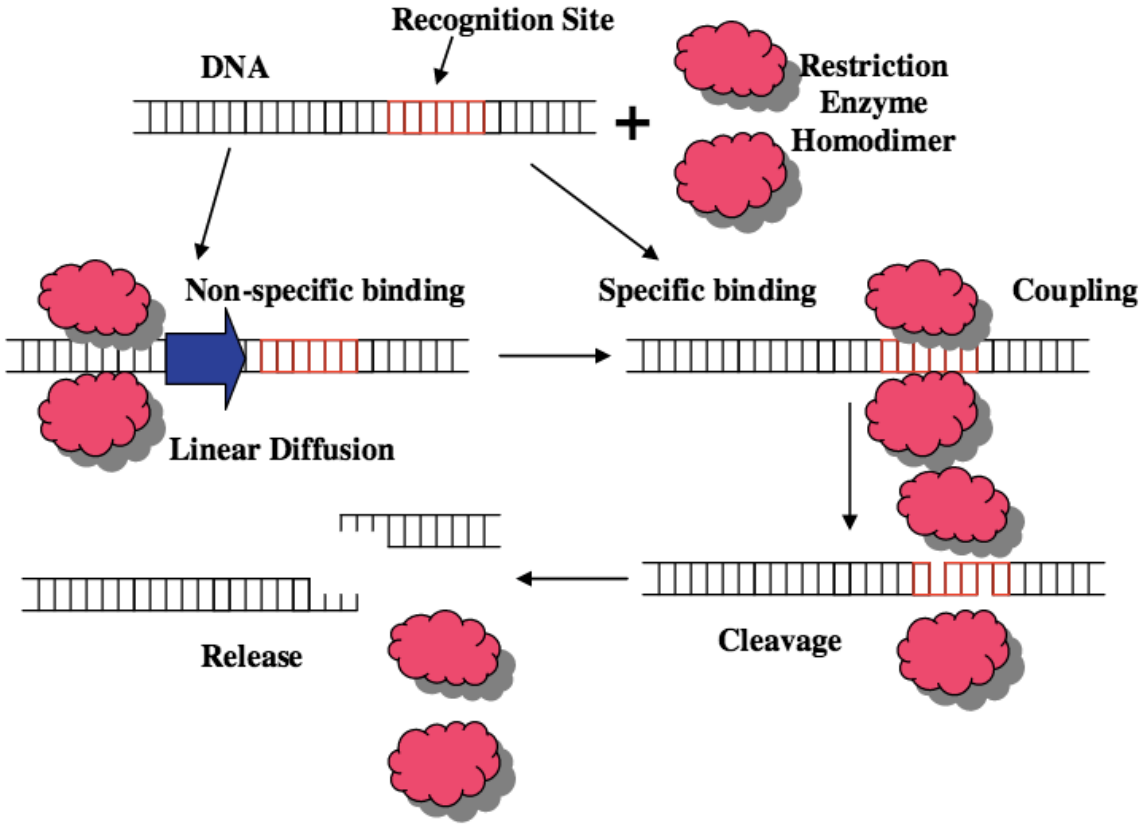


Restriction enzymes cleave DNA at specific sequences

Function as homodimers

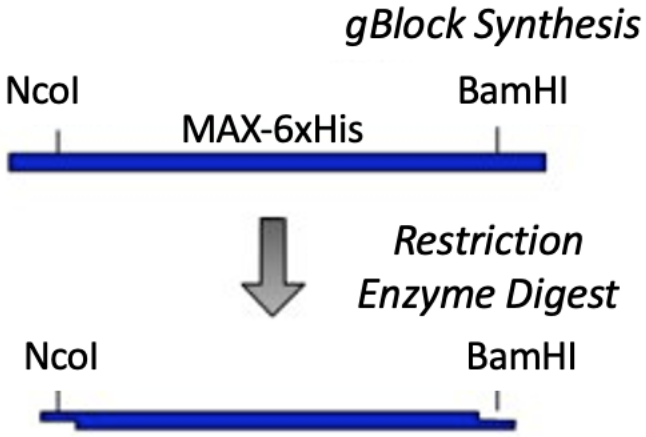
- Each dimer cleaves backbone at site of palindromic recognition sequence

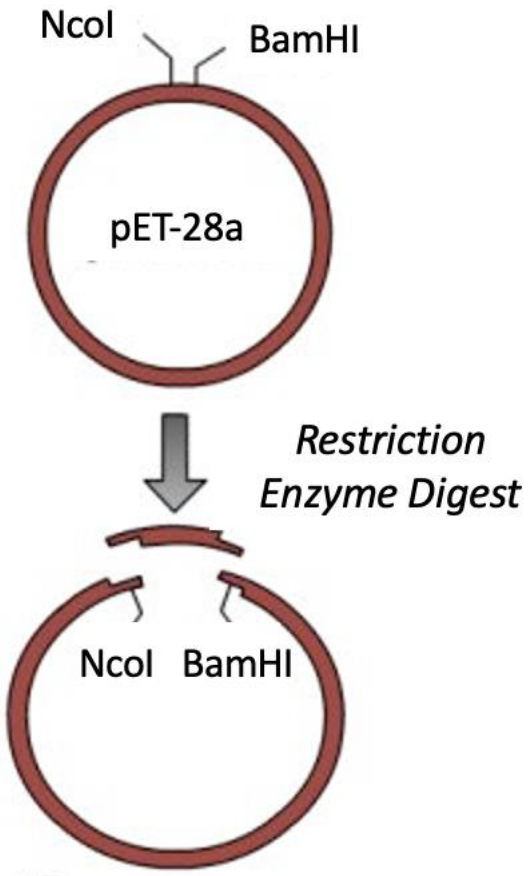
How do restriction enzymes recognize DNA sequences?



Insert & Vector should be digested using the same enzymes

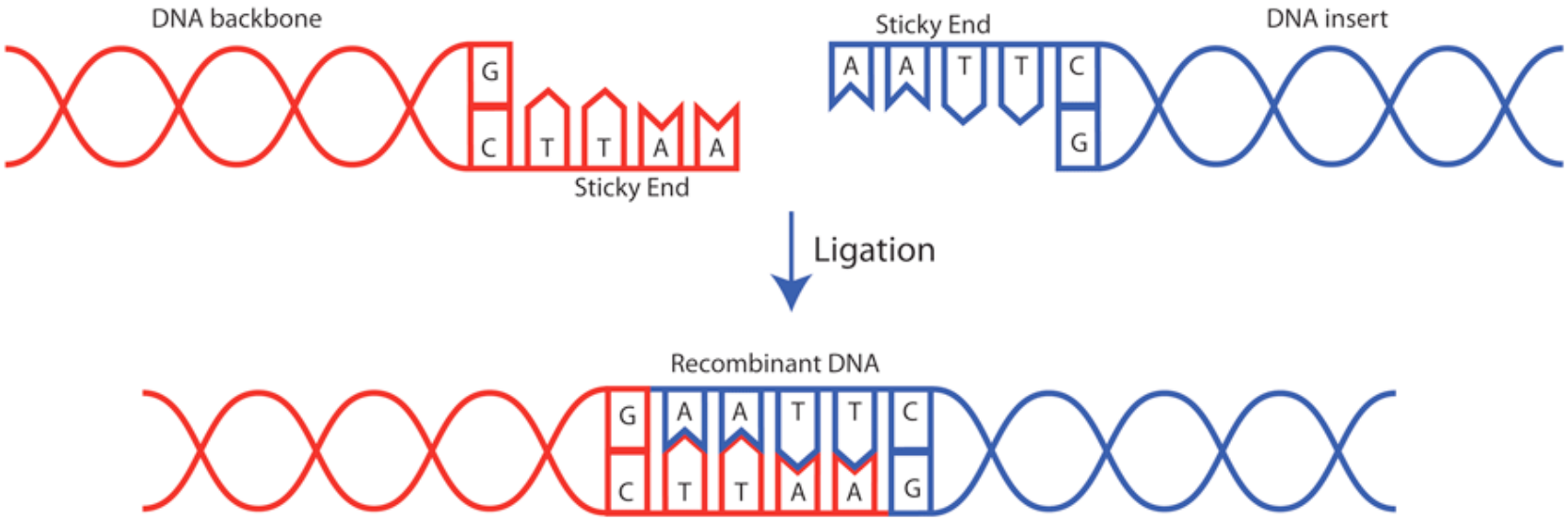
gBlocks allow you to add on whatever RE sites you want – very handy!





3. Ligate insert into vector using ligase enzyme

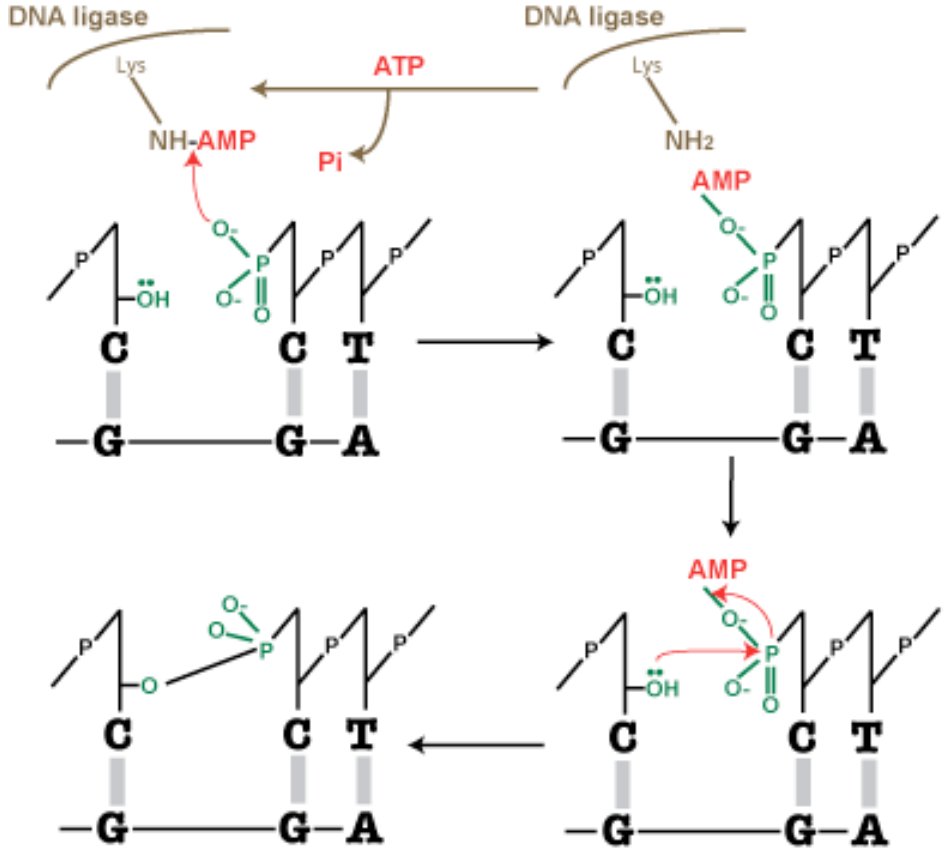
How are compatible DNA ends paired prior to ligation?



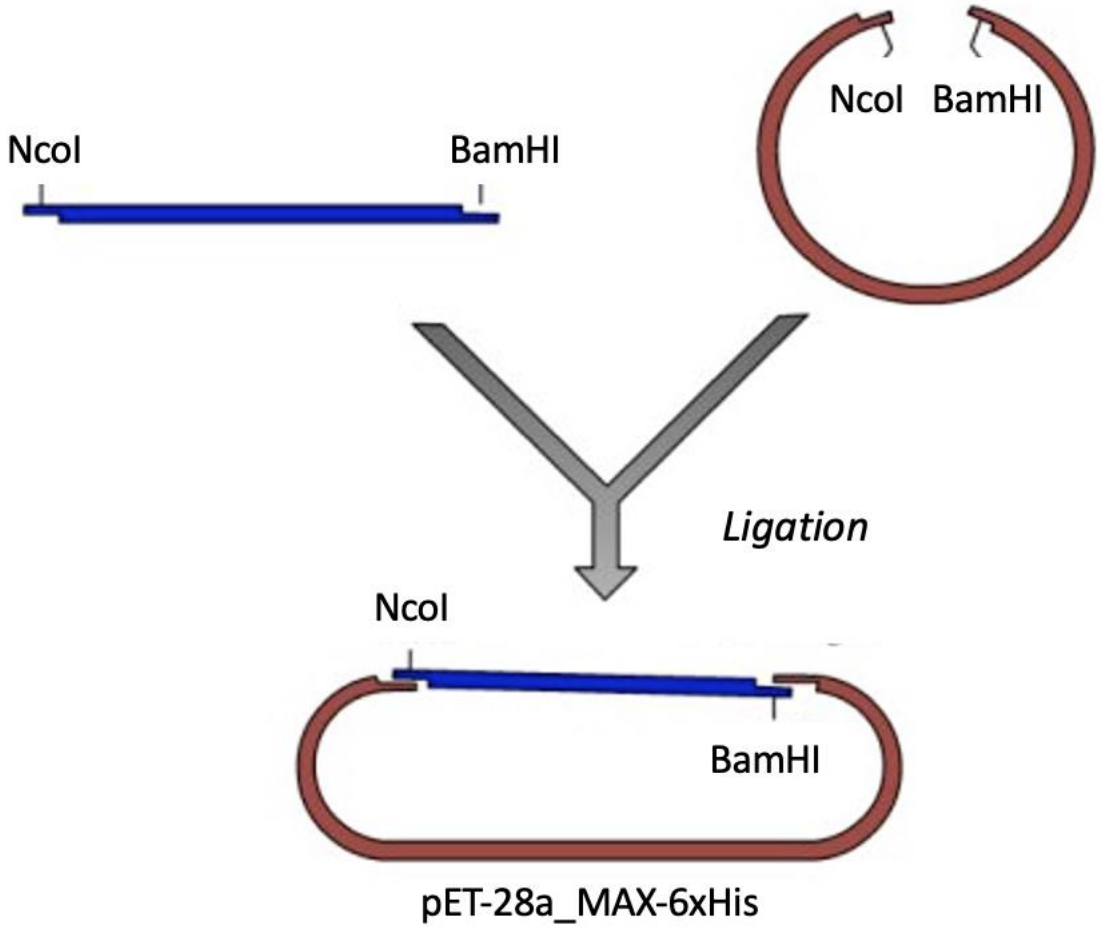
DNA ligase catalyzes formation of covalent phosphodiester bond

DNA ends from 3’ OH acceptor and 5’ phosphate donor joined

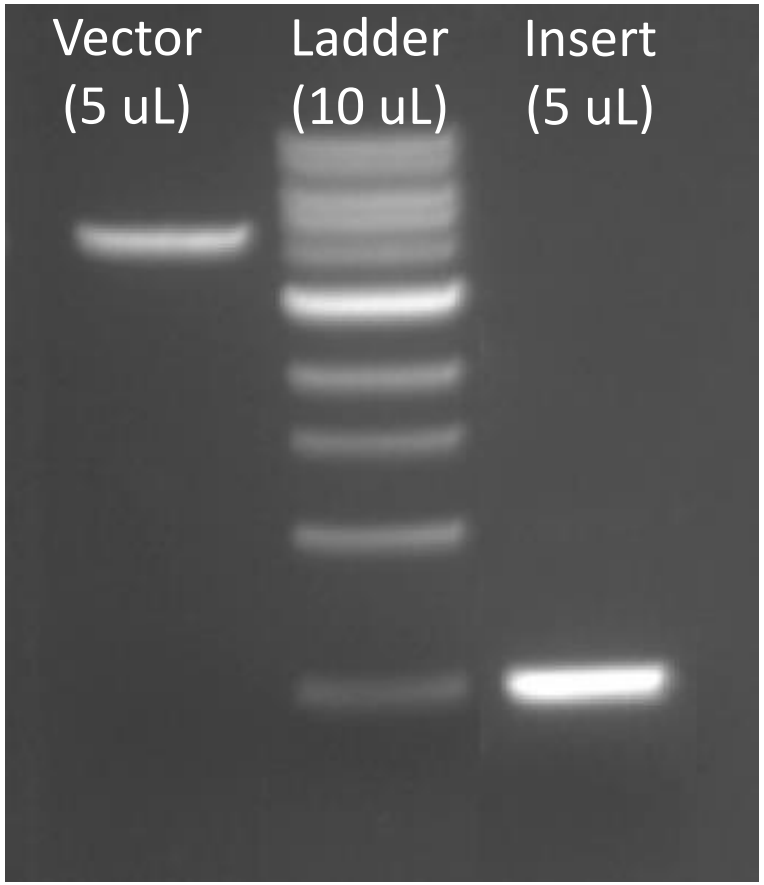
- Requires ATP for adenylation of lysine residue in active site of DNA ligase
- AMP then transferred to 5’ phosphate of DNA base

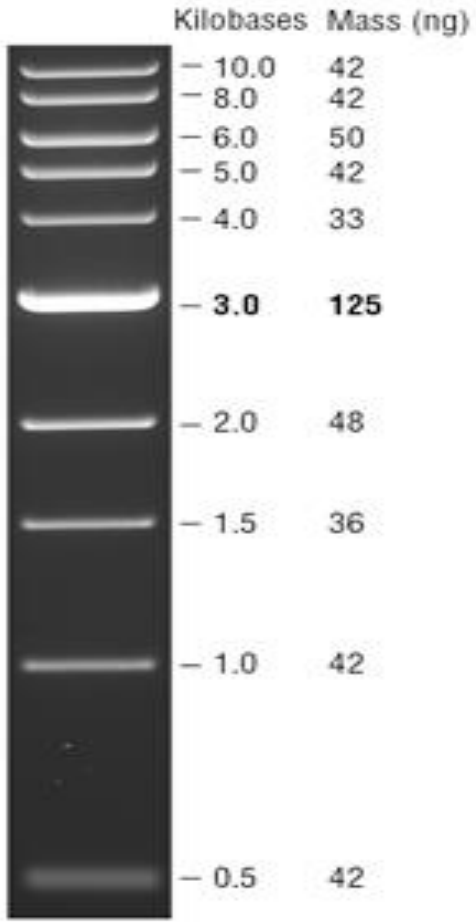


What are we ligating?



How are ligation reactions prepared?





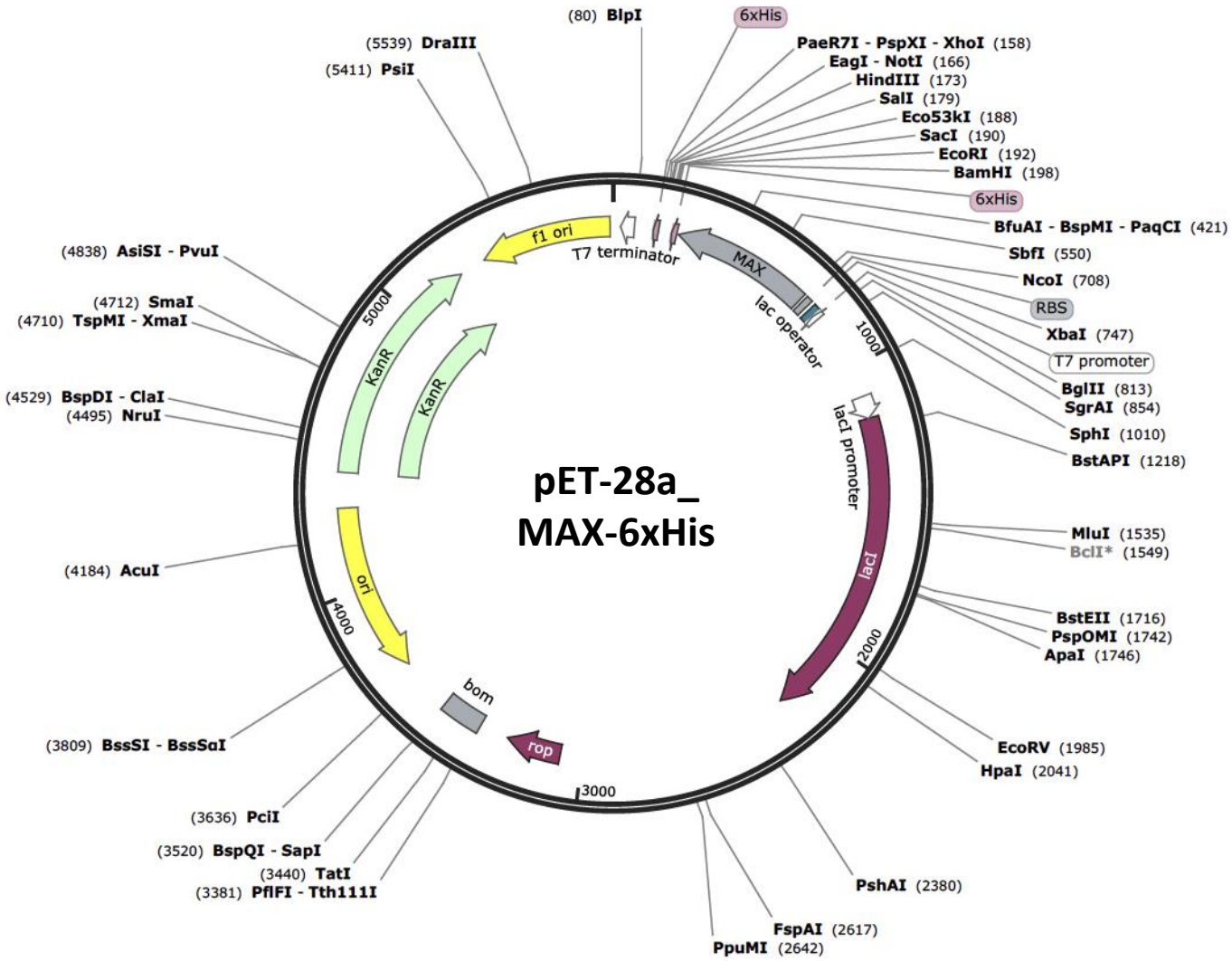
- Ideally, want 3:1 molar ratio of insert:backbone
- Calculate molar amounts from concentrations and sizes of DNA molecules

Ligation calculations

1. Determine volume of backbone
   - Use backbone concentration = [provided in exercise] ng/uL
   - Want 50 – 100 ng
2. Calculate moles of backbone
   - Vector = [determined during exercise] bp, MW bp = 660 g/mol
3. Calculate moles of insert
   - Insert = [determined during exercise] bp, 3:1 ratio of insert:backbone
4. Calculate volume of insert
   - Use insert concentration = [provided in exercise] ng/uL

How do we confirm cloning products?

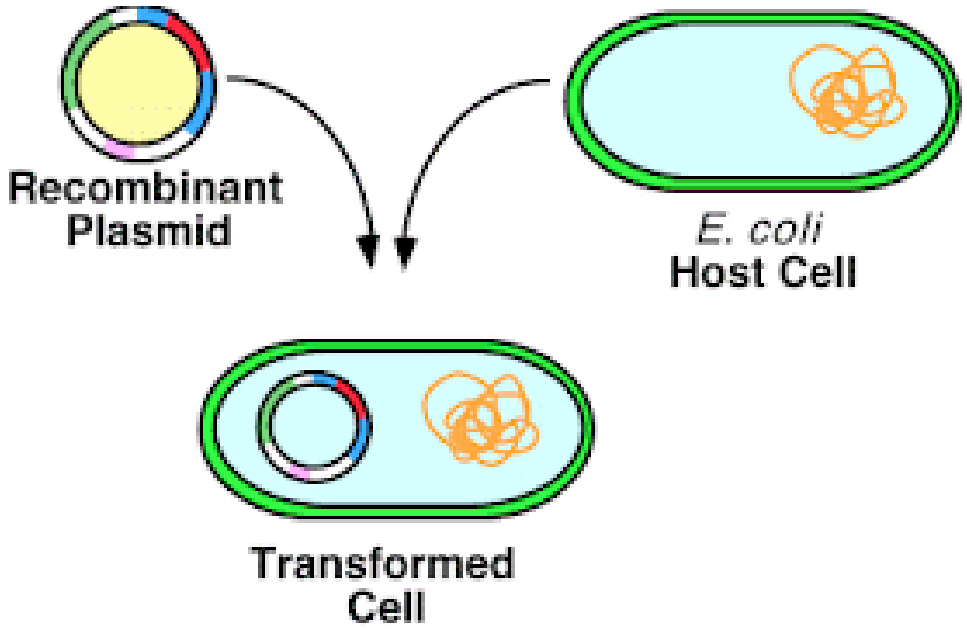
1. Transform plasmid into competent cells
2. Isolate plasmid from cultured cells
3. Digest plasmid using restriction enzymes



1. Transformation used to promote uptake of foreign DNA in bacteria

Why do we transform the ligation product?

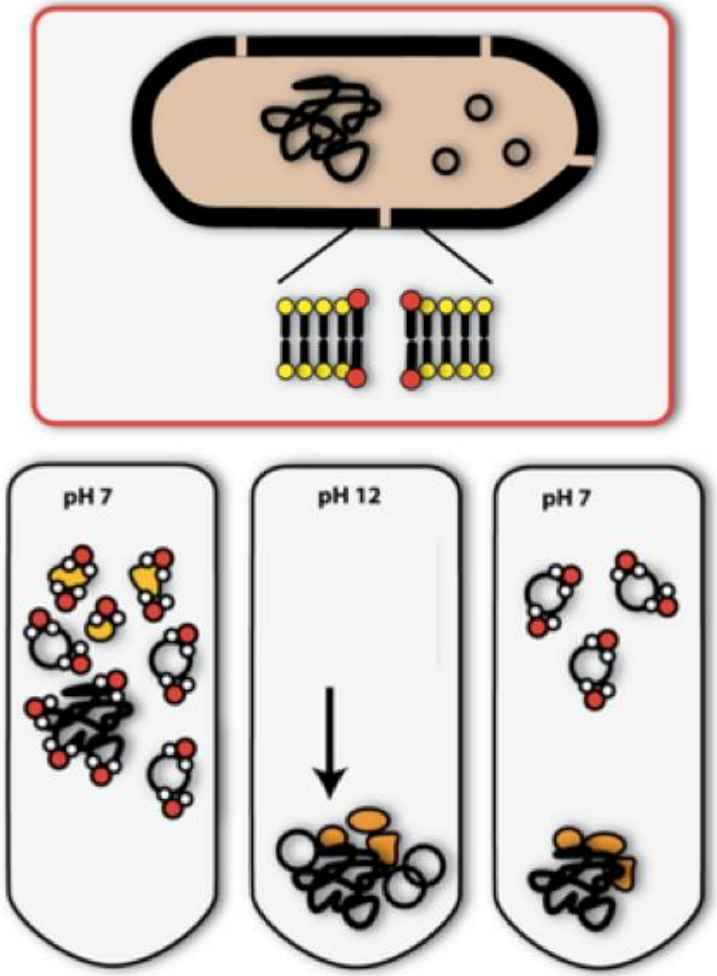
1. Incubation
2. Heat shock
   - DNA taken in by competent cells
3. Recovery
4. Selection



2. Purification used to isolate plasmid DNA

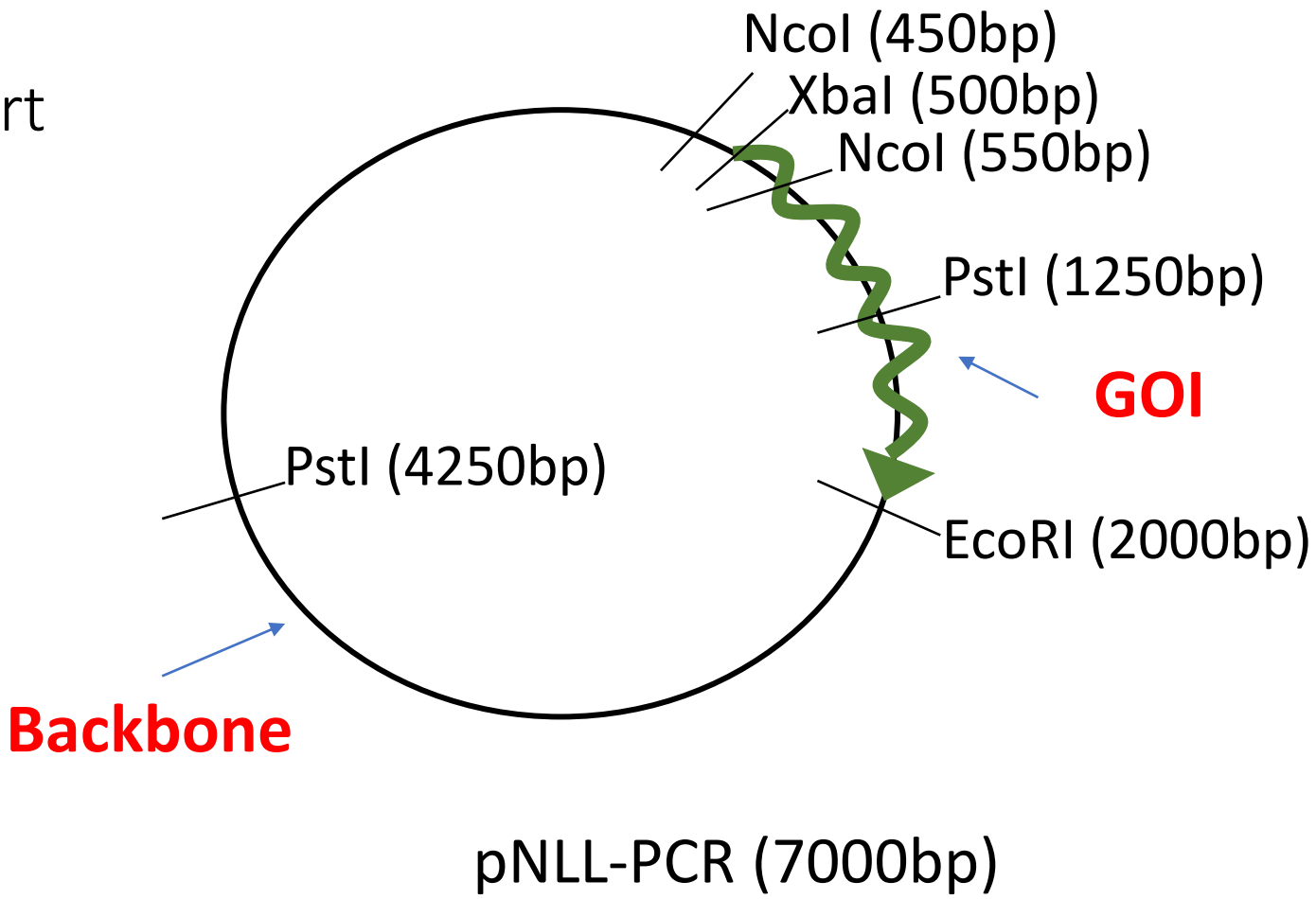
Why do we purify (mini-prep) the ligation product?

1. Resuspend cells
2. Lyse
3. Neutralize
   - Separates chromosomal DNA from plasmid DNA
4. Wash
5. Resuspend or elute DNA



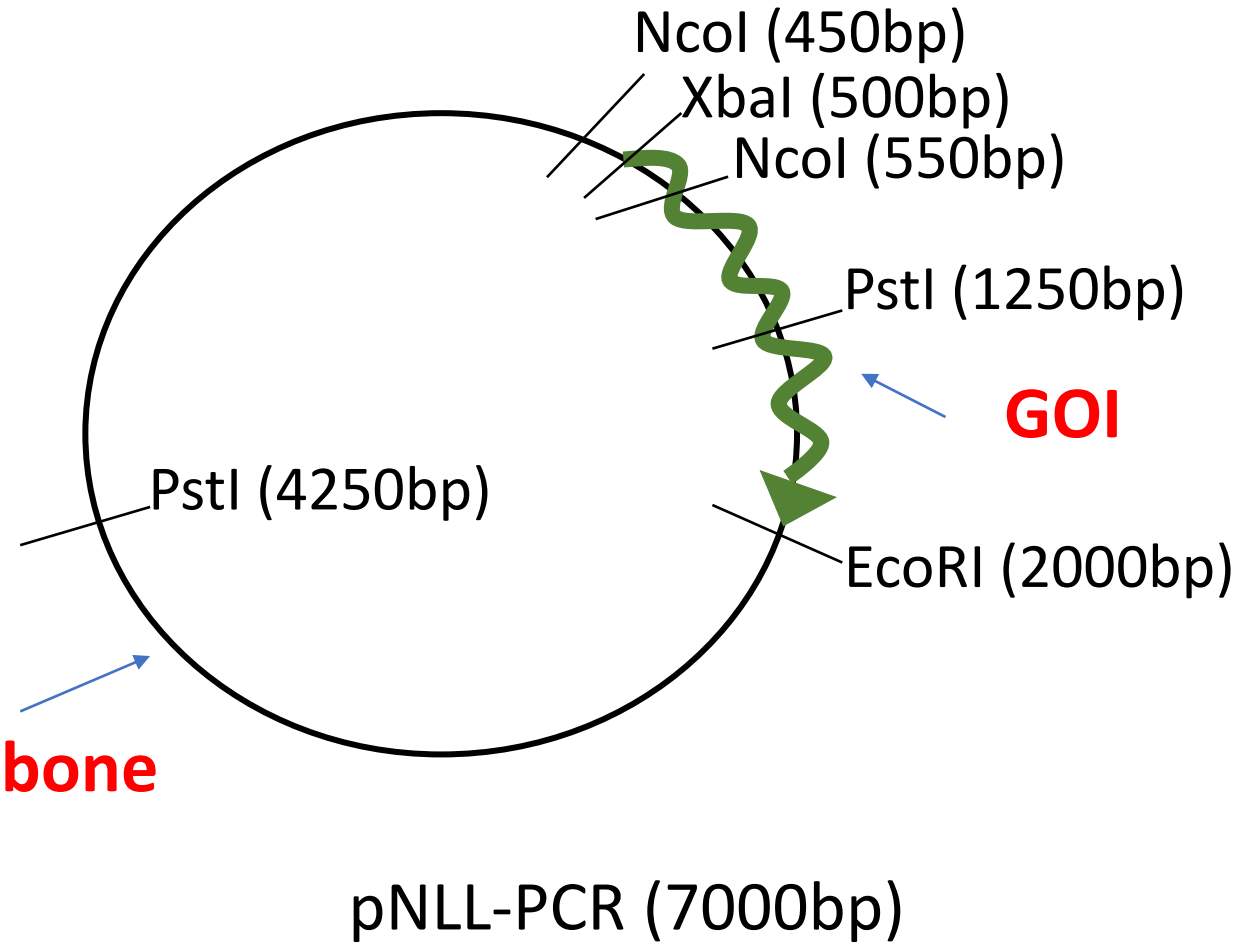
Confirmation digest follows plasmid purification

- Ideally, will cut once in insert and once in vector
  - XbaI and EcoRI?
  - PstI?
  - NcoI?



Confirmation digest follows plasmid purification

- Ideally, will cut once in insert and once in vector
  - XbaI and EcoRI?
    - Can't distinguish between chaining of multiple inserts per backbone
    - 1500 bp, 5500 bp
  - PstI?
    - Wouldn't be able to distinguish band sizes on a gel
    - Two similar frags
  - NcoI?
    - Size difference would be difficult to see
    - One frag is too tiny to resolve (~100bp)



For today…

- Virtual cloning exercise to build pET-28a_MAX-6xHis expression plasmid
- Confirmation digest of pET-28a_MAX-6xHis

For M1D2…

- Answer prompts regarding M1 background information
- Complete in class exercises from M1D1