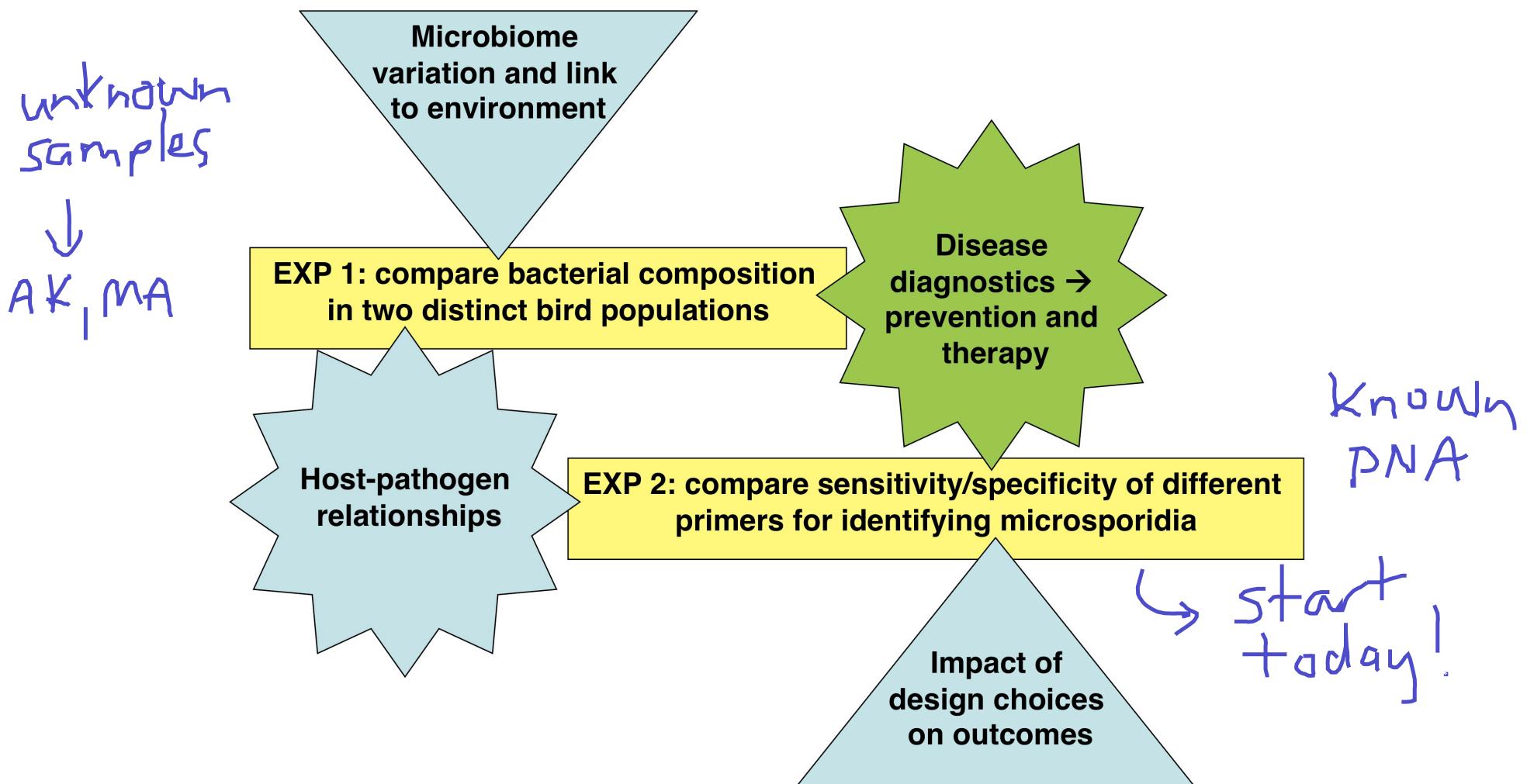


- Announcements
- Pre-lab Lecture
  - ❖ Module 1 overview +  $\mu$ sporidia
  - ❖ Intro to primers and PCR
  - ❖ Module 1 assignments
  - ❖ Today in Lab: M1D1
- Lab Practical (~40-45 min)

# Announcements

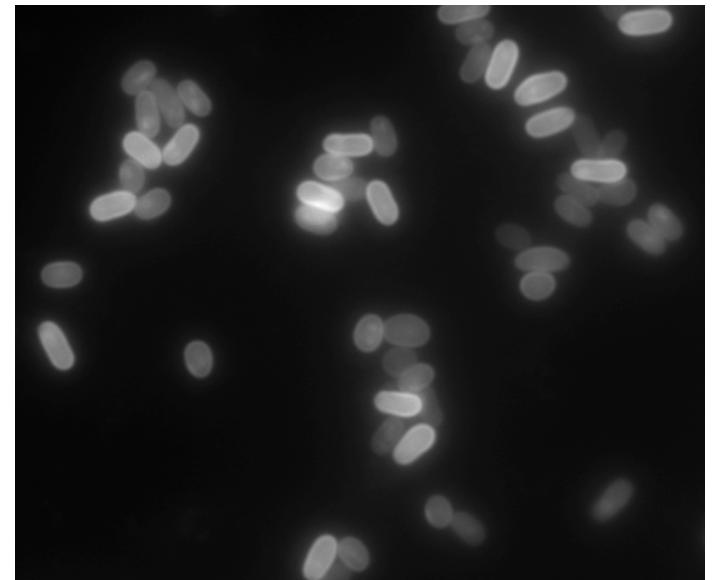
- BE (and other) seminar series:
  - Seminar posters across from BE HQ on 3<sup>rd</sup> floor
  - Full schedule linked from BE website
  - Part of professional development
- Introducing... Ian, your TA for Module 1
- Different equipment for different volumes/tasks

# Module 1 overview



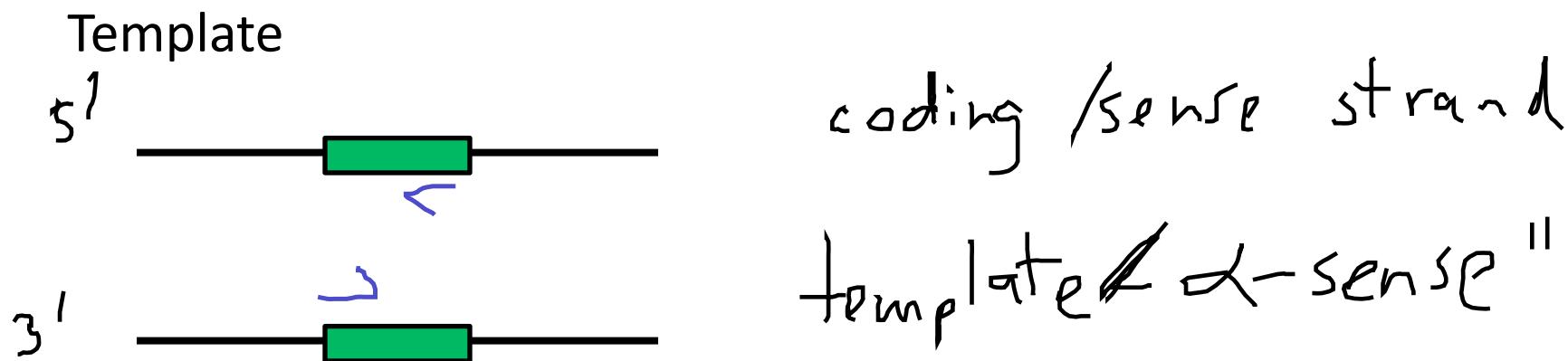
# Microsporidia: fascinating bugs

- Highly evolved fungus
  - toward simplicity
  - few genes *and* few non-coding regions for a eukaryote
  - protist? no!
- Opportunistic infections
  - immunocompromised (*HIV, chemotherapy, age*)
  - travelers
- Tough to isolate!



(HIV, chemotherapy, age)

# Designing PCR primers: topology



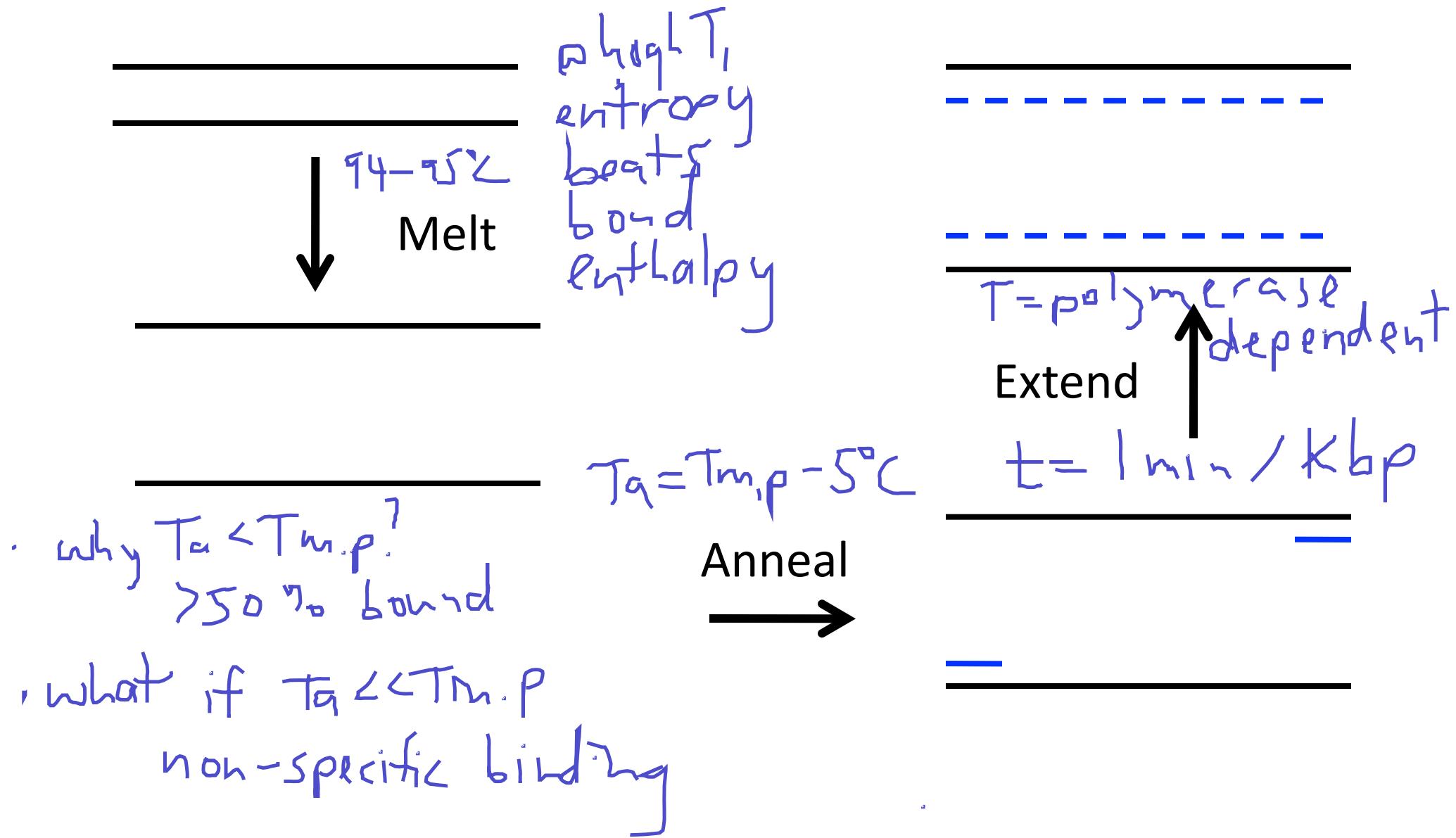
Primers

5'  $\rightarrow$  Forward { binds to d-sense  
reads as sense

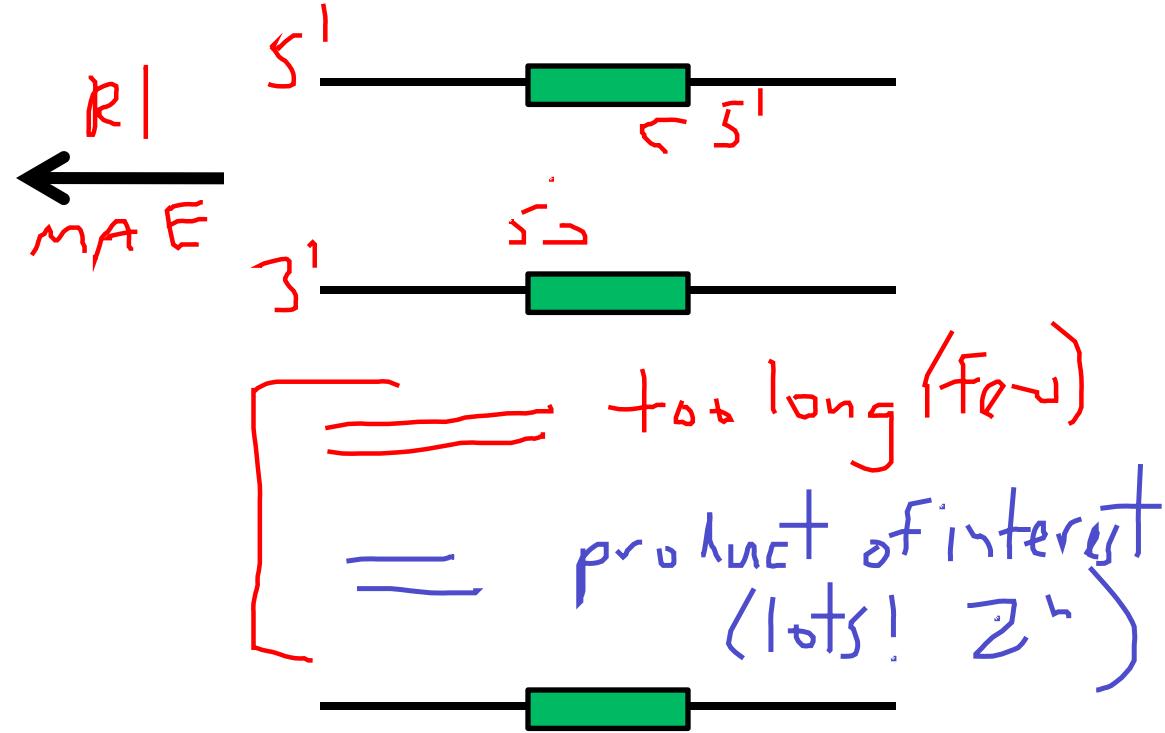
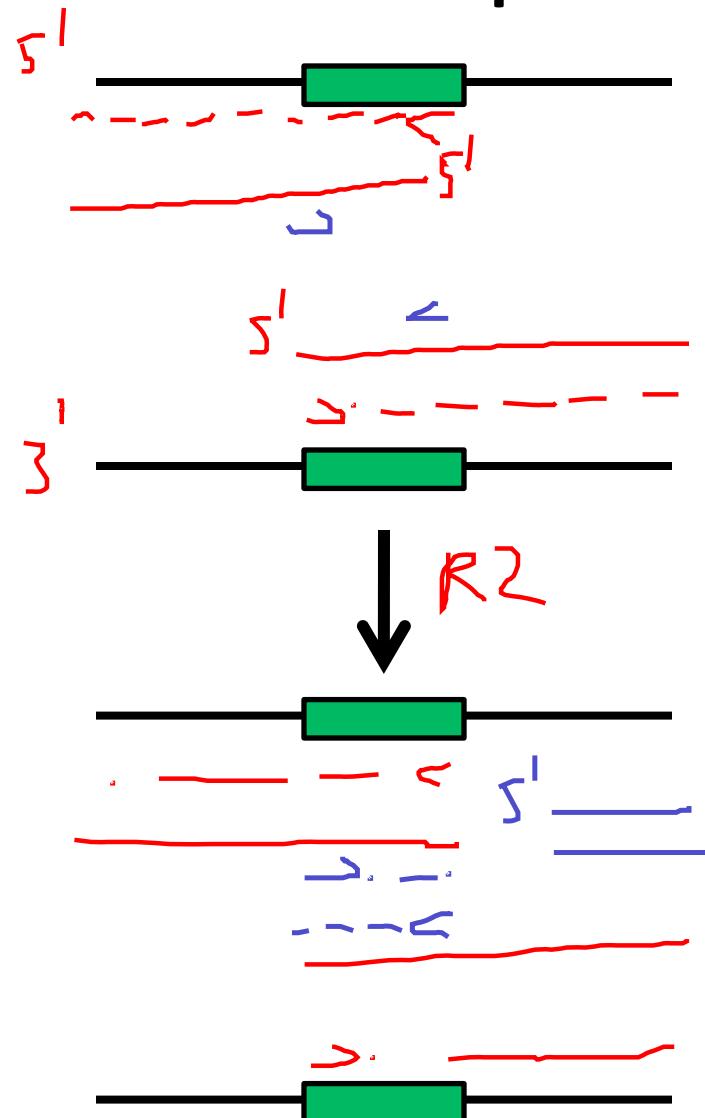
$\leftarrow$  5' Reverse binds to sense

The diagram illustrates PCR primer placement. It shows two primers: a forward primer (blue arrow pointing right) and a reverse primer (blue arrow pointing left). The forward primer is labeled "Forward" with a brace indicating it "binds to d-sense" and "reads as sense". The reverse primer is labeled "Reverse" with a brace indicating it "binds to sense". Both primers are aligned with the template strand shown in the previous diagram.

# PCR process: three TD-driven steps



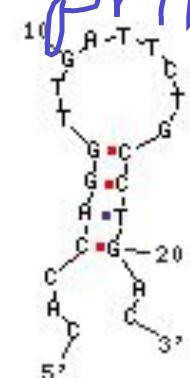
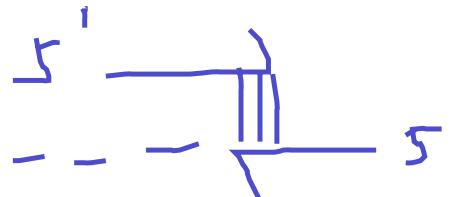
# PCR process: three rounds



# Designing PCR primers: properties

- Length: why is 17 bp the magic number?  
human genome  $\sim 3 \cdot 10^9$  bp  $\sim 17 \sim 2 \cdot 10^{10}$  bp
- Melting + annealing temperature  $\Rightarrow$  efficiency  
 $T_a \sim 55-60^\circ\text{C}$
- G/C content: why is 40-60% best?
- Avoid long runs of same/similar base  $\rightarrow$  mispriming
- Secondary structure considerations  $\rightarrow$  poor priming
- Binding considerations (energy; self, other)

GC clamp  
(3' end esp.)



# Mod 1 written assignments

- Lab report re: bacterial communities (15%)
  - Traditional format (intro, methods, etc.)
  - WAC training begins next time
  - Written in pairs *HW individually*
  - Can be revised for up to 1.33 letter grade higher
- Primer design summary (5%)
  - Short text and table summarizing design strategy
  - Short text and figure summarizing result
  - Written alone
  - Not subject to revision

# Mod 1 oral assignment

- Journal club (10%)
  - Purpose: summarize a recent research article
  - Sign up for Day 6 (Feb 28/Mar 1) or Day 8 (Mar 7/8)
  - Paper list available ?next week?
- Preparation
  - WAC training will be on Day 3 (Feb 14/15)
  - Will also practice discussing an article in-class on M1D3: start reading the paper this weekend
- Presentations will be videotaped, reviewed

# Participation and reflection in 20.109

- 1%: our perception of your engagement and contributions
- 2%: four reflections on your own learning
  - journal club self-assessment
  - module 1 report lessons learned
  - module 2 report lessons revisited
  - grab-bag: meeting with peers or instructors; discussion of outside research article
  - *extra-credit reflections*
  - *our hope: make learning gains more concrete*

# Today in Lab: M1D1

- Complete lab practical
- Explore existing diagnostic primers for  $\mu$ sporidia
- Design new primers
  - sensitivity *or* specificity challenge
  - sign up on M1D1 “Talk” page
- Notebooks start today!
  - primer table will be used in your M1 design summary
- For next time
  - keep exploring wiki... and add to it
  - start reading paper for M1D3 discussion