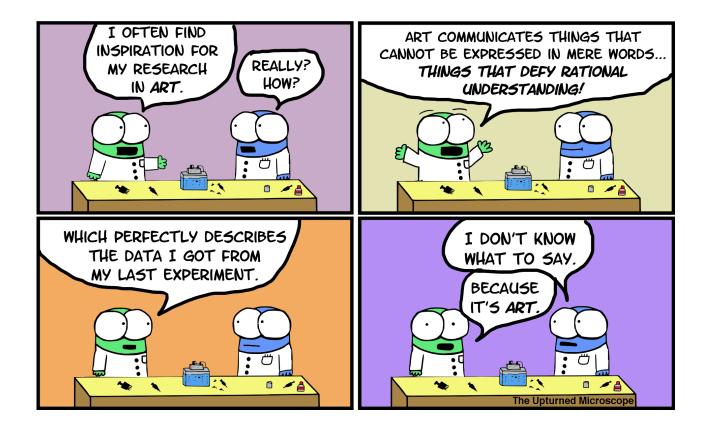
M1D4: Complete data analysis for γ H2AX experiment

- 1. Quiz
- 2. Prelab, part 1
- 3. Image analysis for γ H2AX assay
- 4. Paper discussion with Noreen
- 5. Prelab, part 2
- 6. Make a CometChip



Mod1 Overview

Last lab:

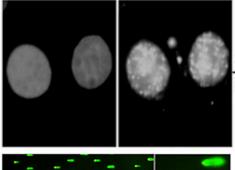
Completed yH2AX staining

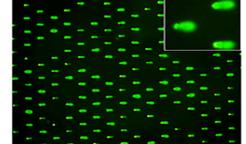
This lab:

yH2AX data analysis Paper Discussion Pouring Comet Chip

Next lab:

Completing Comet Chip





- **1**. Use repair foci experiment to measure DNA breaks
- Examine effect of H₂O₂ +/- As on double strand DNA breaks by measuring γH2AX foci formation

2. Use high-throughput genome damage assay to measure DNA damage

 Measure effects of H₂O₂ +/- As on DNA damage by measuring DNA migration in agarose matrix

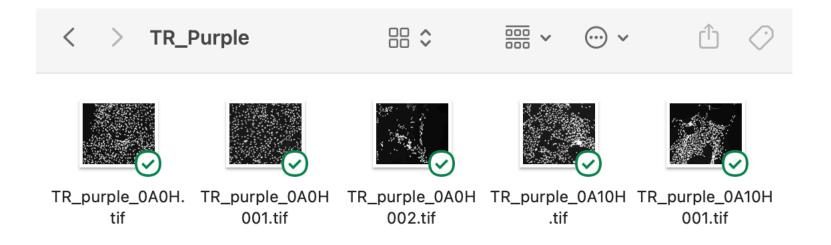
Image analysis has some potential pitfalls

- Data can be skewed dramatically by bias (conscious or unconscious)
 - Microscopy images are vulnerable to this because they are often used as representative of a much larger population
- How do we mitigate bias when taking and analyzing images?
 - Blind imaging or analysis
 - Set parameters ahead of time (i.e. select images randomly in the DAPI channel without looking at H2AX staining)
 - Try to create a field of view that encompasses multiple cells

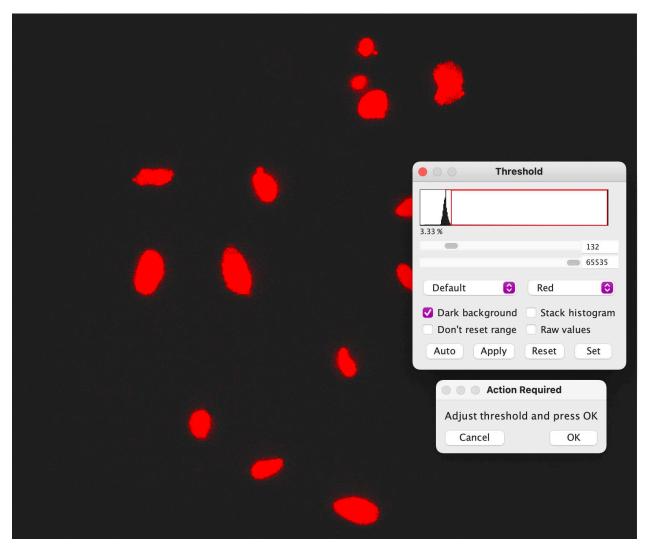
How will you analyze your images for the Data Summary?



- Use macro developed by Joshua Corrigan in Engelward lab
 - The DAPI channel used to create a "mask" of the nuclei
 - Gamma-H2AX foci are identified by pixel maxima readings in the FITC channel
 - You will be able to compare you "by eye" assessment of punctae to the count identified by the program
 - Average the number of foci per nuclei per image to get data point

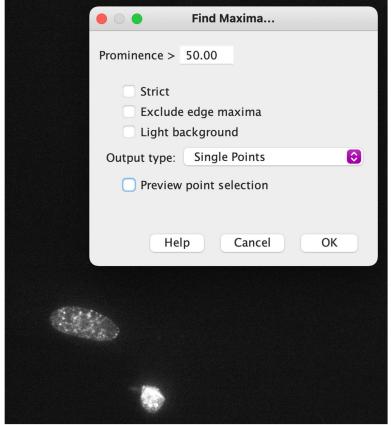


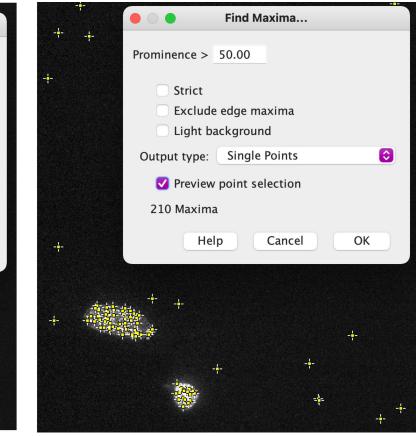
Set nuclei threshold to create region of interest to count foci



- Adjust threshold to capture discrete nuclei
 - May not be perfect!
- Program will also watershed the images to separate nuclei that are close together

Set prominence for the FITC/488 channel image





- Find a prominence setting that allows most visible foci to be counted in a condition while minimizing background counting
 - Select output of Single Points
 - Check Preview point selection

Compile results in Excel

| | | | Results | | | | | |
|----|--------------------------------------|-------|---------|-----|-----|-------|--------|--|
| | Label | Area | Mean | Min | Max | Circ. | IntDen | |
| 1 | 5H10As_40x_117-0002 Maxima:0004-0548 | 5972 | 0.000 | 0 | 0 | 0.267 | 0 | |
| 2 | 5H10As_40x_117-0002 Maxima:0005-0630 | 8132 | 0.000 | 0 | 0 | 0.287 | 0 | |
| 3 | 5H10As_40x_117-0002 Maxima:0007-0936 | 9354 | 0.000 | 0 | 0 | 0.359 | 0 | |
| 4 | 5H10As_40x_117-0002 Maxima:0009-1017 | 8844 | 0.000 | 0 | 0 | 0.321 | 0 | |
| 5 | 5H10As_40x_117-0002 Maxima:0013-1653 | 12860 | 0.000 | 0 | 0 | 0.412 | 0 | |
| 6 | 5H10As_40x_117-0002 Maxima:0014-1681 | 9359 | 0.000 | 0 | 0 | 0.264 | 0 | |
| 7 | 5H10As_40x_117-0002 Maxima:0017-2047 | 10956 | 0.000 | 0 | 0 | 0.423 | 0 | |
| 8 | 5H10As_40x_117-0002 Maxima:0002-0252 | 8709 | 0.029 | 0 | 255 | 0.326 | 255 | |
| 9 | 5H10As_40x_117-0002 Maxima:0008-1004 | 21650 | 0.012 | 0 | 255 | 0.371 | 255 | |
| 10 | 5H10As_40x_117-0002 Maxima:0015-1952 | 8416 | 0.030 | 0 | 255 | 0.301 | 255 | |
| 11 | 5H10As_40x_117-0002 Maxima:0001-0230 | 9846 | 0.052 | 0 | 255 | 0.495 | 510 | |
| 12 | 5H10As_40x_117-0002 Maxima:0003-0307 | 10179 | 0.050 | 0 | 255 | 0.295 | 510 | |
| 13 | 5H10As_40x_117-0002 Maxima:0006-0938 | 13402 | 0.038 | 0 | 255 | 0.233 | 510 | |
| 14 | 5H10As_40x_117-0002 Maxima:0011-1481 | 13157 | 0.058 | 0 | 255 | 0.260 | 765 | |
| 15 | 5H10As_40x_117-0002 Maxima:0010-1038 | 14512 | 0.176 | 0 | 255 | 0.229 | 2550 | |
| 16 | 5H10As_40x_117-0002 Maxima:0016-1983 | 15859 | 0.338 | 0 | 255 | 0.325 | 5355 | |
| 17 | 5H10As_40x_117-0002 Maxima:0012-1541 | 24834 | 0.226 | 0 | 255 | 0.354 | 5610 | |

• Results should have a Max of 0 or 255

 Integrated Density should be in multiples of 255

Data analysis required for Data Summary

- Complete the analysis of images in all conditions for your group (3 replicates per condition)
- Then complete the image analysis for a biological replicate of **pilot data** from instructors
 - Divide the work amongst your lab team!
- Once the numbers are recorded for each experiment, take the average number of foci for <u>each</u> image (i.e. treat each image as n=1)
 - This is a special circumstance for this class!
 - Statistics are another lab session
- The average number of foci in each treatment condition will become a figure in the Data Summary

Homework

Data figure

Data figure example

- Image **should not** be the entire page
 - Only needs to be large enough to be clear / visible
 - 1/3 1/2 of a page in portrait orientation
- Title **should** be conclusive
 - Don't include what you did, rather state what you found (take home message)
- Caption should not detail the methods or interpret the data
 - Define abbreviations, symbols, etc.
 - Info needed to "read" figure
 - Figure captions with multiple panels need to start with a topic sentence

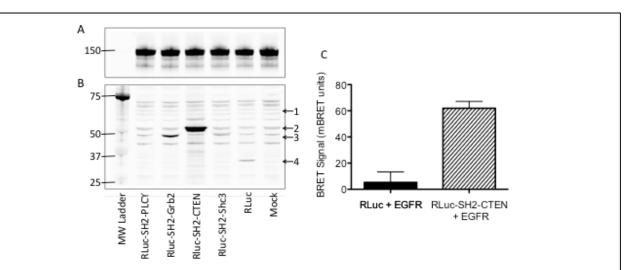


Figure 1: Development of BRET assay to monitor EGFR and SH2 domain interactions. CHO-K1 cells were transfected with Citrine-EGFR (A) and renilla luciferase (RLuc)-tagged SH2 domains from PLCg, Grb2, CTEN, and Shc3 (B). Western blots of CHO-K1 lysates were probed with anti-EGFR (A) or anti-RLuc (B) antibodies. Arrowheads indicate the expected molecular weight of the RLuc-tagged proteins; (1) RLuc-SH2-PLCg, (2) RLuc-SH2-CTEN, (3) RLuc-SH2-Grb2 and RLuc-SH2-Shc3, and (4) RLuc alone. Mock indicates no cDNA was utilized during transfection. (C) For CTEN only, BRET signal was quantified using a luminometer after stimulation of CHO-K1 with 100 ng/mL EGF for 15 min.

Data Summary = pptx file with slides set at 8.5 x 11" portrait

In lab today:

- 1. Work on image analysis until 2:45pm
- 2. Paper discussion from 2:45-3:30ish
- 3. Prelab part 2: making a CometChip gel
- 4. Work in teams to pour CometChip gels

HW due M1D5

- 1. Create a data figure of H2AX results with title and caption
- 2. Write up a short summary of your Comm Lab visit.