

DNA Engineering

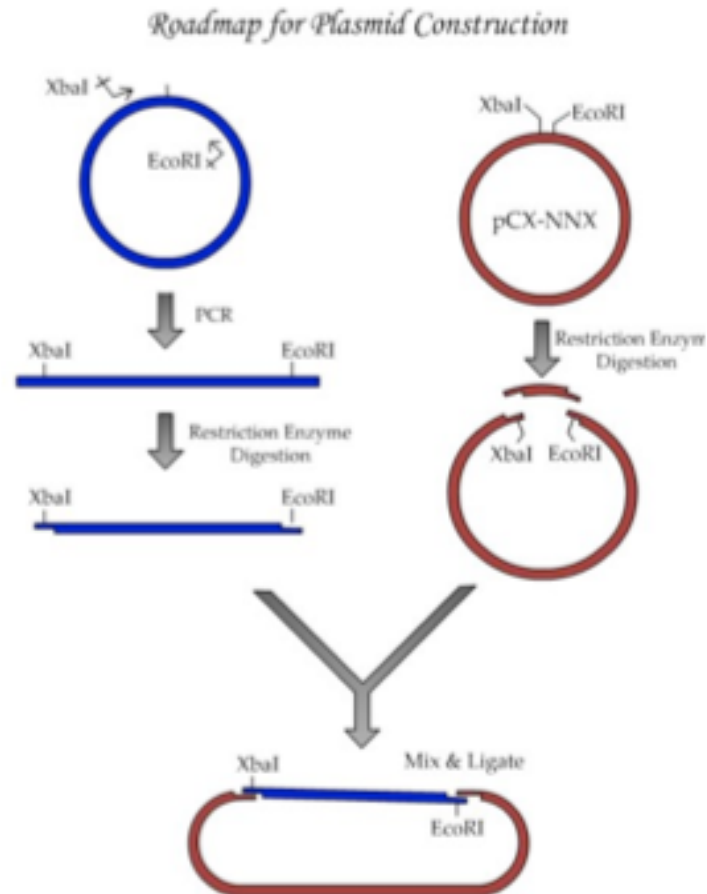
Mod 1 Day 5

Pre-lab Lecture

20.109 (F13)

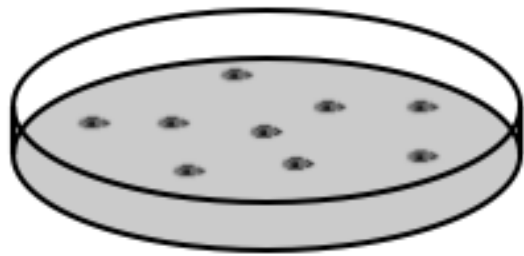
9/26/2013

M1D5

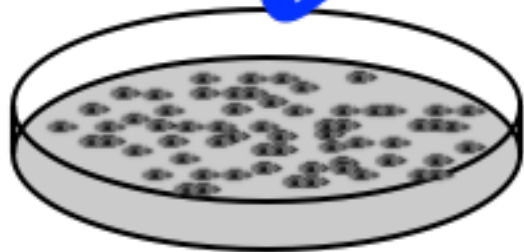


- Quiz
- Pre-lab discussion
- ½ class to TC
- Purify clones
- Set-up diagnostic digests
- Evaluate transformation

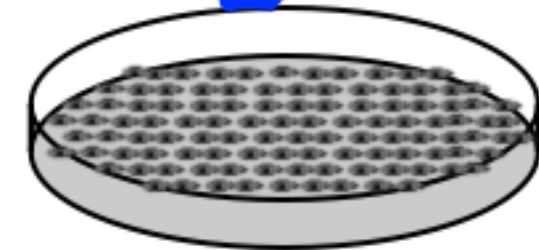
Tissue Culture!



Seeding \rightarrow Seeding Density

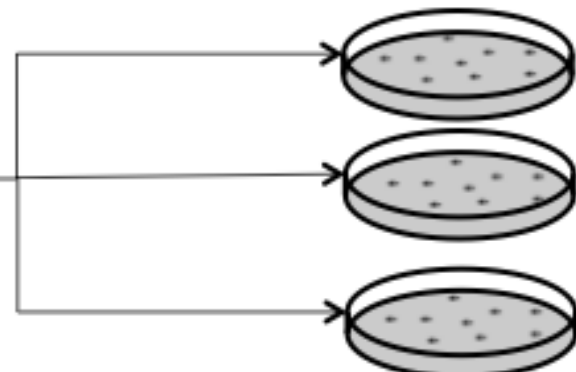


Proliferation Rate

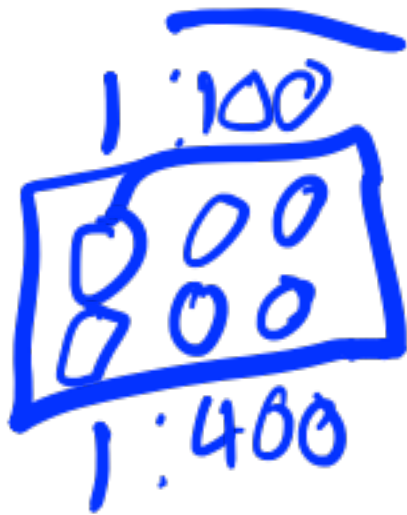


Confluency

Passage/
Split

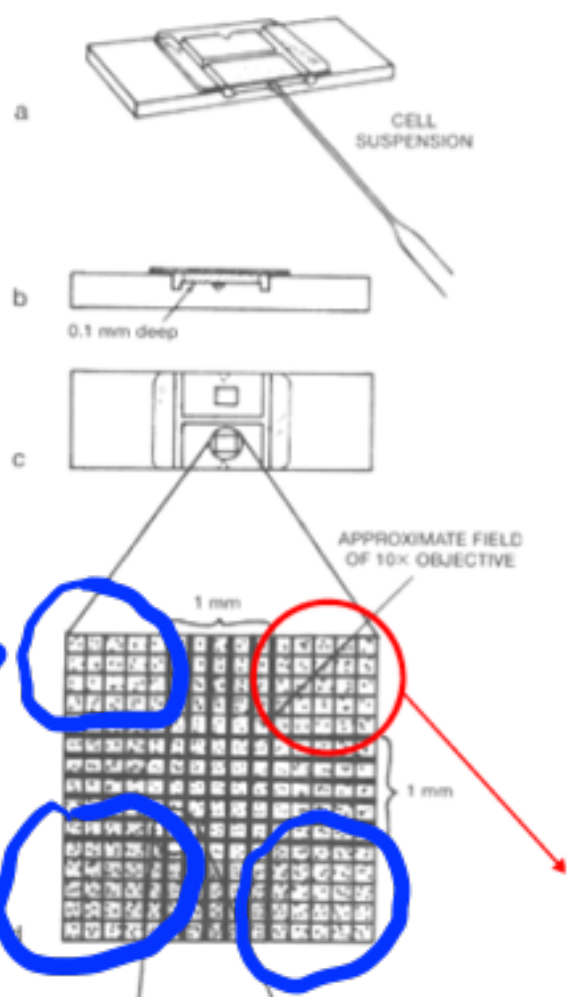


RBC's




1 square
= 0.1 μ L

Hemocytometer!



1. Count 4 corners
 2. Avg
 3. Calc Density (cells/mL)
- Avg $\times 10^4$



Review of Experiment + Controls

Experiment Condition	pCX-EGFP (5ng)	Backbone + Insert (no ligase)	Backbone + Ligase	Backbone + Insert + Ligase
Why?	Pos. Control Lots! ☺	Neg ☹	∅	EXP!
What if...	Few - Low den. - [AB] 	- Uncut bKb - nbn E. coli cont.	- Single digest	

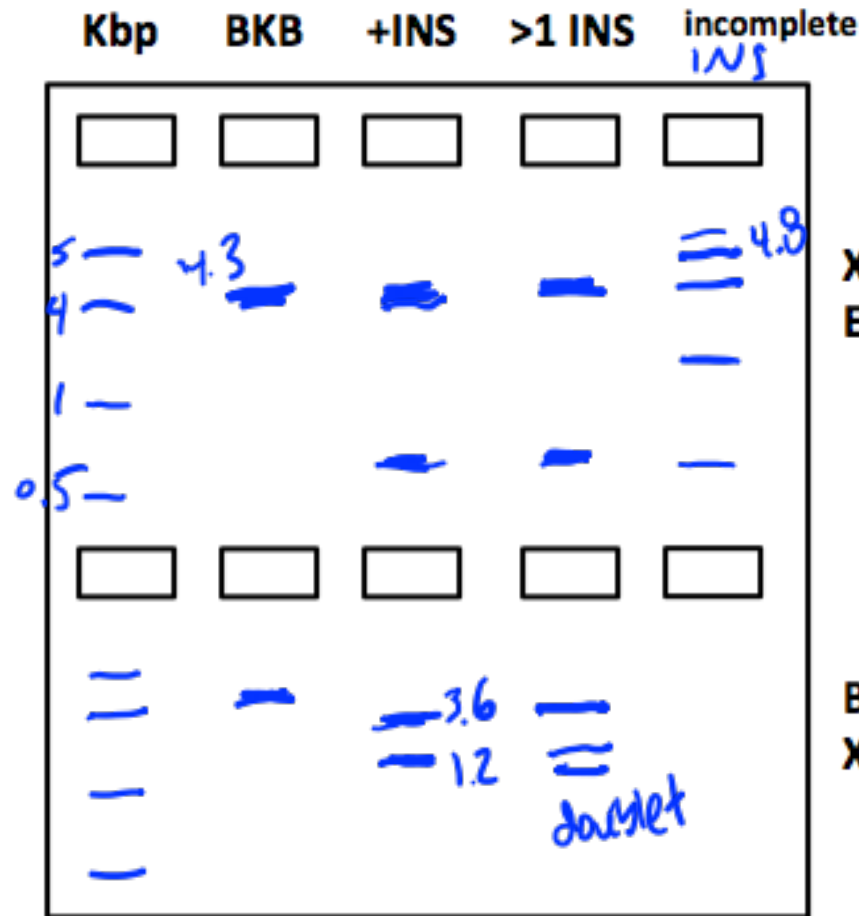
Variations in Experimental Samples

- Backbone-to-Insert ratio
- Incubation time
- Salt in ligation
- Unequal recovery

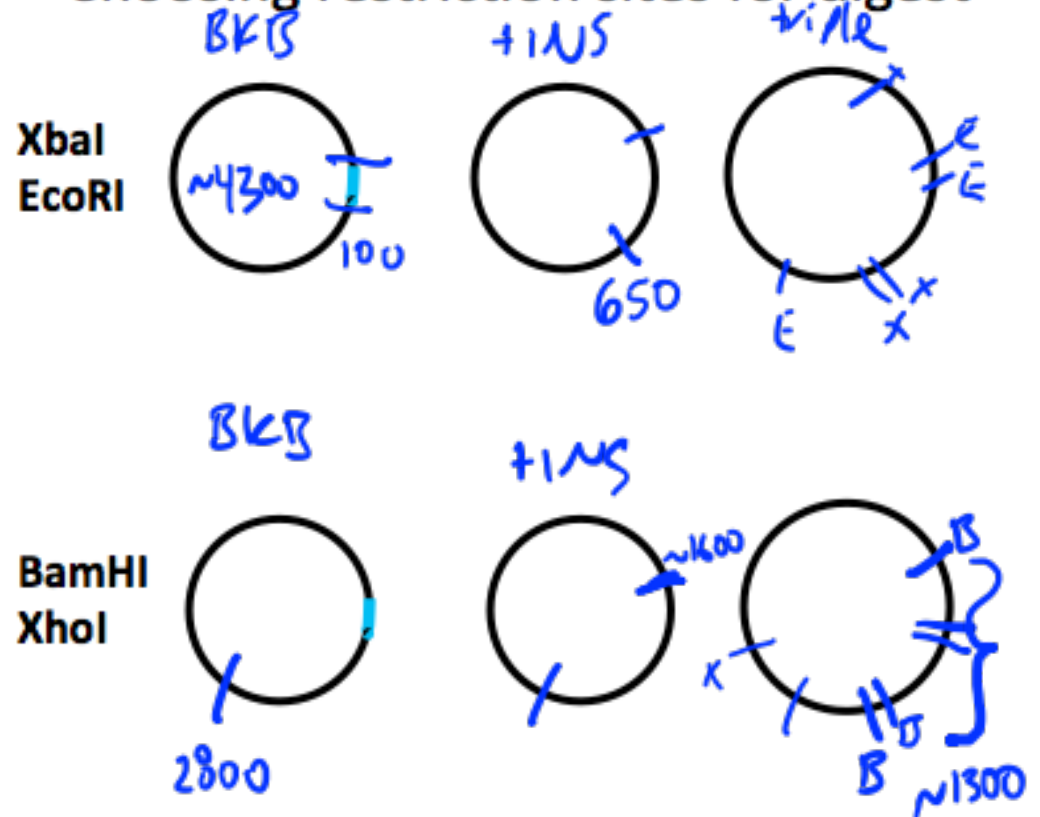
Extracting DNA from XL1-Blue

Step	Contains	Purpose
Prepare	EDTA Buffer, glucose	- weaken cell envelope - keep otherwise stable
Lyse	SDS  NaOH	- disrupt/solubilize lipid membranes, protein - denatures DS → SS DNA
Neutralize	Acetic acid/KAc	- neutralize pH, precipitate SDS 
Transfer	N/A	keep supernatant
Wash, collect	A) EtOH B) dry, water	- precipitate DNA - but EtOH interferes w/ digest

Diagnostic DNA gels



Choosing restriction sites for digest



Today in Lab (M1D5)

- Miniprep three $\Delta 5$ -EGFP candidates, and bacteria transformed with pCX-NNX
 - tip: orient tubes in centrifuge
 - pCX-NNX = control for *technique*
- Set up digests
 - tip: make reaction cocktail \rightarrow efficiency
 - add loading dye before leaving lab
- Count and post colony #s on M1D5 *Talk* page
 - we will discuss briefly before heading to TC
- TC practice session: don't need notebook, just a piece of scrap paper