

# 20.109 Spring 2015 Module 2 – Lecture 6

## System Engineering and Protein Foundations



Shannon Hughes

Noreen Lyell

Leslie McLain

Nova Pishesha (TA)

Leona Samson (Lectures)

Zachary Nagel (help with development) Alex Chaim



## What experimental question will you ask in Module 2?

How efficiently does DNA repair by the Non Homologous End Joining (NHEJ) pathway act on DNA damage with different topologies?



## This raises the following questions

- How does DNA get damaged?
- What is DNA repair?
- Why does DNA repair exist?
- Why do we care about how efficient DNA repair is?
- How does one actually measure DNA repair efficiency?

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A · W · A · R · D

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INTERVIEW

June 16<sup>th</sup> 2009,  
8am!

# Developing Novel Methods to Measure DNA Repair Capacity in Human Populations

*Leona D. Samson*

MIT

Biological Engineering Department

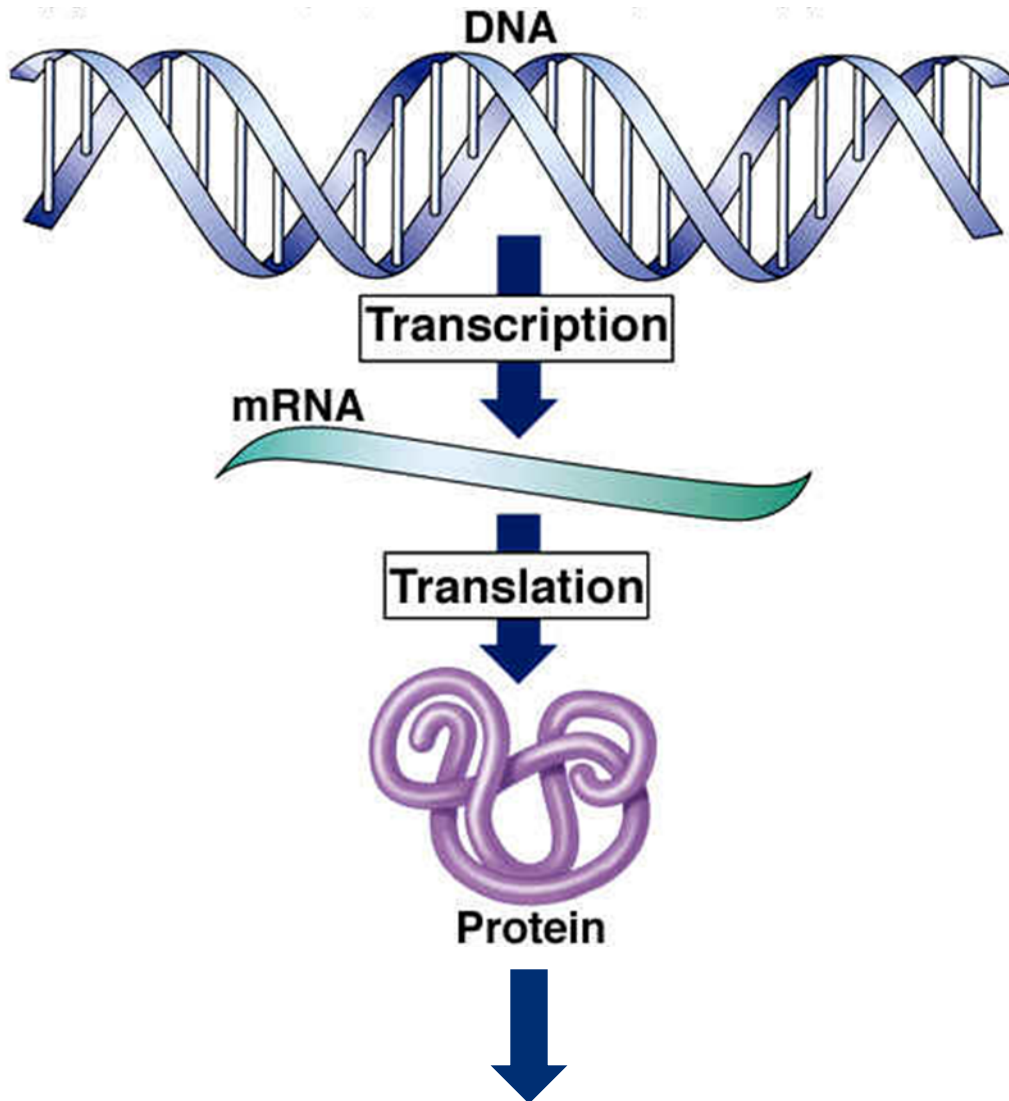
Biology Department

Center for Environmental Health Sciences

Koch Institute for Integrative Cancer Research

Computational and Systems Biology Initiative

Broad Institute (Harvard and MIT)



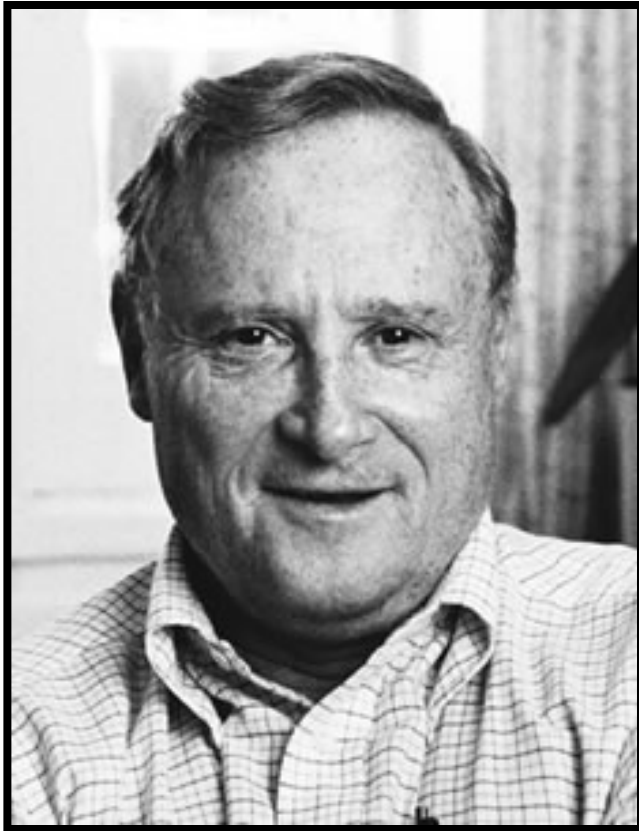
SNPs – GWAS  
Genome sequencing

mRNA (miRNA, lncRNA)  
Profiling  
Exome Sequencing

Proteomic Analyses

*In vitro / In vivo*  
Functional Assays

# The Proposal was based on the Pioneering work of:



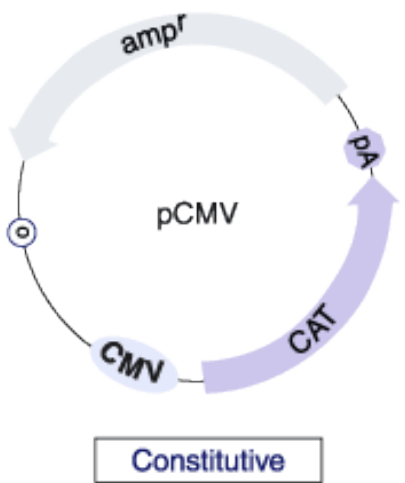
Dr. Lawrence Grossman  
(1924–2006)



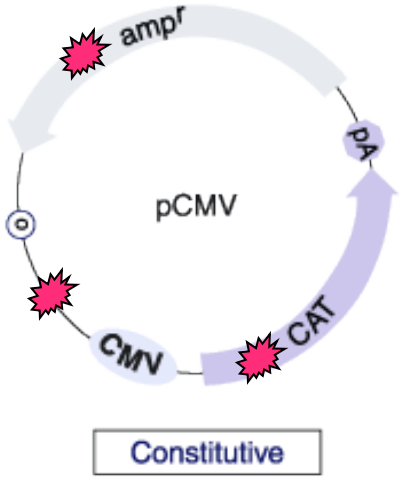
Dr. Qingyi Wei

# Reactivation of UV damaged DNA by Host cell Reactivation (HCR)

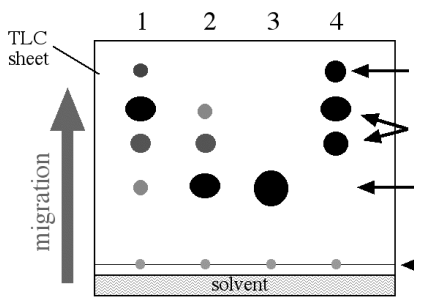
Athas & GROSSMAN  
Cancer Res. 1991



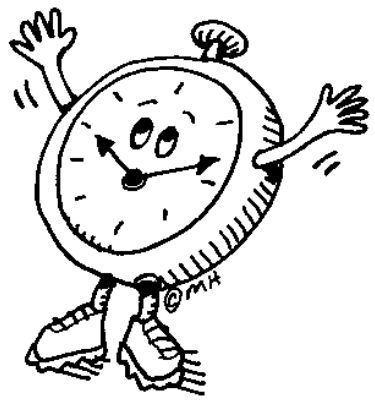
+ UV  
light



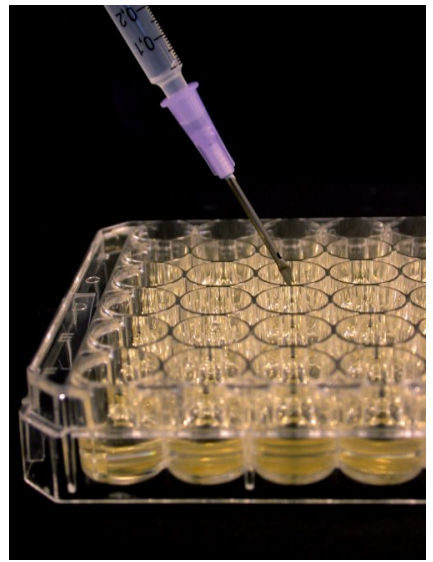
Transient  
transfection  
peripheral  
blood  
lymphocytes



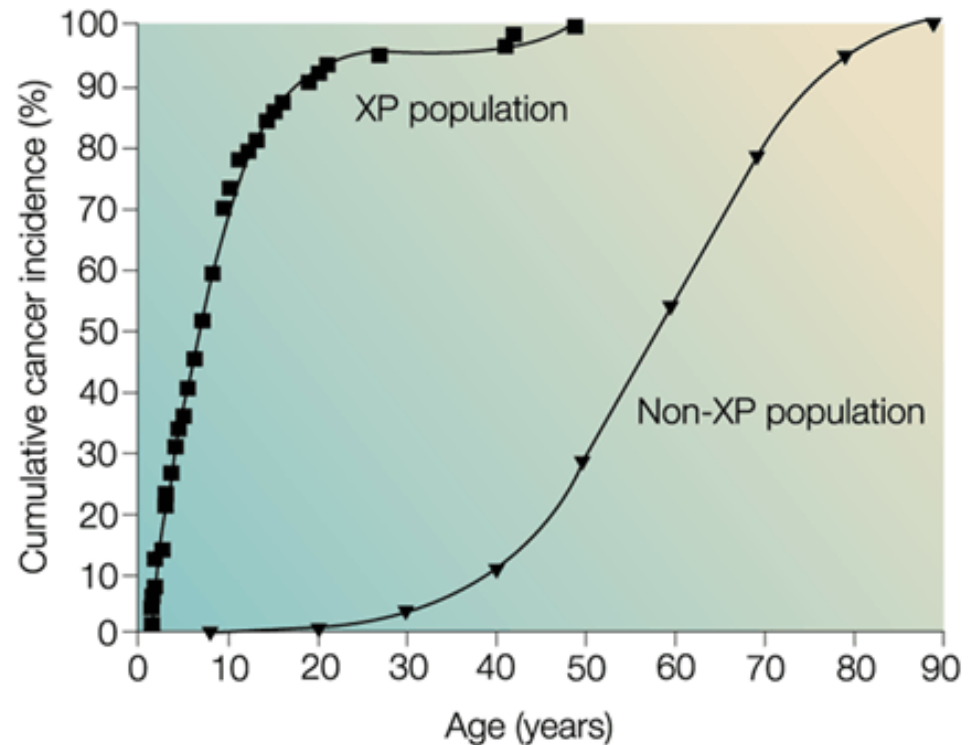
CAT Assay



Time to repair

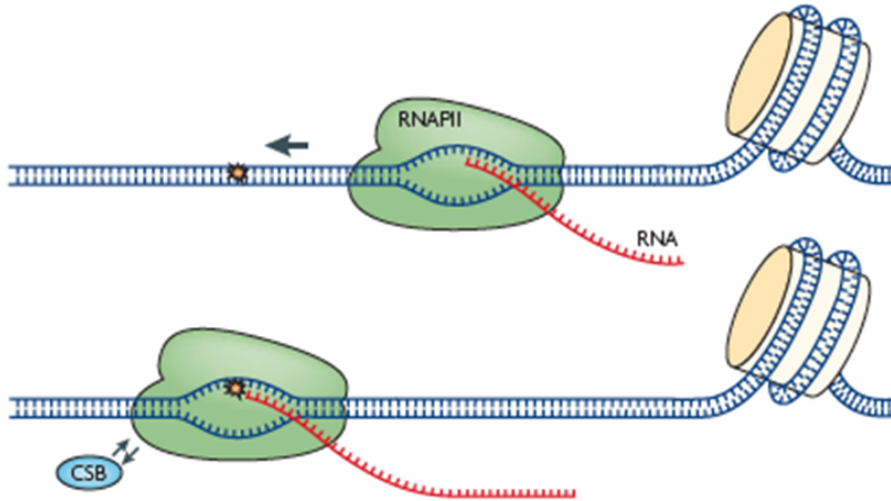


# Lack of NER accelerates the onset of skin cancer in Xeroderma Pigmentosum patients



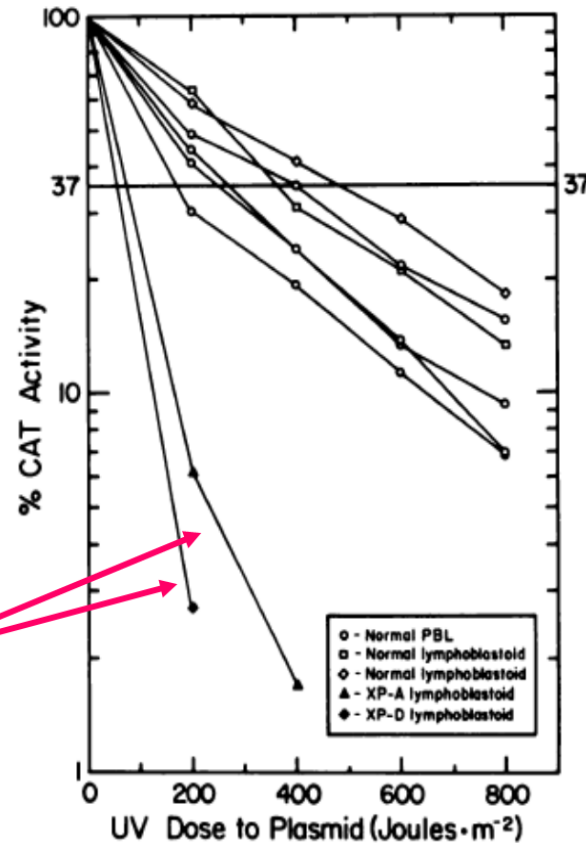
1, 22-33 (October 2001)

# RNA Polymerase II is exquisitely sensitive to DNA lesions



XP cells  
NER  
deficient

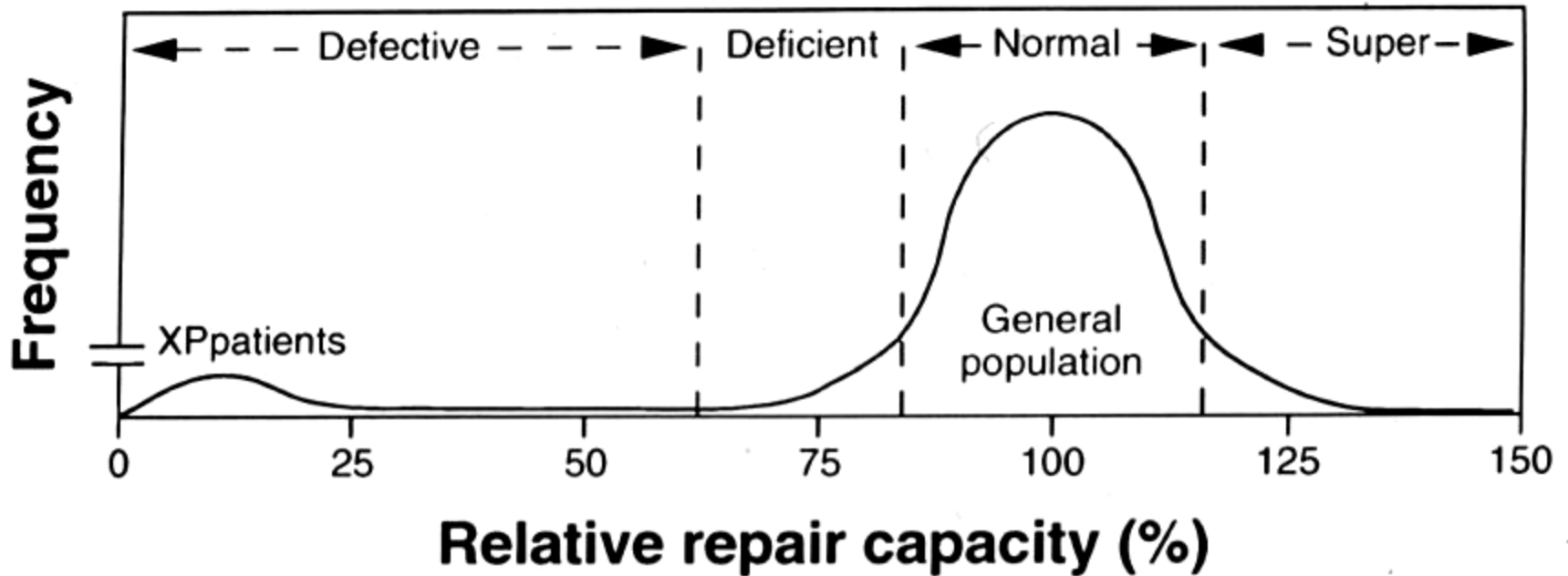
Fresh Circulating Lymphocyte  
Plasmid HCR in XP and Normal PBL



'WT' for  
NER



# Interindividual Variation in DNA Repair Capacity

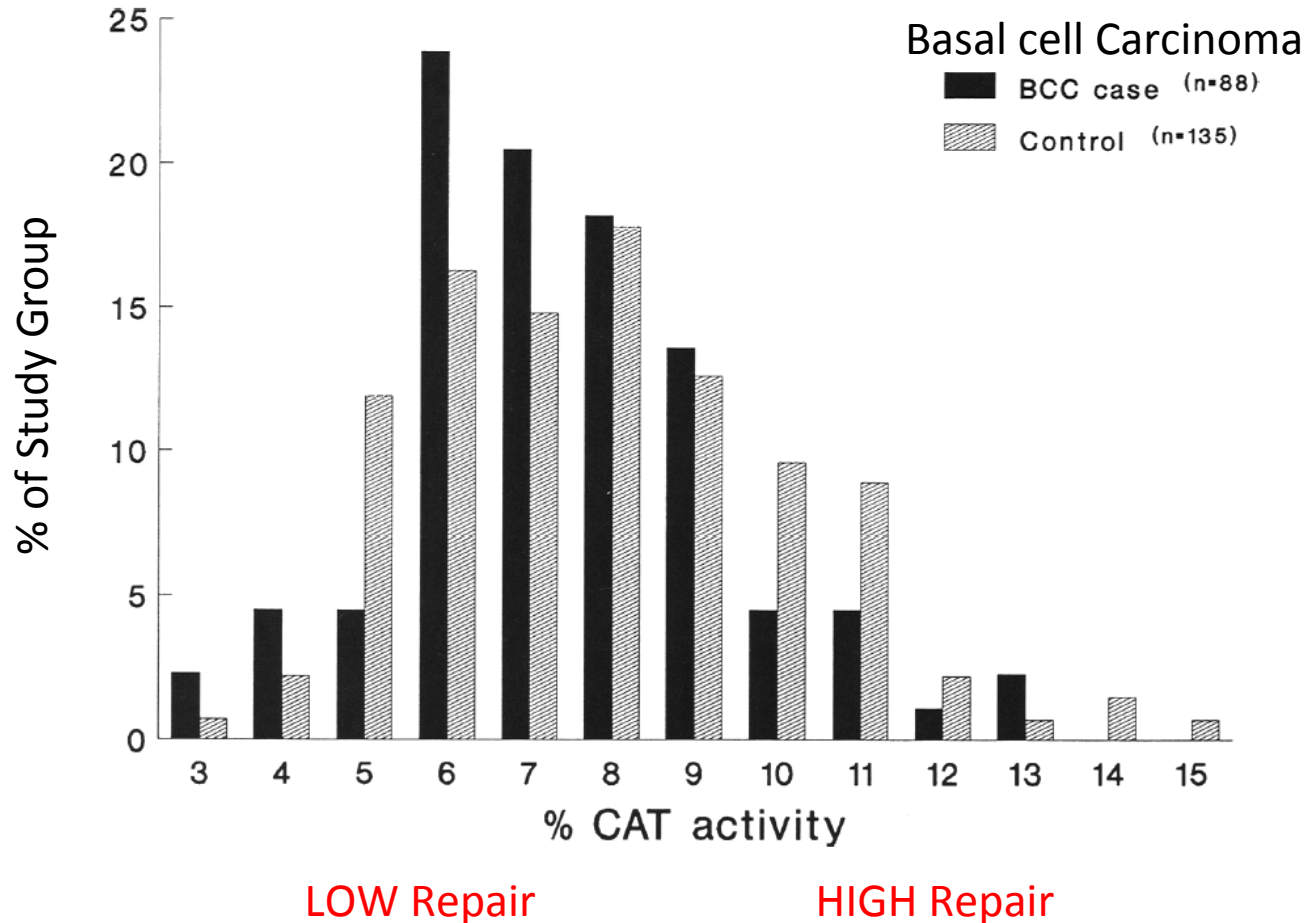


Adapted from **GROSSMAN and Wei (1995)** Clinical Chem 41: 1854-1863

XP frequency =  $\sim 1:250,000$  giving theoretically  $\sim 28,000$  cases worldwide with **2,000-fold increased skin cancer risk**

Even if just 1% of the population is relatively repair deficient, could have tens of millions with several-fold increased risk

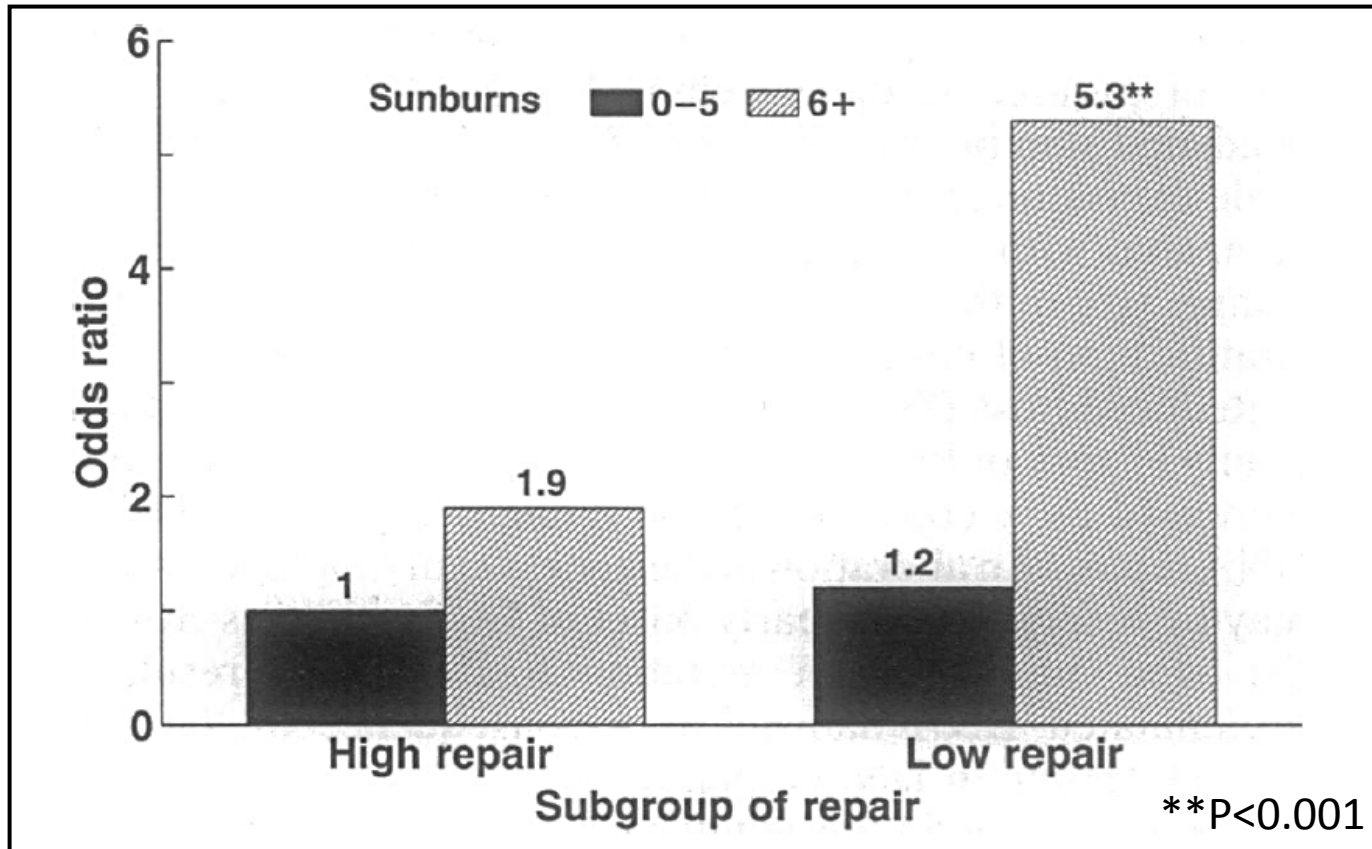
# Case-Control Study monitoring DNA Repair Capacity (DRC) by Host Cell Reactivation (HCR) of plasmids containing DNA damage



[CANCER RESEARCH 54, 437-44(i), January 15, 1994]

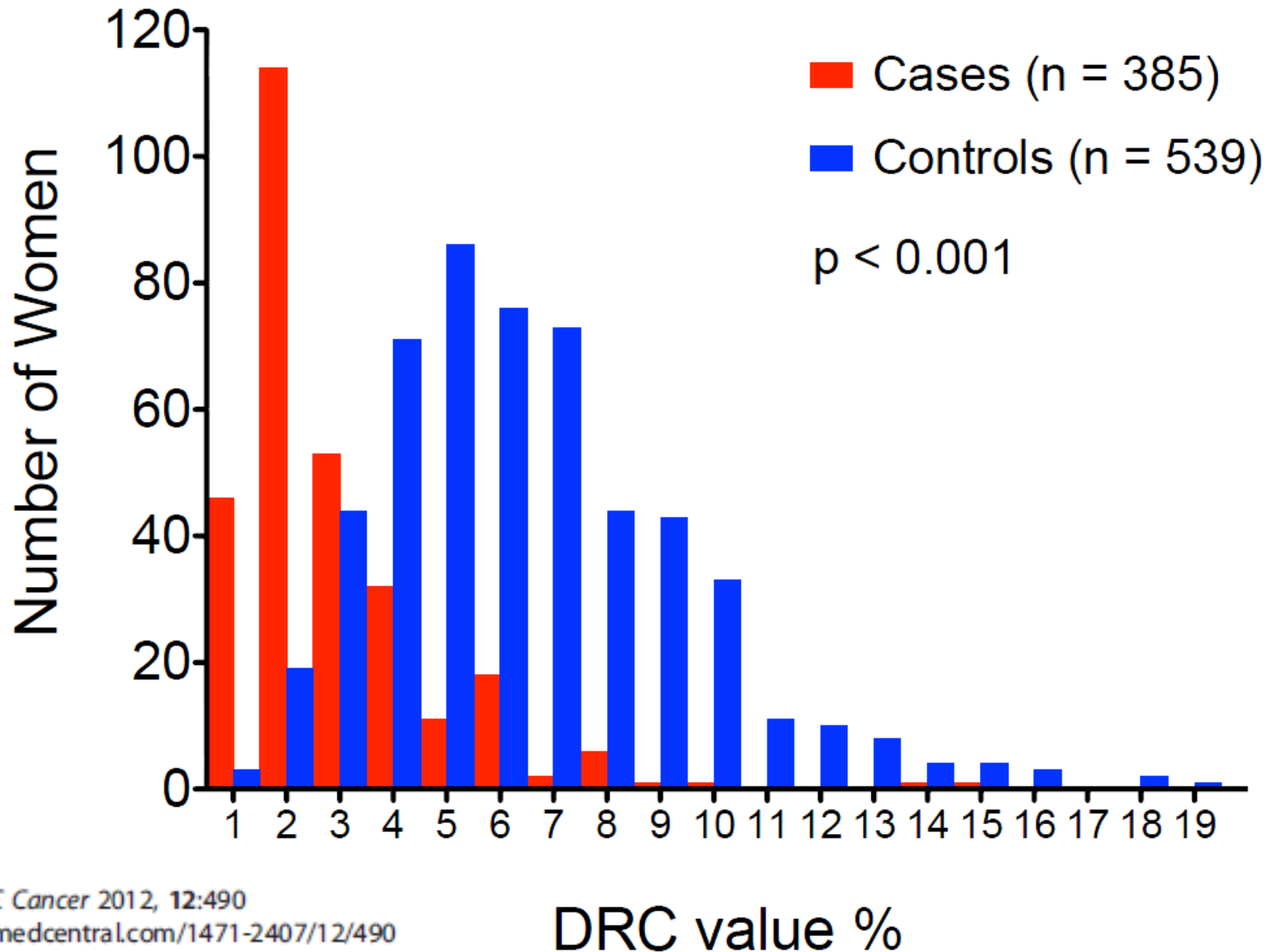
**Qingyi Wei**, Genevieve M. Matanoski, Evan R. Farmer, Mohammad A. Hedayati, and **Lawrence GROSSMAN**

# Low NER Repair status combined with excessive sun exposure is very dangerous

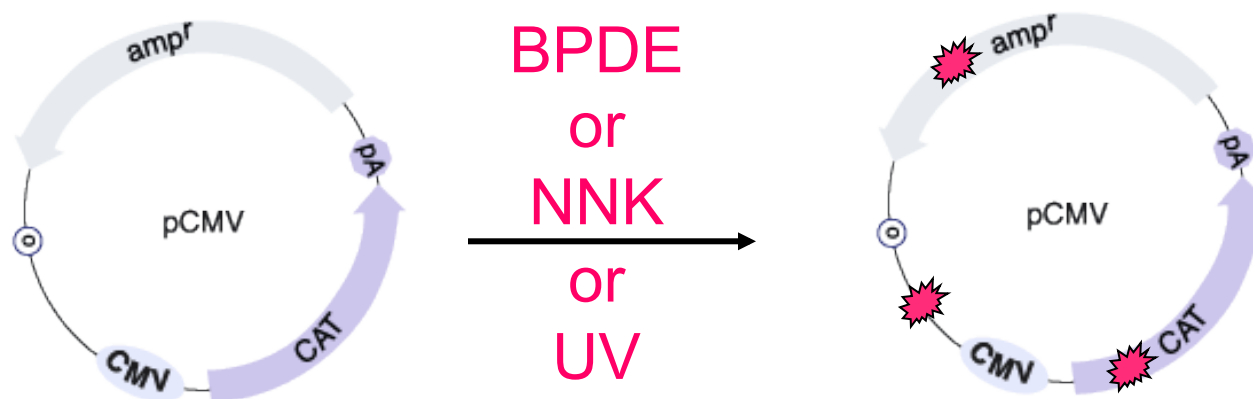


**Wei Q**, Matanoski GM, Farmer ER, Hedayati MA, **GROSSMAN L**. Proc Natl Acad Sci U S A. 1993 90:1614-8.

# DNA Repair Capacity (DRC) in **Breast Cancer** versus **Control** Patients

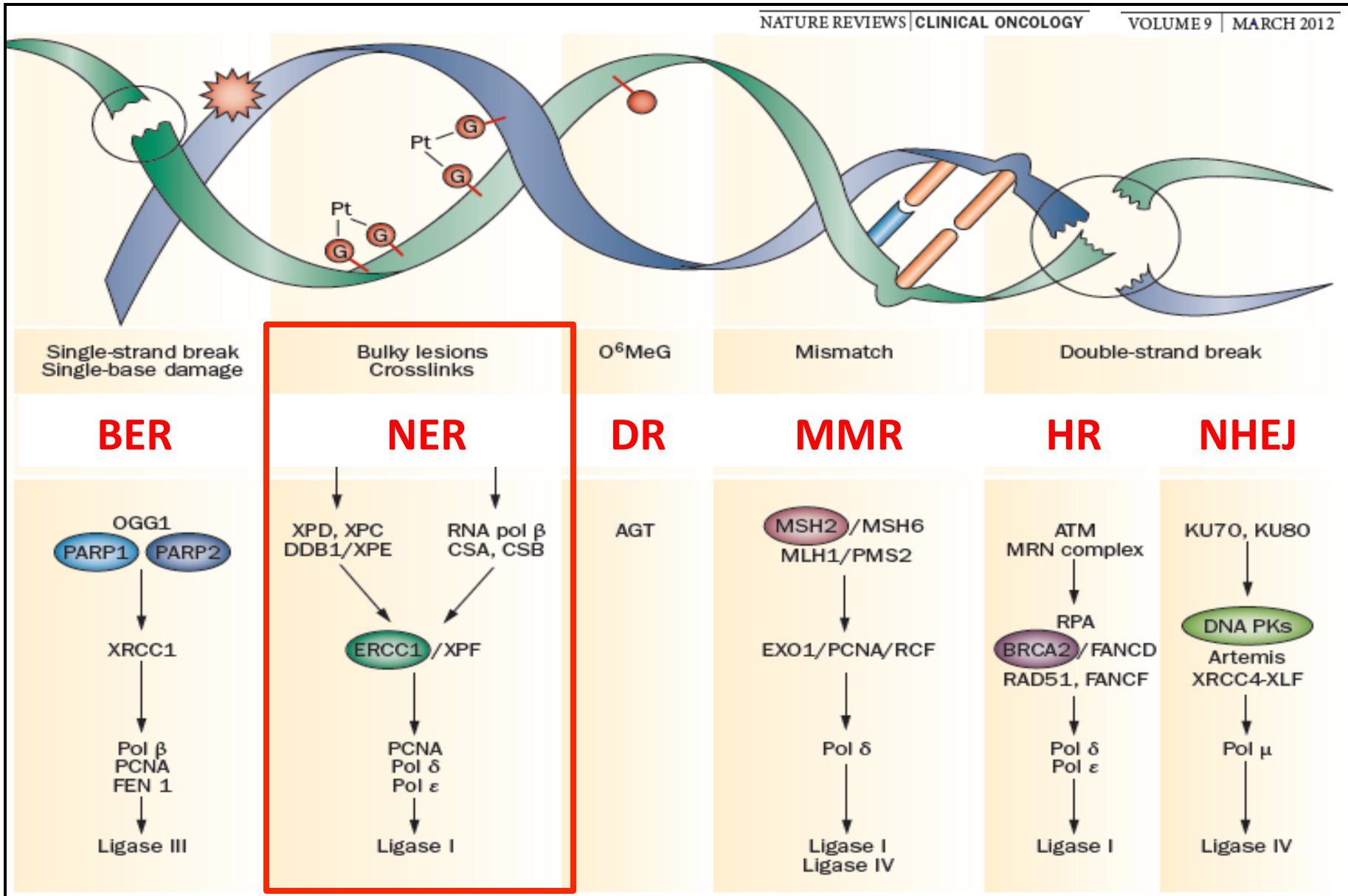


# Many cancer case/control studies have monitored Nucleotide Excision Repair (NER)



Damage	Cancer	Increased Risk Low v High Repair	Source
BPDE	Lung	5.7	Wei, 1996
"	Head & Neck	2.2	Cheng, 2005
"	Breast	3.4	Shi, 2004
NNK	Lung	3.2	Wang, 2007
UV	Skin	2.0	Wei, 2002
"	Melanoma	2.0	Wei, 2002
"	Breast	>10	Matta, 2012

# Six Major DNA Repair Pathways



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INTERVIEW

June 16<sup>th</sup> 2009,  
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# Developing Novel Methods to Measure DNA Repair Capacity in Human Populations

*Leona D. Samson*

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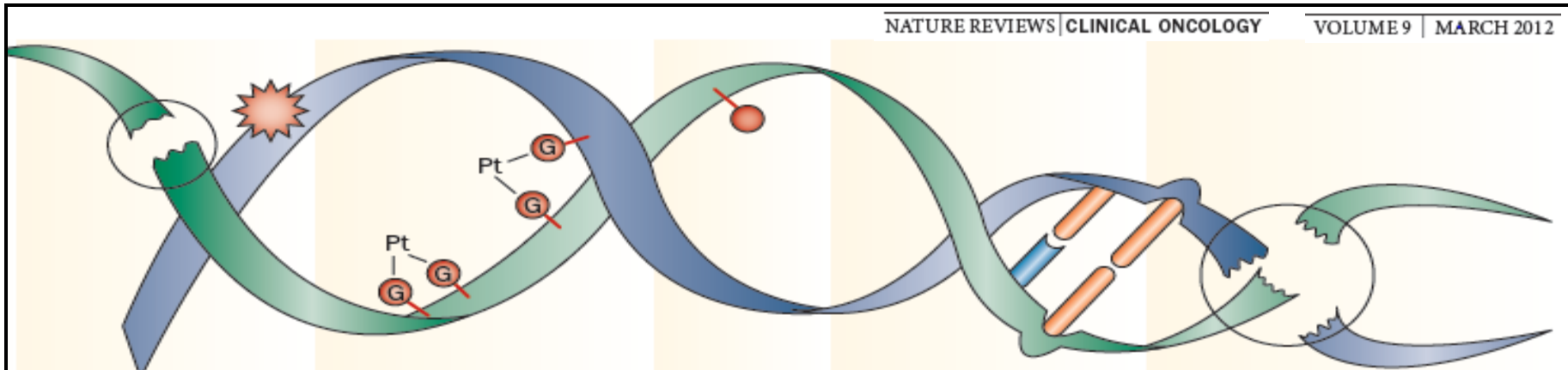
Koch Institute for Integrative Cancer Research

Computational and Systems Biology Initiative

Broad Institute (Harvard and MIT)

# Six Major DNA Repair Pathways

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Single-strand break  
Single-base damage

Bulky lesions  
Crosslinks

O<sup>6</sup>MeG

Mismatch

Double-strand break

**BER**

**NER**

**DR**

**MMR**

**HR**

**NHEJ**

OGG1  
PARP1 PARP2

XRCC1

Pol β  
PCNA  
FEN 1

Ligase III

XPD, XPC  
DDB1/XPE

RNA pol β  
CSA, CSB

ERCC1/XPF

PCNA  
Pol δ  
Pol ε

Ligase I

AGT

MSH2/MSH6  
MLH1/PMS2

EXO1/PCNA/RCF

Pol δ

Ligase I  
Ligase IV

ATM  
MRN complex

RPA

BRCA2/FANCD  
RAD51, FANCF

Pol δ  
Pol ε

Ligase I

KU70, KU80

DNA PKs

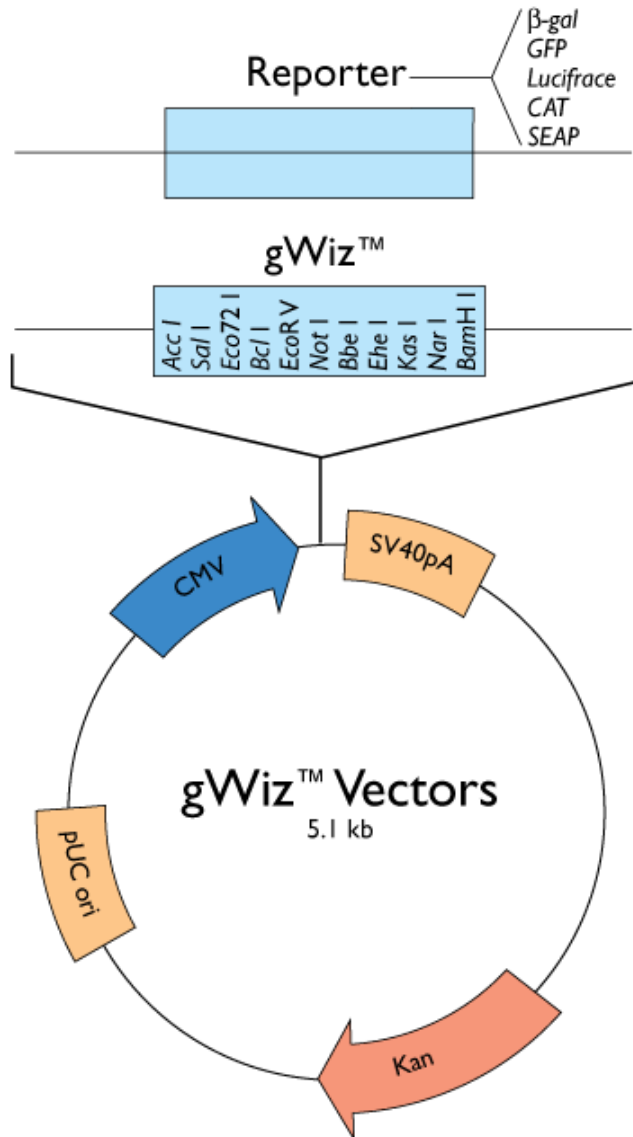
Artemis  
XRCC4-XLF

Pol μ

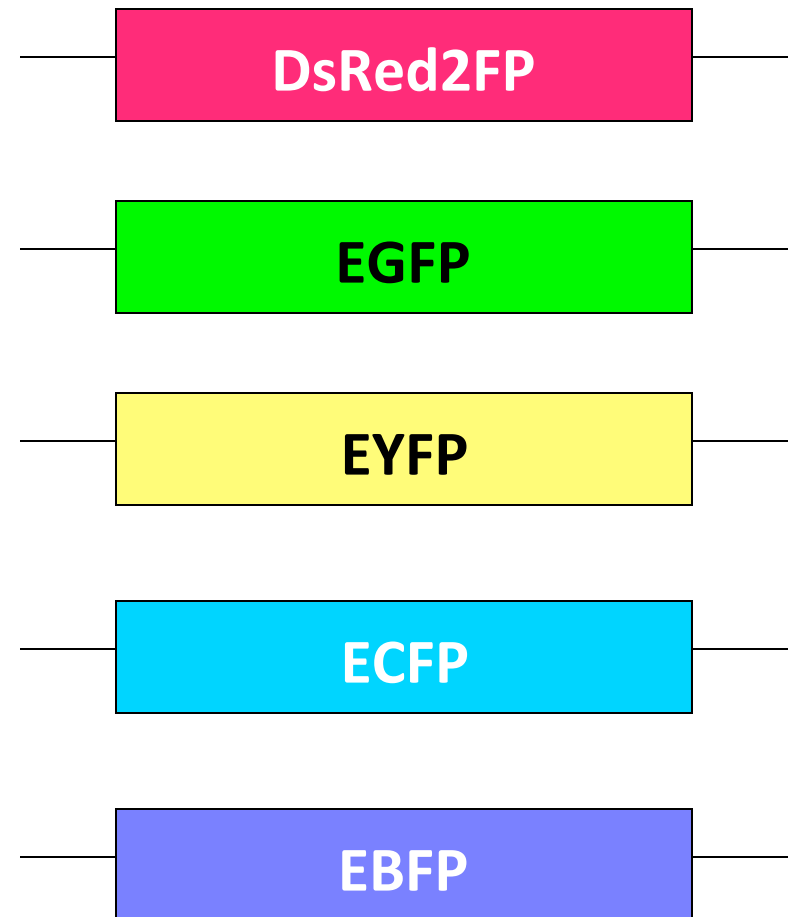
Ligase IV



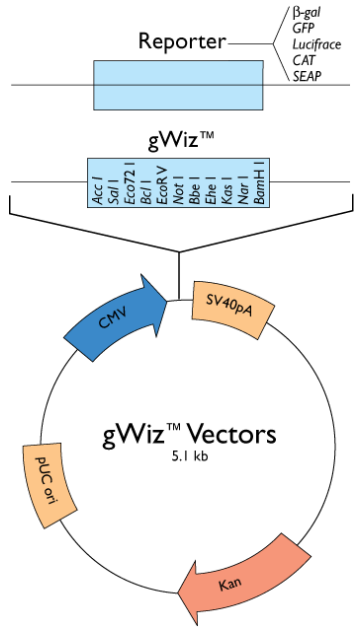
# Reactivation of damaged DNA - multiplexed



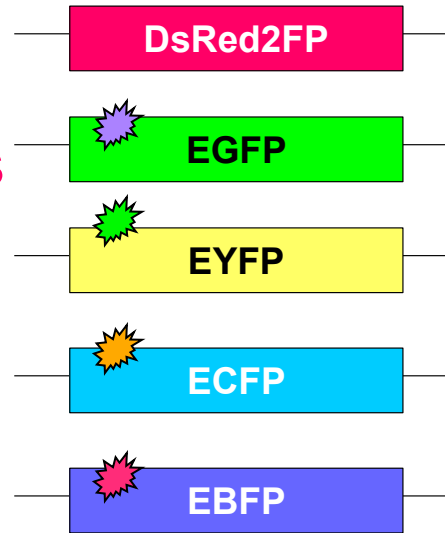
Each Fluorescent Protein gene will harbor a different type of DNA damage



# Reactivation of damaged DNA - multiplexed

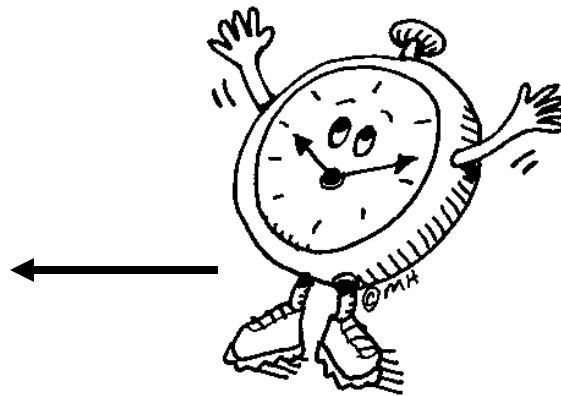
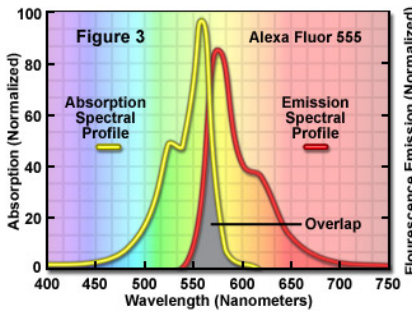


+ different  
DNA lesions

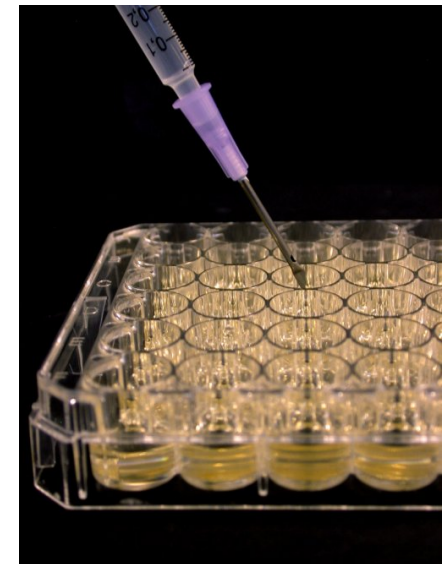


Transient  
transfection  
of mixture

Fluorophore Absorption and Emission Profiles

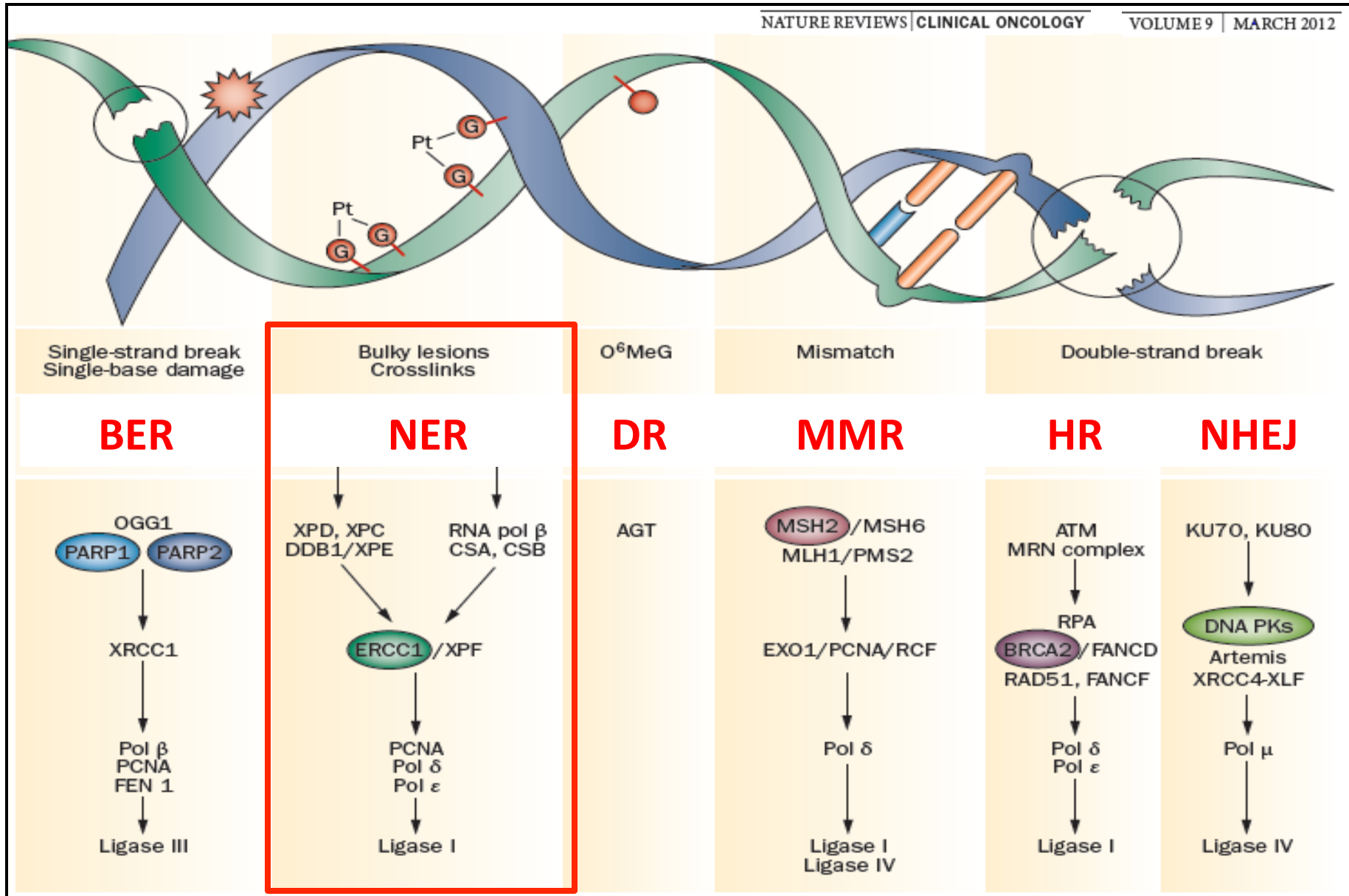


Time to repair

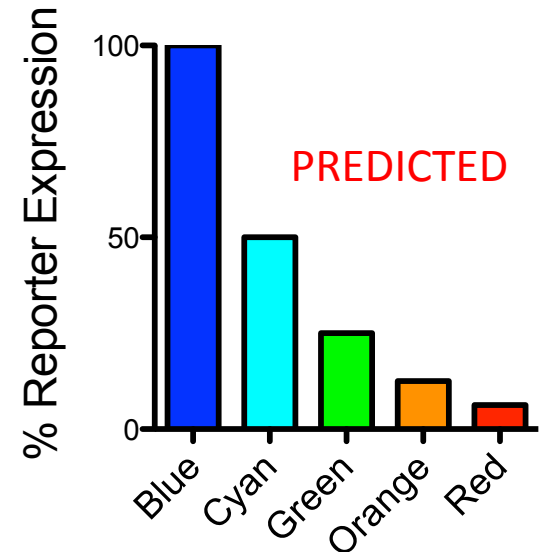
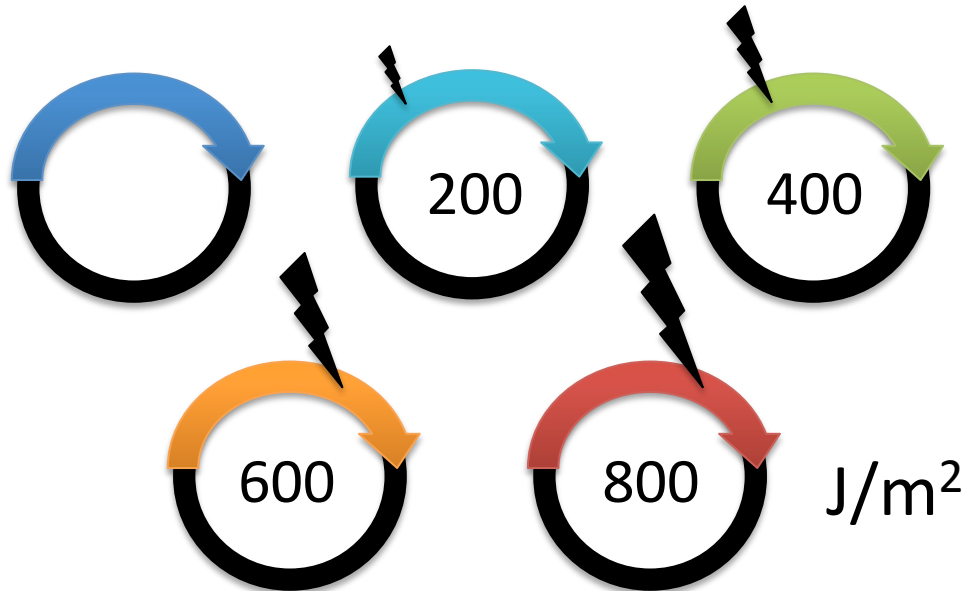
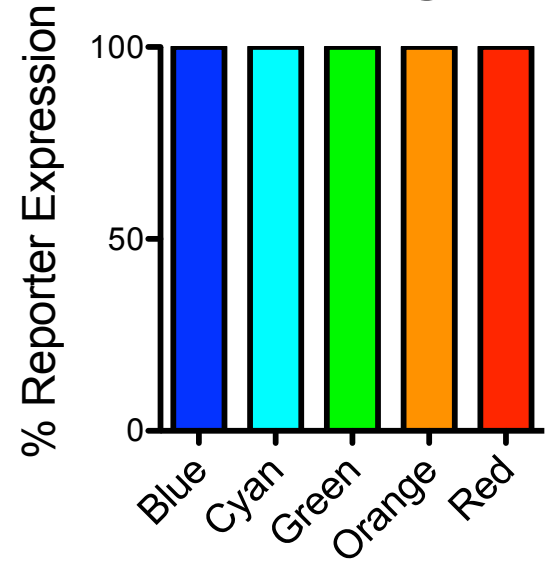
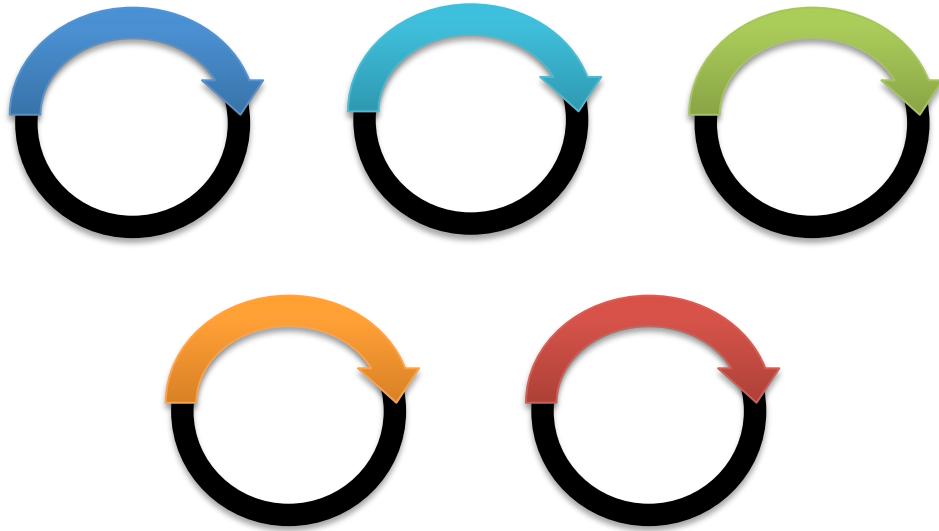


Fluorescence  
quantitation

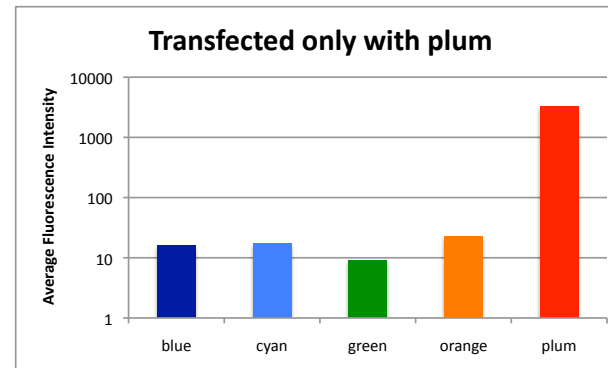
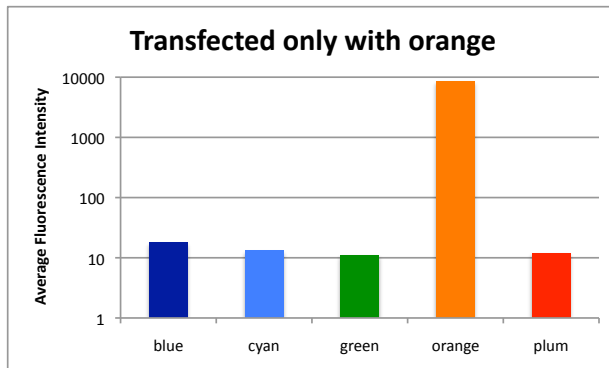
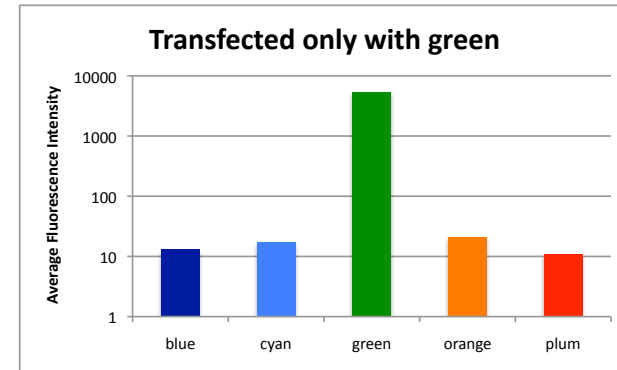
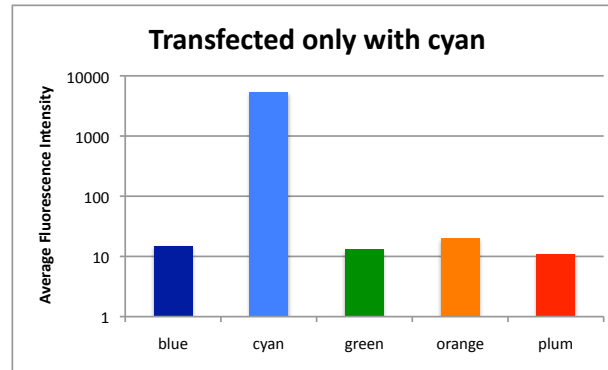
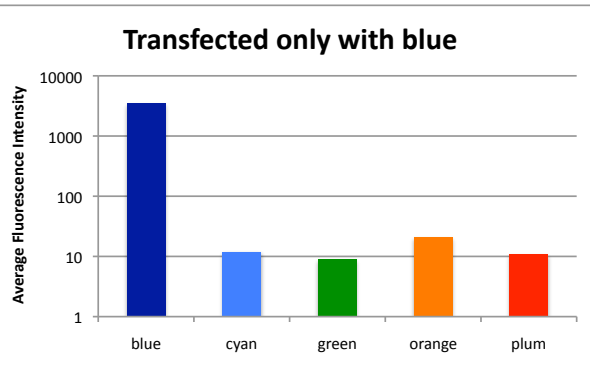
# Six Major DNA Repair Pathways



Before trying different damages - tried different doses of the same damage (UV)

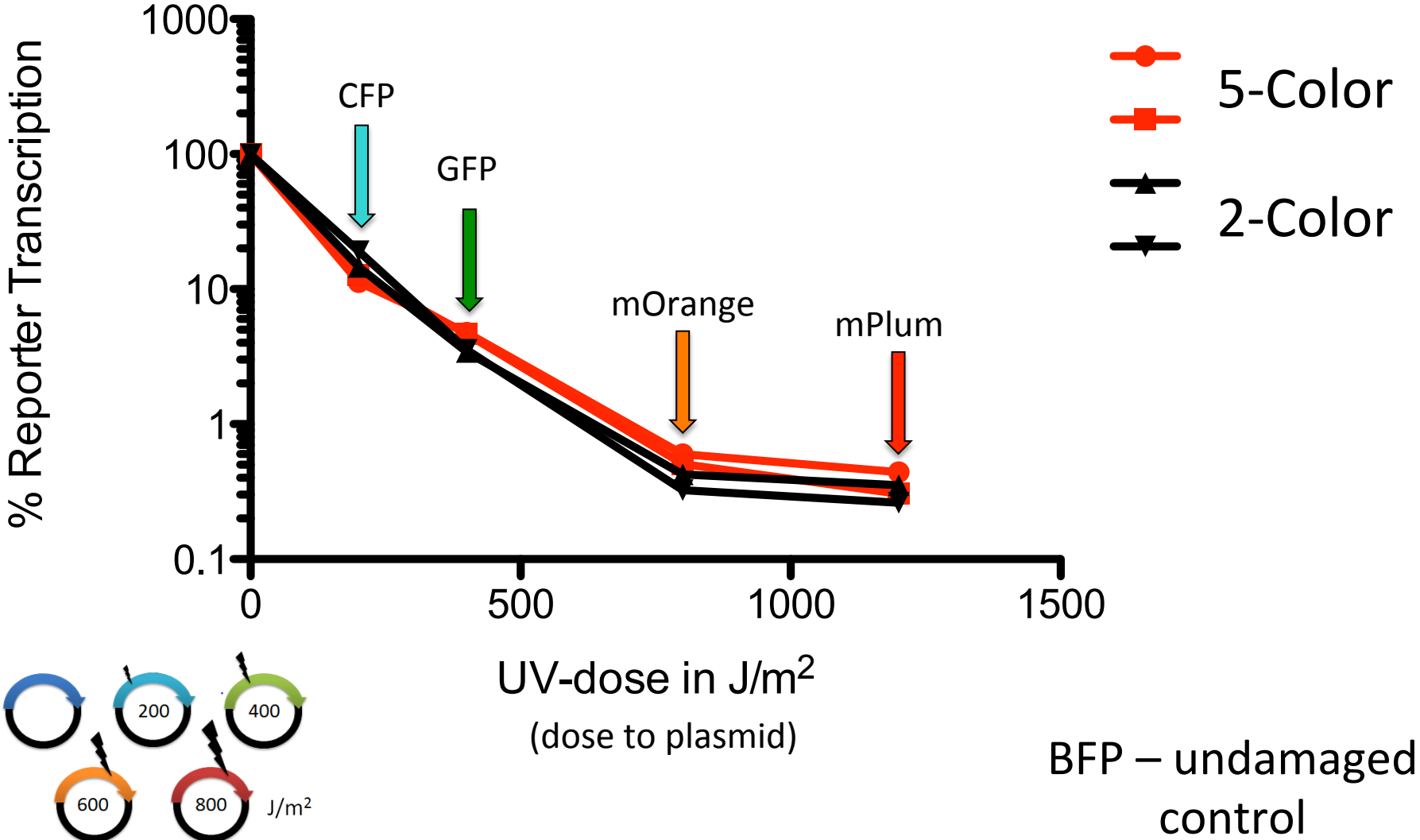


# Sanity Check: Is it even feasible detect 5-colors independently?:

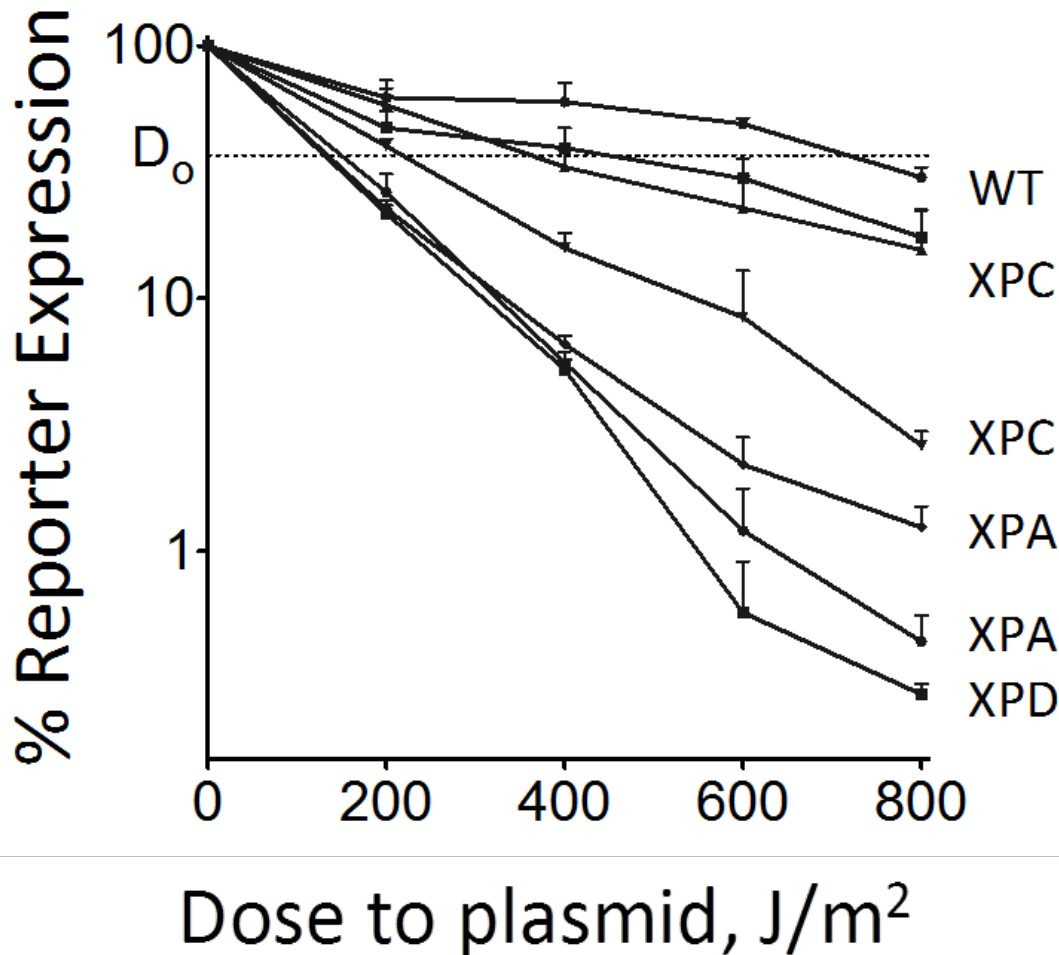


# 2-color versus 5-color HCR of UV-irradiated plasmids

UV HCR: XPA - deficient cell line at 16 hours



# FM-HCR for UV damaged Plasmids (Nucleotide Excision Repair)

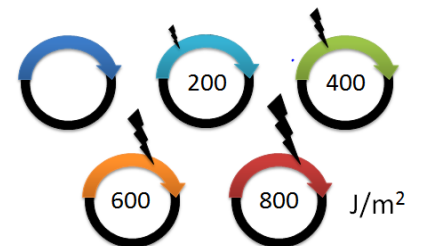


**FM-HCR**

**Fluorescence**  
**Multiplexed**

**Host**  
**Cell**

**Reactivation**



# Development and field-test validation of an assay for DNA repair in circulating human lymphocytes. *Cancer Res.*1991 51:5786-93.

**Athas, Hedayati, Matanoski, Farmer & GROSSMAN**

**Table 2** *Phenotype and plasmid HCR response ( $D_0$  and %CAT<sub>300</sub>) in XP homozygote, and apparent normal lymphoblastoid cell lines*

Cell line	Phenotype	Mean $\pm$ SD	n	95% CI <sup>a</sup>	%CAT <sub>300</sub> <sup>b</sup>
GM0536	Apparent normal	385 $\pm$ 60	3	235–534	59.4
GM0892	Apparent normal	595 $\pm$ 22	4	559–630	57.1
★ GM1953	Apparent normal	717 $\pm$ 78	3	523–91	67.7
★ GM1989	Apparent normal	594 $\pm$ 76	3	406–783	58.0
★ GM3657	Apparent normal	381 $\pm$ 15	4	357–405	47.0
GM2250	XP-A homozygote	90 $\pm$ 9	3	67–112	3.0
★ GM2344	XP-A homozygote	132 $\pm$ 9	3	110–155	6.4
★ GM2345	XP-A homozygote	90 $\pm$ 9	3	69–111	3.0
★ GM2246	XP-C homozygote	165 $\pm$ 19	4	134–195	22.1
★ GM2249	XP-C homozygote	256 $\pm$ 12	5	241–270	31.2
★ GM2253	XP-D homozygote	75 $\pm$ 5	3	62–88	1.8
GM2485	XP-D homozygote	97 $\pm$ 12	3	67–125	4.5
GM2450	XP-E homozygote	312 $\pm$ 42	5	260–364	

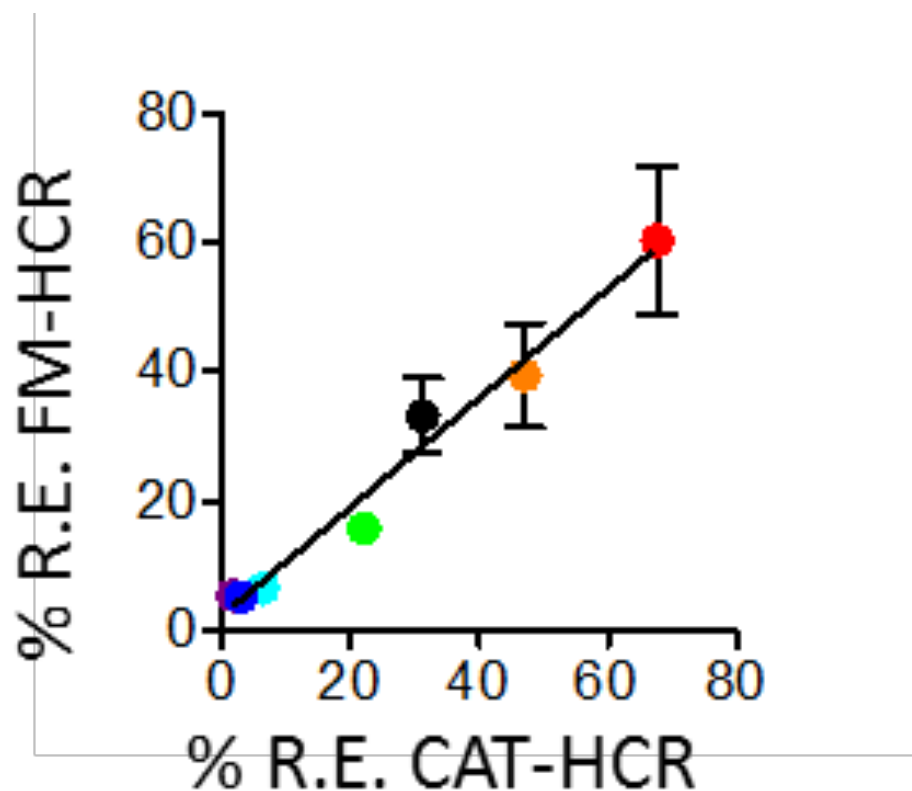
<sup>a</sup> CI, confidence interval.



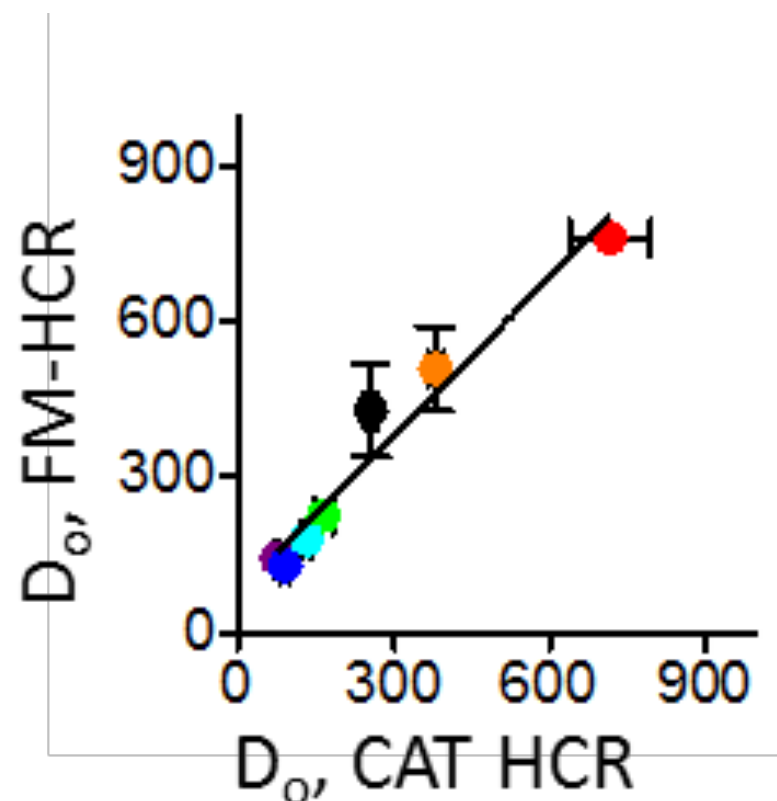
How does our **FM-HCR** data stack up against  
CAT-HCR, Grossman *et al.*, > 20 years ago?  
Cancer Research. 1991; **51** (21): 5786-93

# How does our FM-HCR data stack up against CAT-HCR, Grossman *et al.*, > 20 years ago?

Cancer Research. 1991; 51 (21): 5786-93

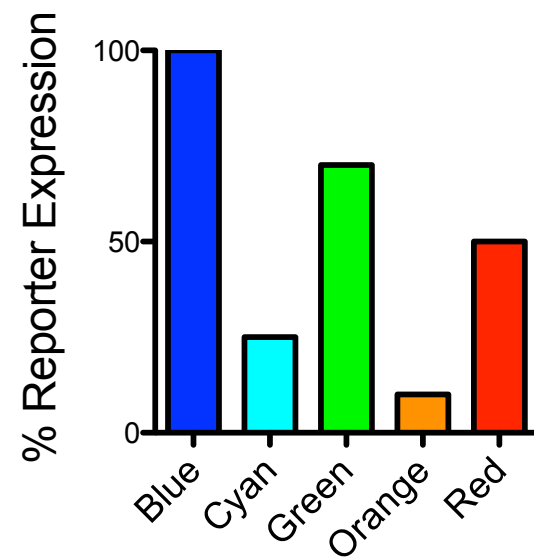
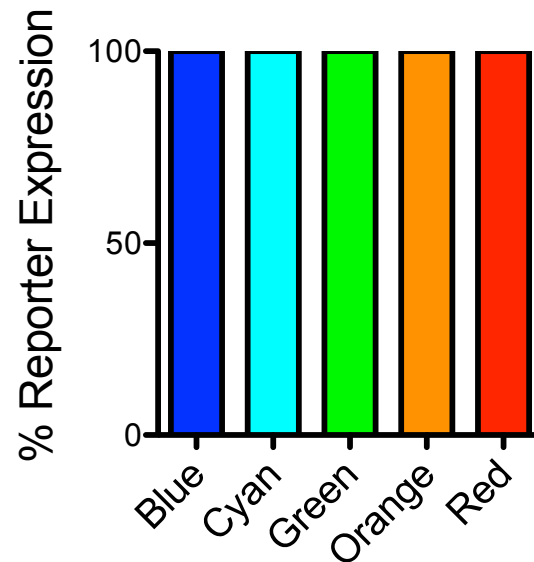
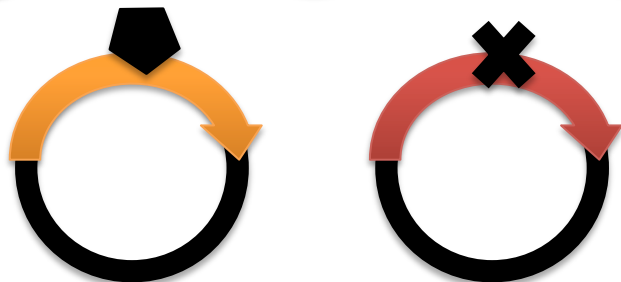
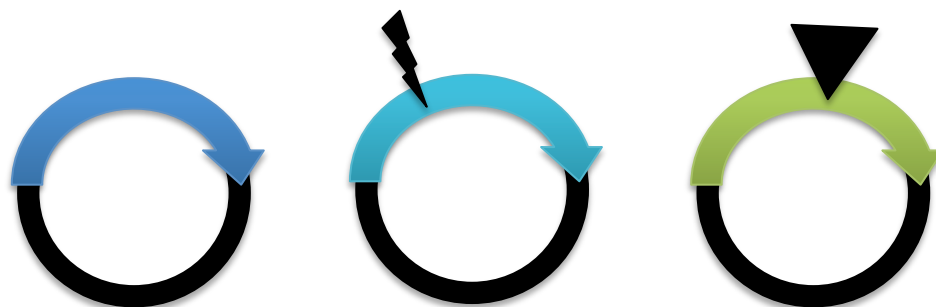
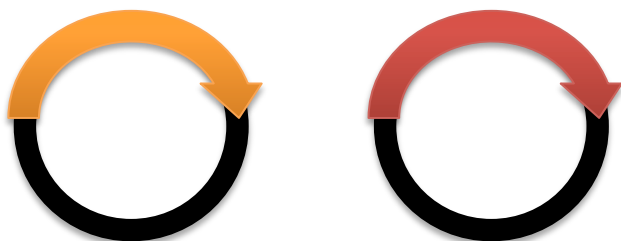
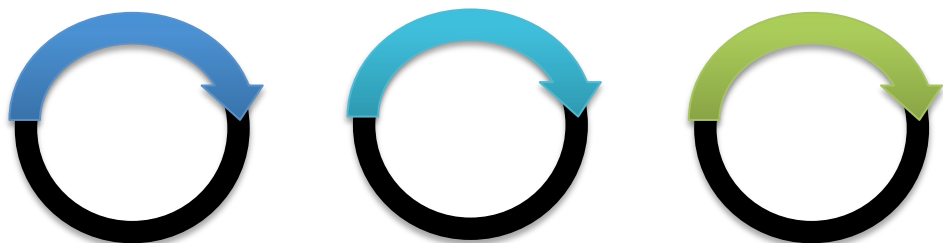


$R^2 = 0.92, p = 0.0006$



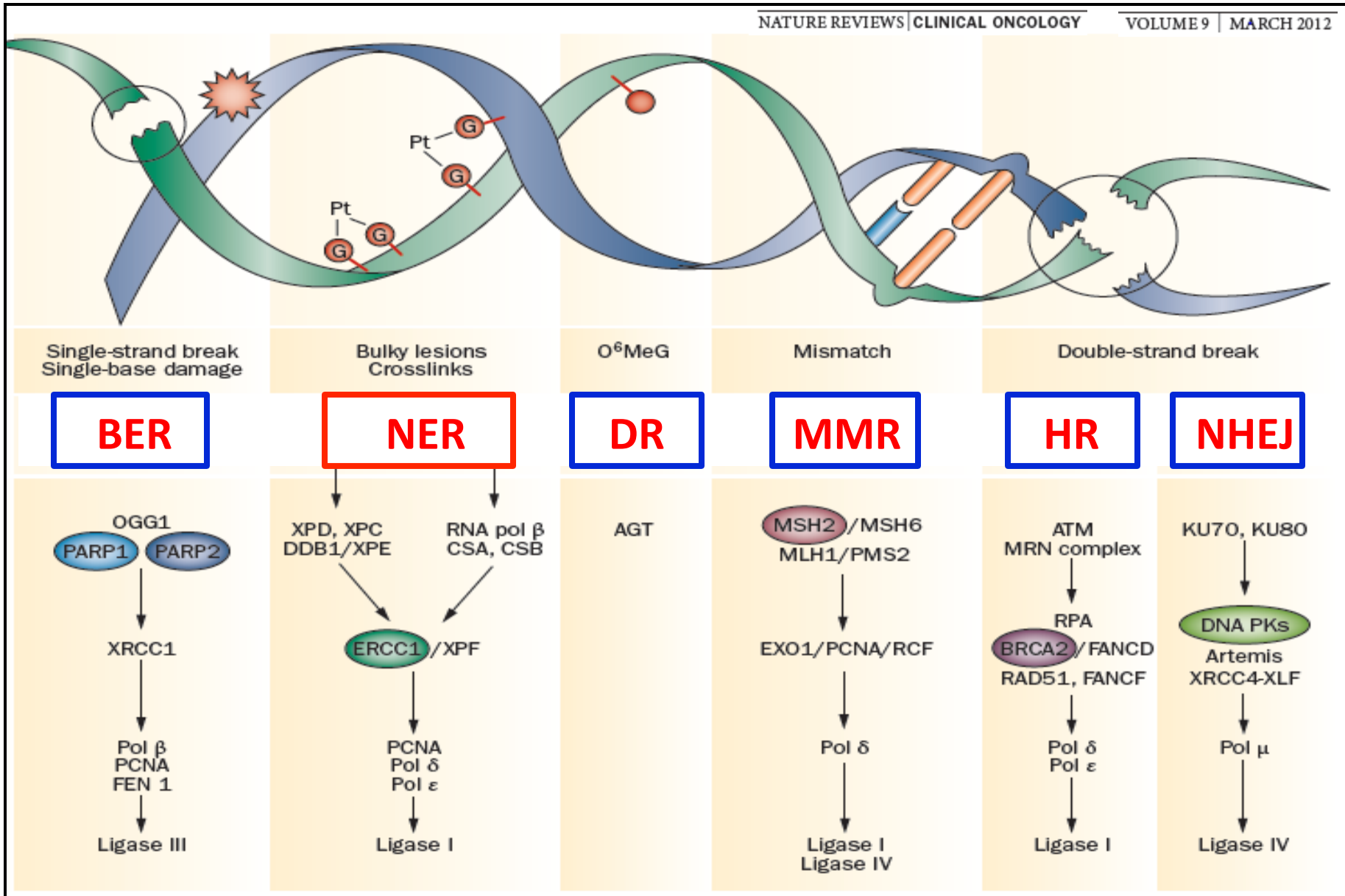
$R^2 = 0.92, p = 0.0001$

# 5 color HCR assay applications

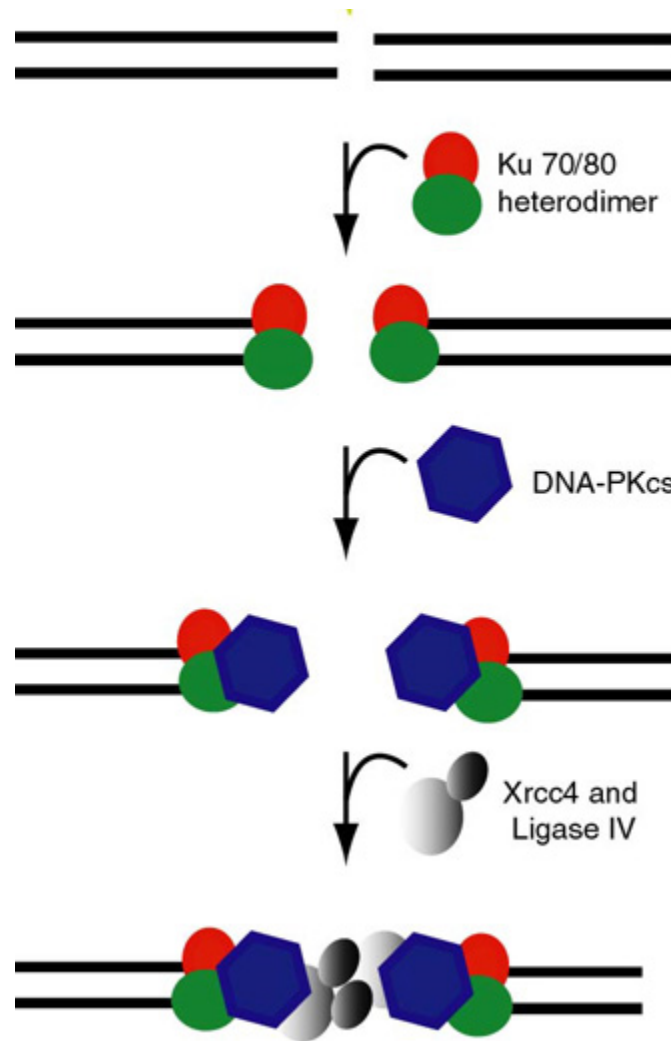


# Six Major DNA Repair Pathways

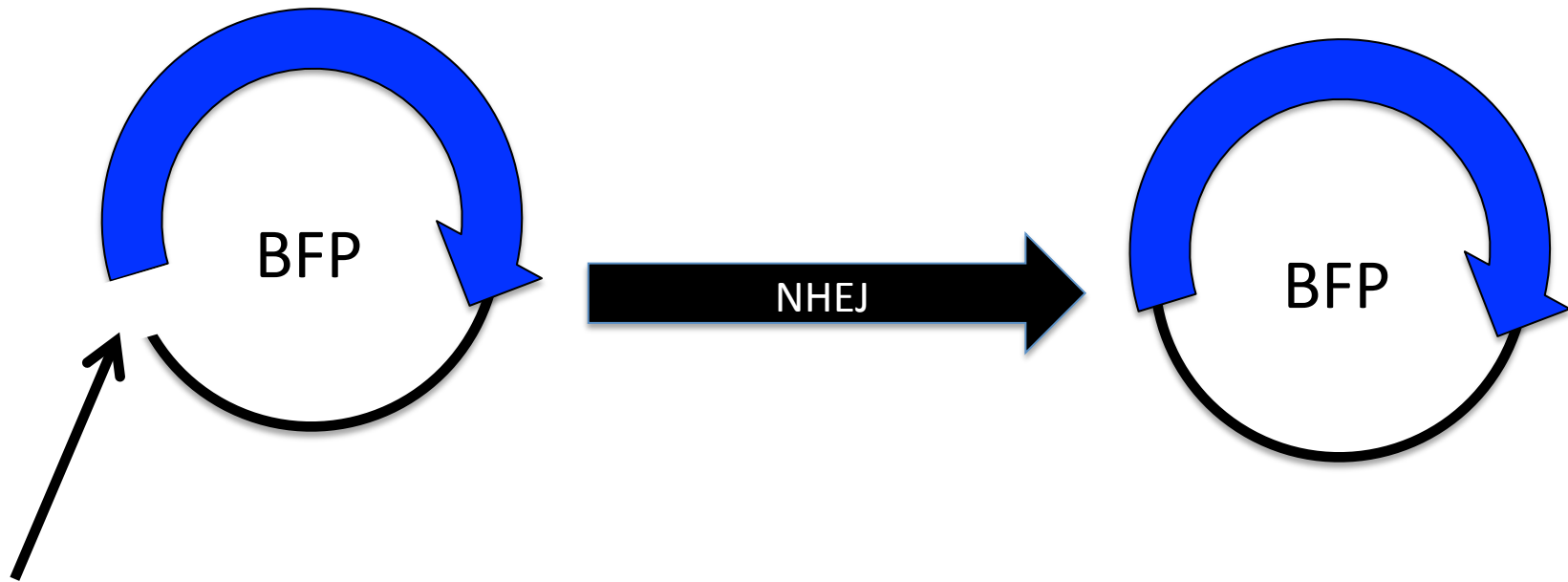
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# Non-Homologous End Joining (NHEJ) DNA Double Strand Break Repair

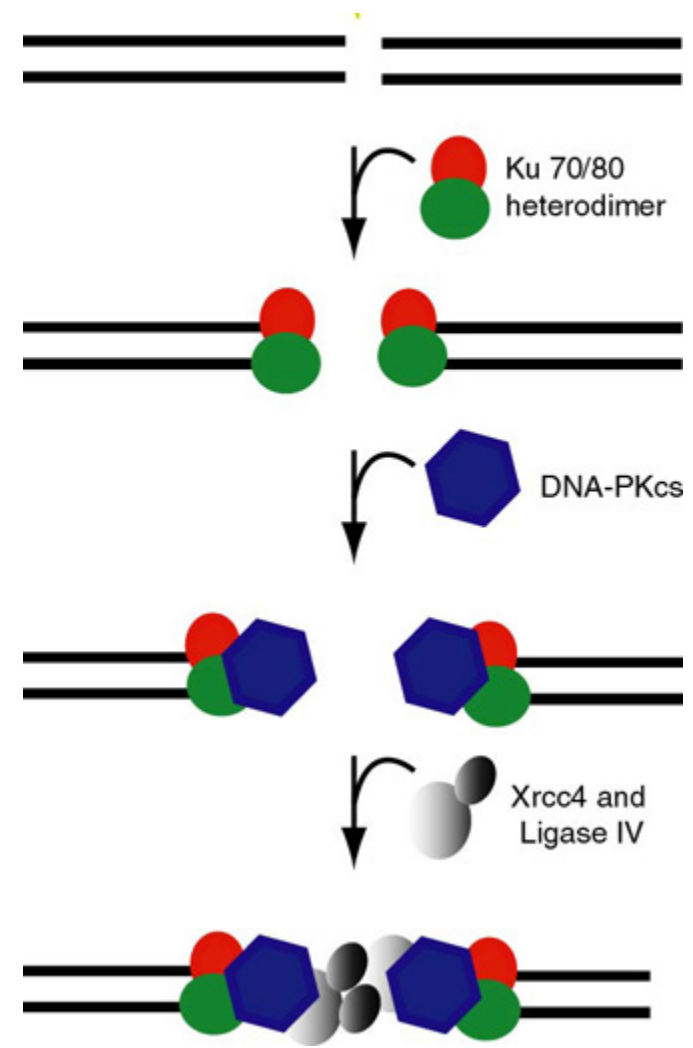
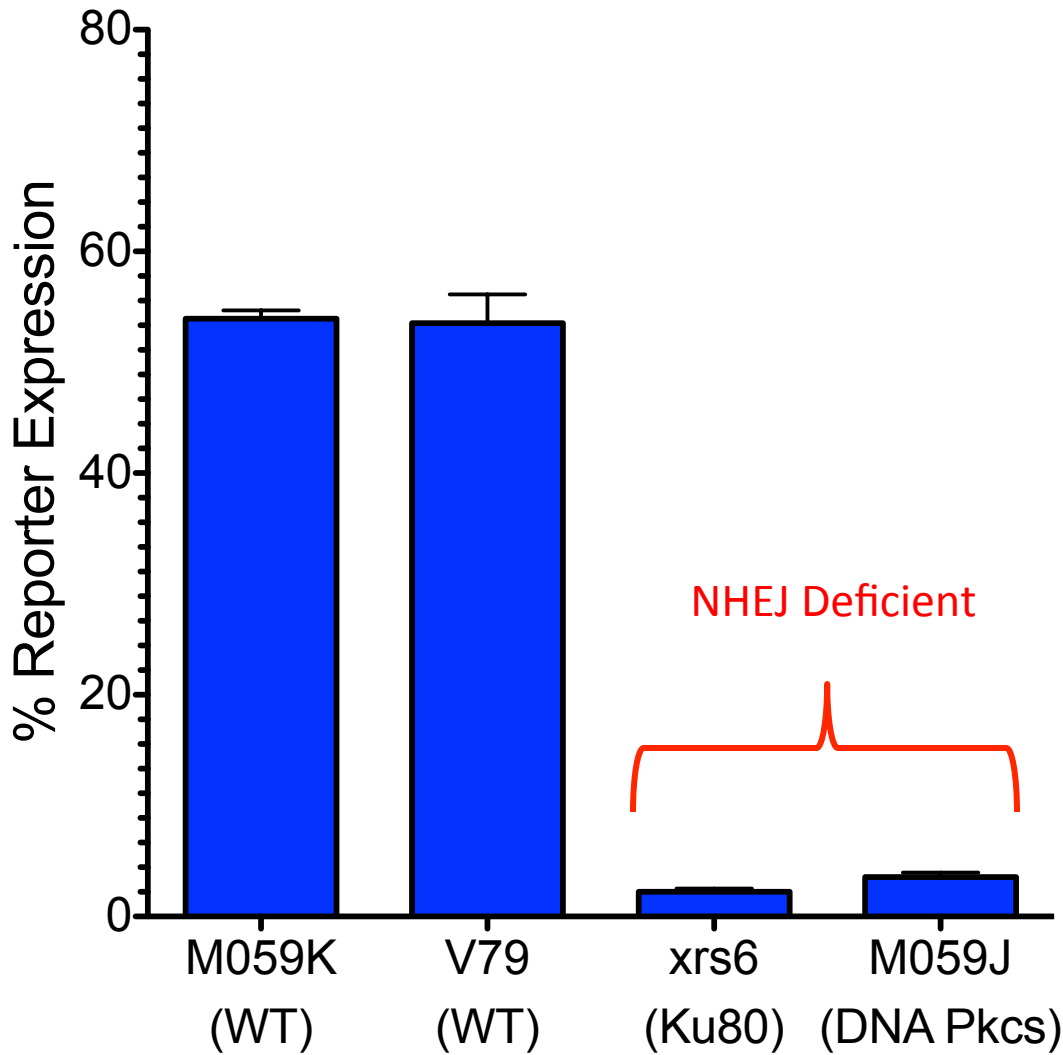


# Non-Homologous End Joining (NHEJ)

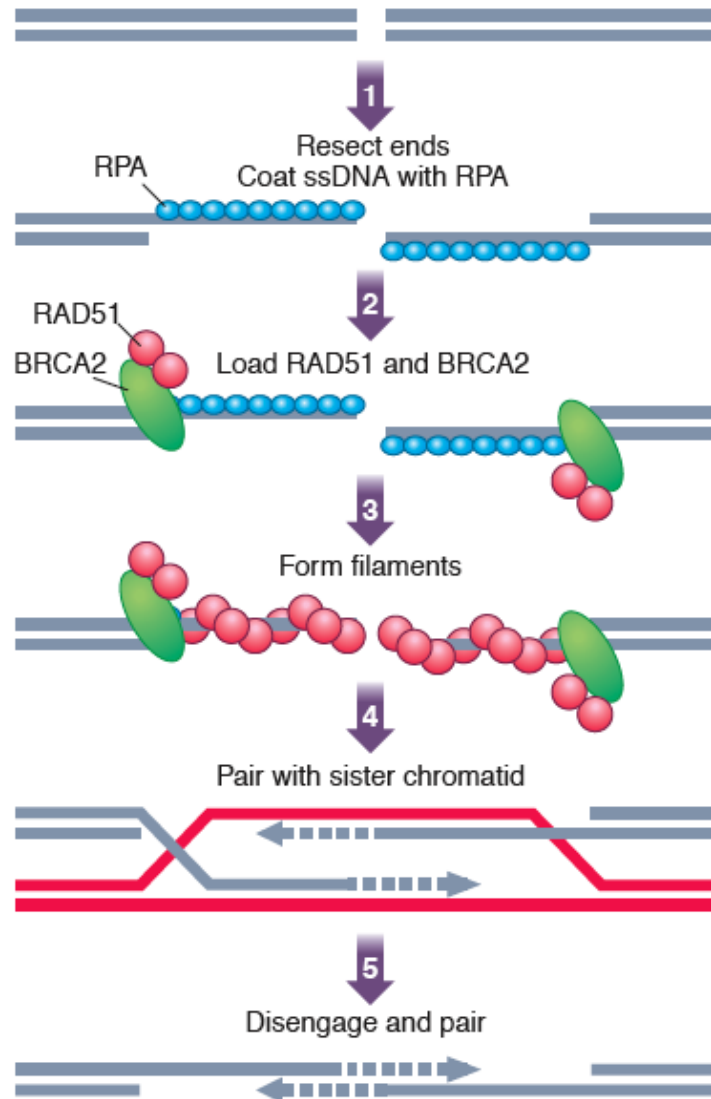


Substrate contains a blunt-end DSB in the 5' UTR

# NHEJ HCR in WT and NHEJ defective cells:

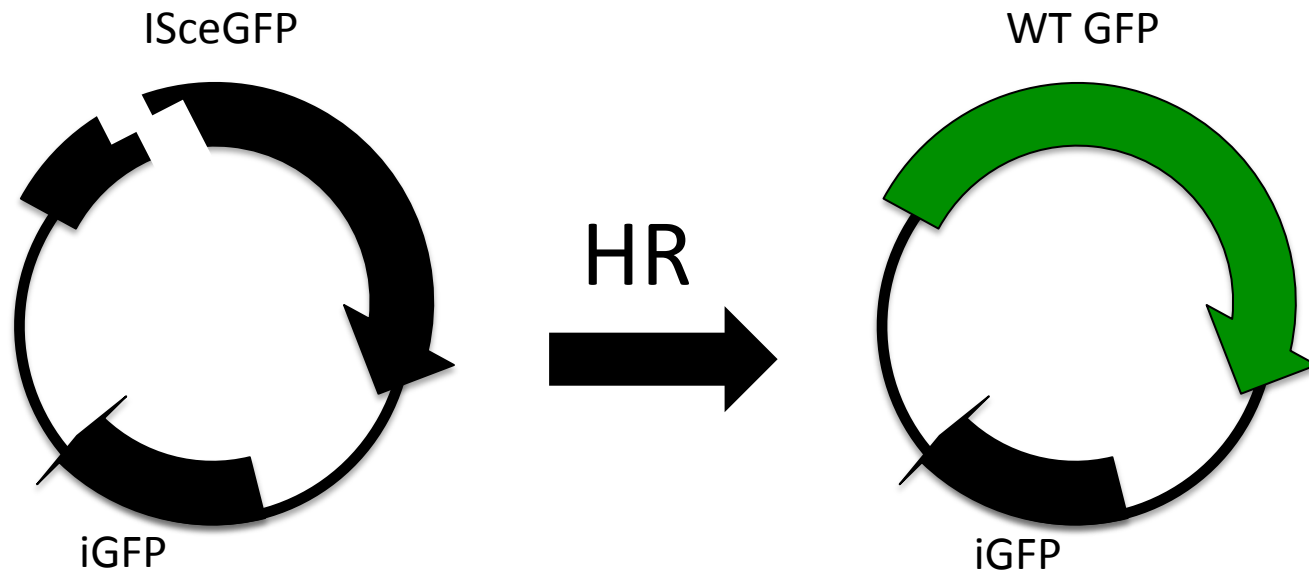


# DNA Double Strand Break Repair Homologous Recombination (HR)





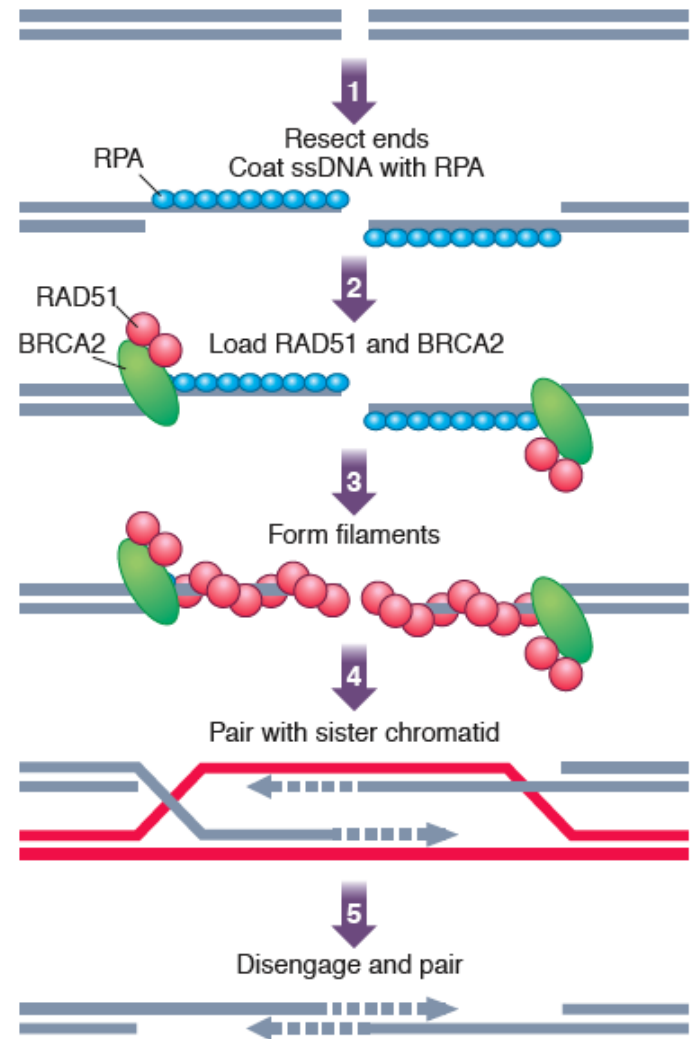
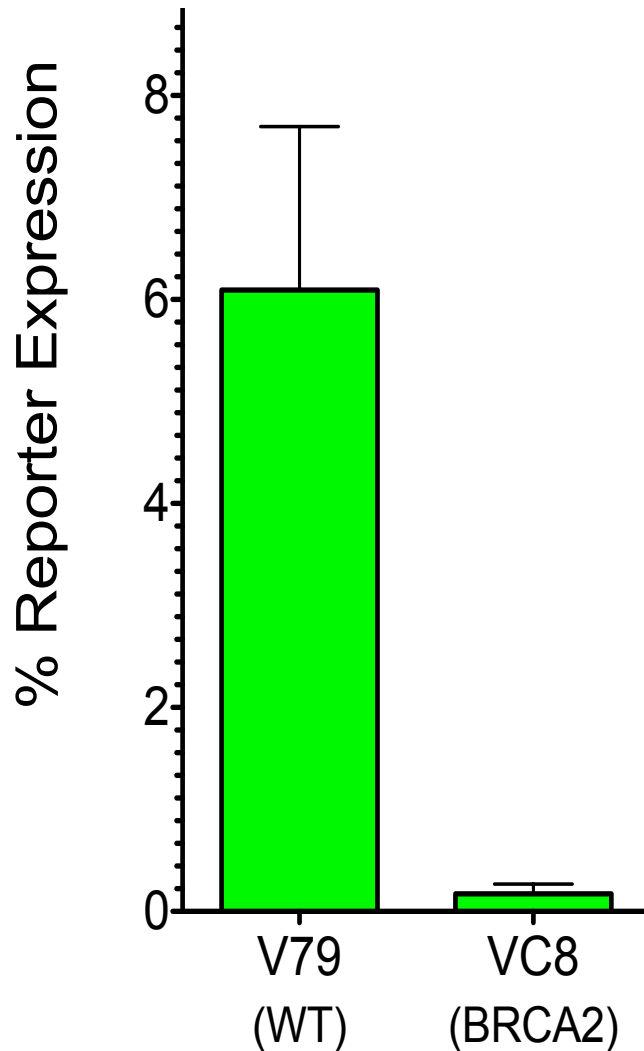
# DNA Double Strand Break Repair Homologous Recombination (HR)



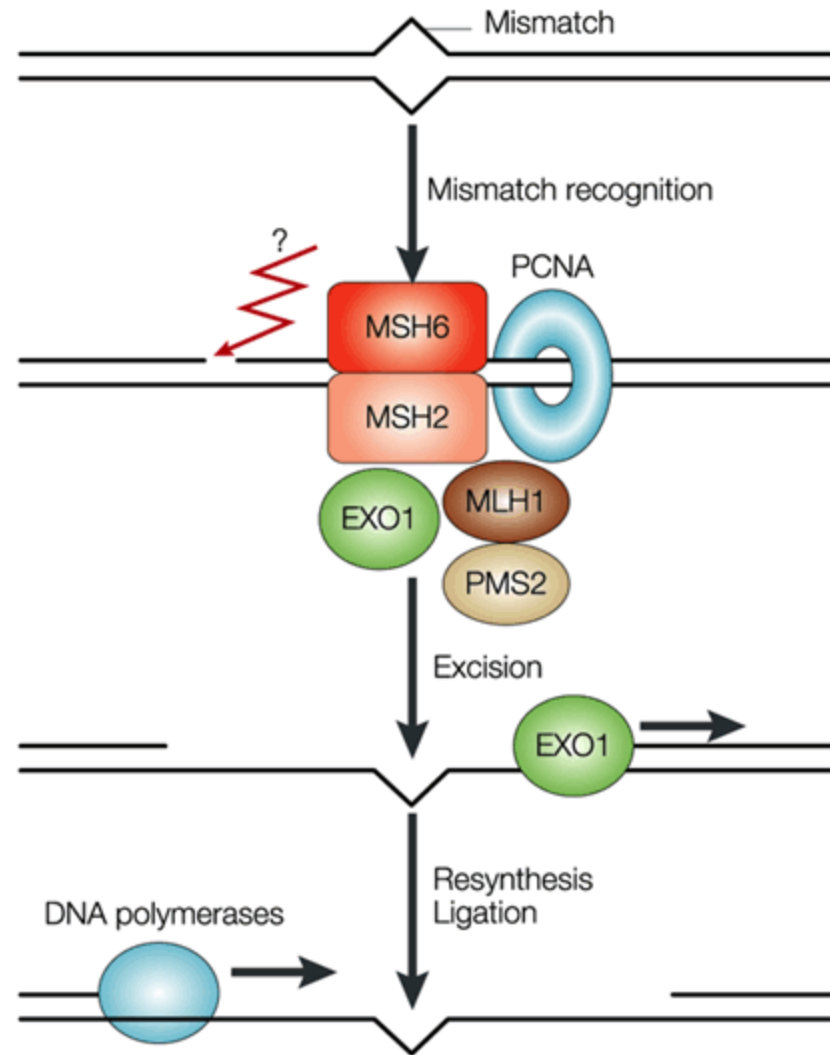
Adapted from Maria Jasin

Pierce et al. (1999) *Genes and Development* (13) 2633-2638

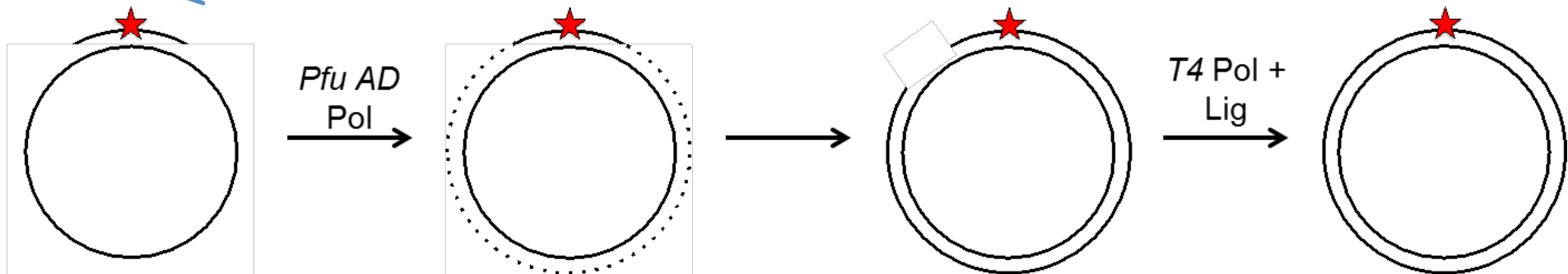
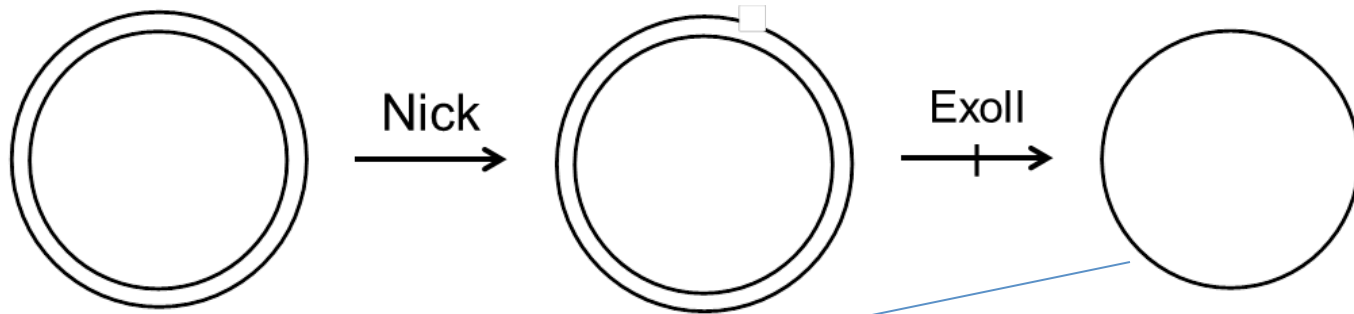
# FM-HCR for DSB repair by Homologous Recombination (HR)



# DNA Mismatch Repair (MMR)



# How to build a site-specific reporter? "Primer-Extension"

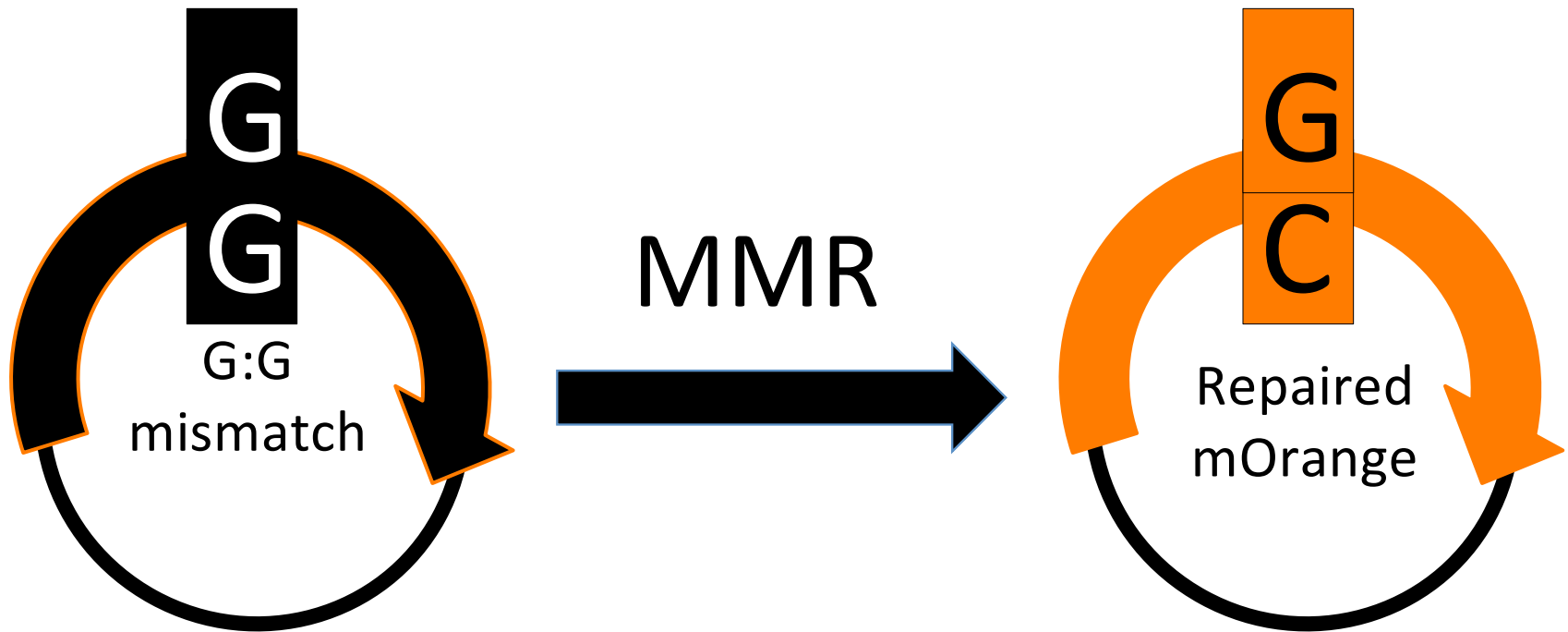


Original protocol from Baerenfaller et al. 2006 *Meth in enzymology*

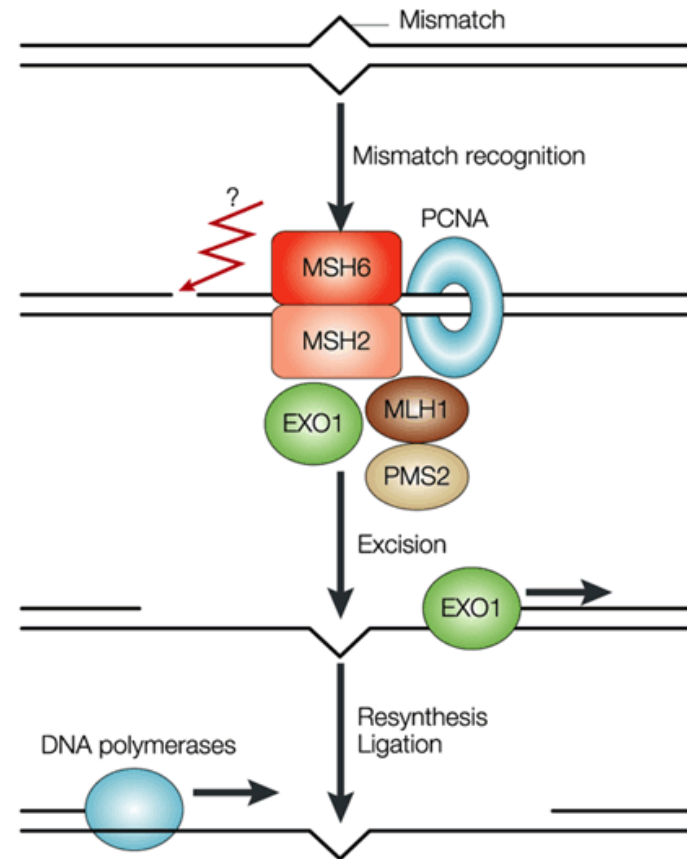
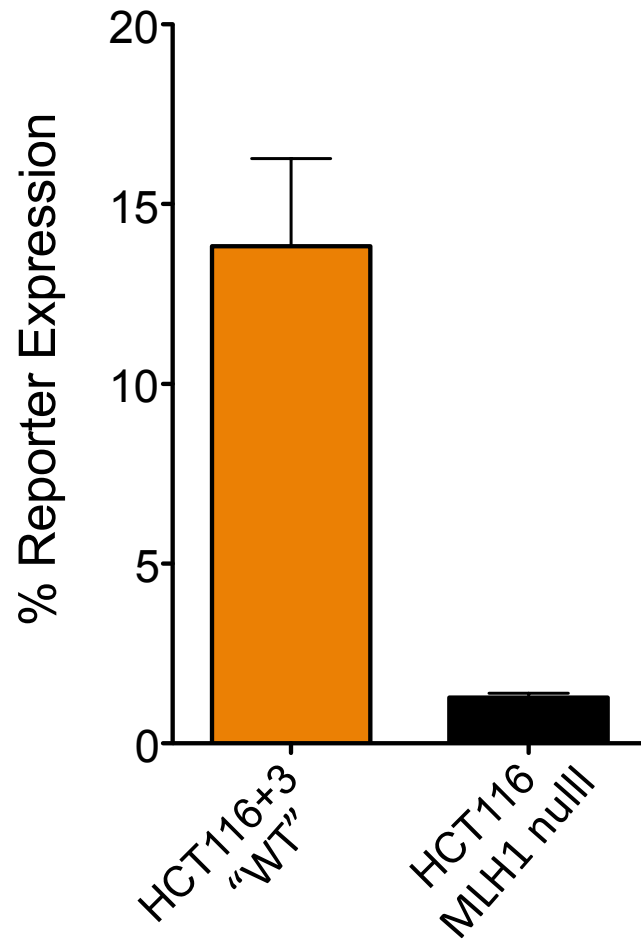
36

Modified and optimized by Alex Chaim, Zachary Nagel and Patrizia Mazzucato

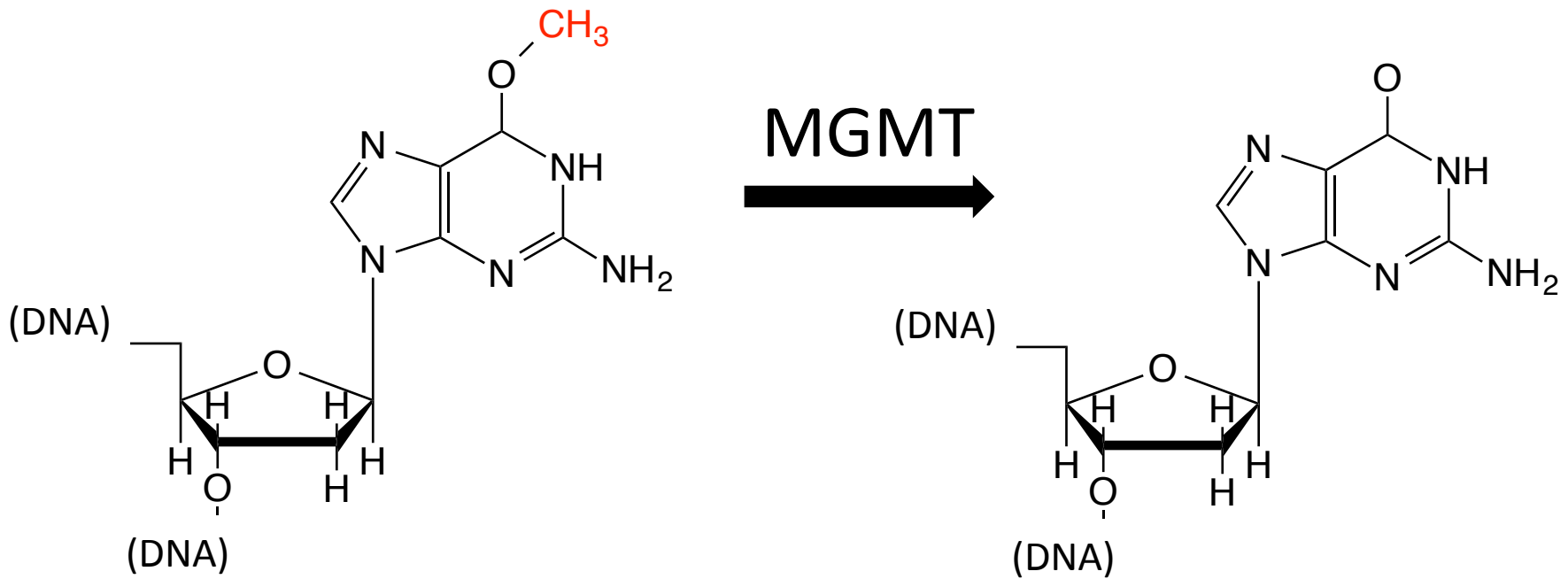
# DNA Mismatch Repair (MMR)



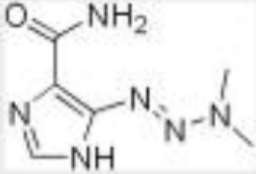
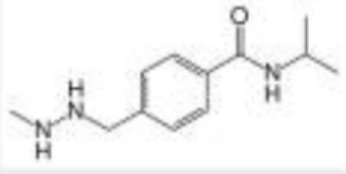
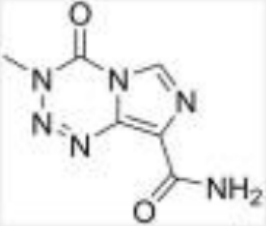
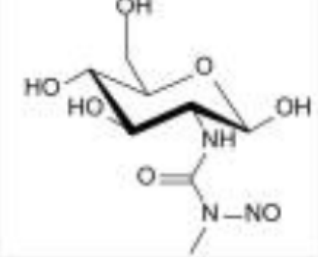
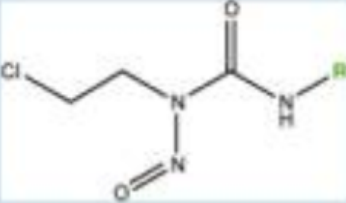
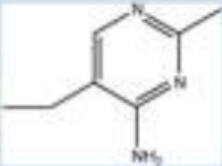

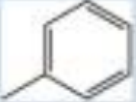
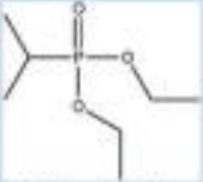
# FM-HCR for DNA Mismatch Repair



# $O^6$ -Methylguanine DNA Methyltransferase MGMT – Repairs by Direct Reversal (DR)

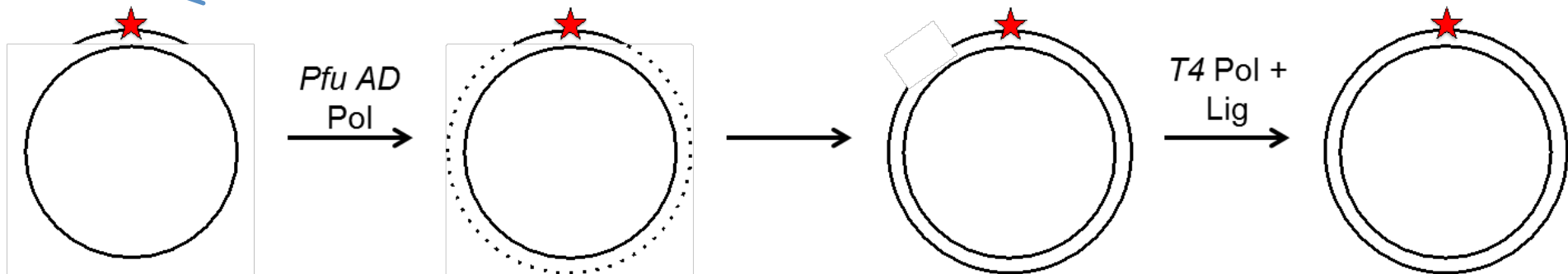
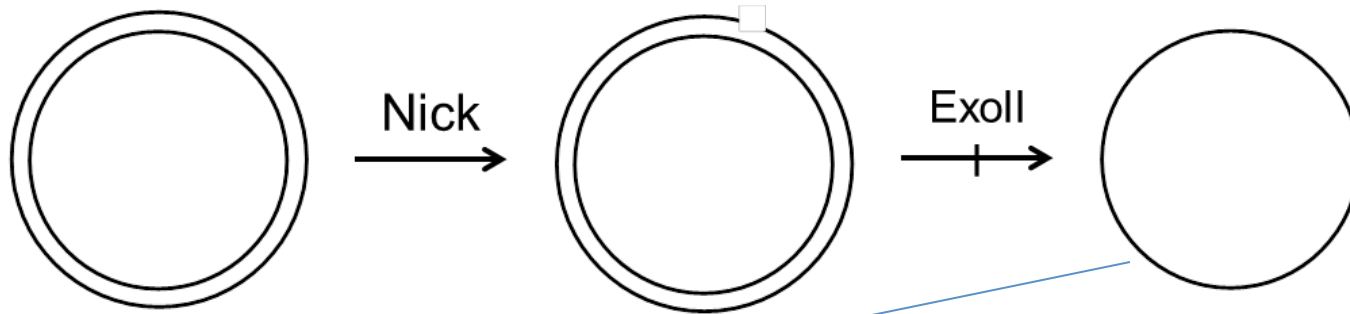


# Alkylating agents used in the cancer clinic

<b>a Monofunctional</b>				<b>Lesions<sup>1</sup></b>
<p><u>Triazene</u></p> <div style="display: flex; justify-content: space-around;"> <div style="text-align: center;">  <p><b>Dacarbazine</b> Metastatic melanoma Hodgkin's lymphoma Sarcoma</p> </div> <div style="text-align: center;">  <p><b>Procarbazine</b> Malignant gliomas Hodgkin's lymphoma</p> </div> <div style="text-align: center;">  <p><b>Temozolomide</b> Malignant gliomas</p> </div> </div> <p style="text-align: right;"><u>Nitrosourea</u></p> <div style="text-align: right;">  <p><b>Streptozotocin</b> Pancreatic islet cell cancer</p> </div>				<p>7meG 3meA O<sup>6</sup>meG</p>
<p><u>Chloroethylating Nitrosoureas</u></p> <div style="display: flex; justify-content: space-around;"> <div style="text-align: center;">  </div> <div style="text-align: center;"> <p>R=</p>  <p><b>ACNU</b> (Nimustine) Brain tumors Solid tumors</p> </div> <div style="text-align: center;">  <p><b>BCNU</b> (Carmustine) Brain tumors Lymphomas Melanoma</p> </div> <div style="text-align: center;">  <p><b>CCNU</b> (Lomustine) Brain tumors Lymphomas Melanoma</p> </div> <div style="text-align: center;">  <p><b>Fotemustine</b> Metastatic melanoma</p> </div> </div>				<p>7-alkylG O<sup>6</sup>Cl-ethylG N1,O<sup>6</sup>-EG G-C x-link G-G x-link</p>



# How to build a site-specific reporter? "Primer-Extension"



Original protocol from Baerenfaller et al. 2006 *Meth in enzymology*

41

Modified and optimized by Alex Chaim, Zachary Nagel and Patrizia Mazzucato

# DNA lesions from an RNA polymerase perspective

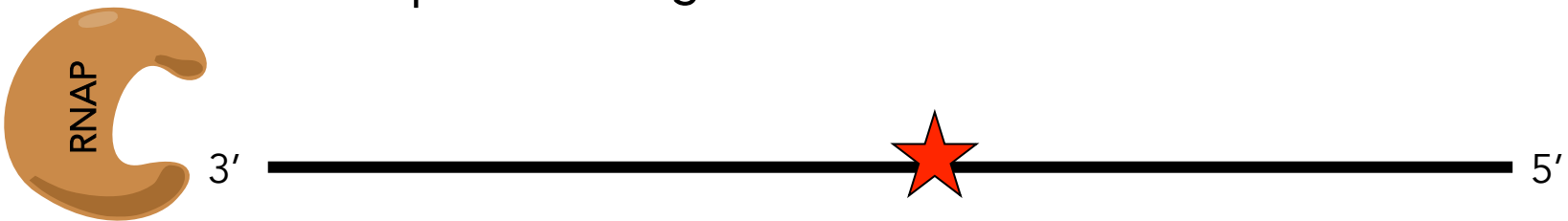
Block Transcription



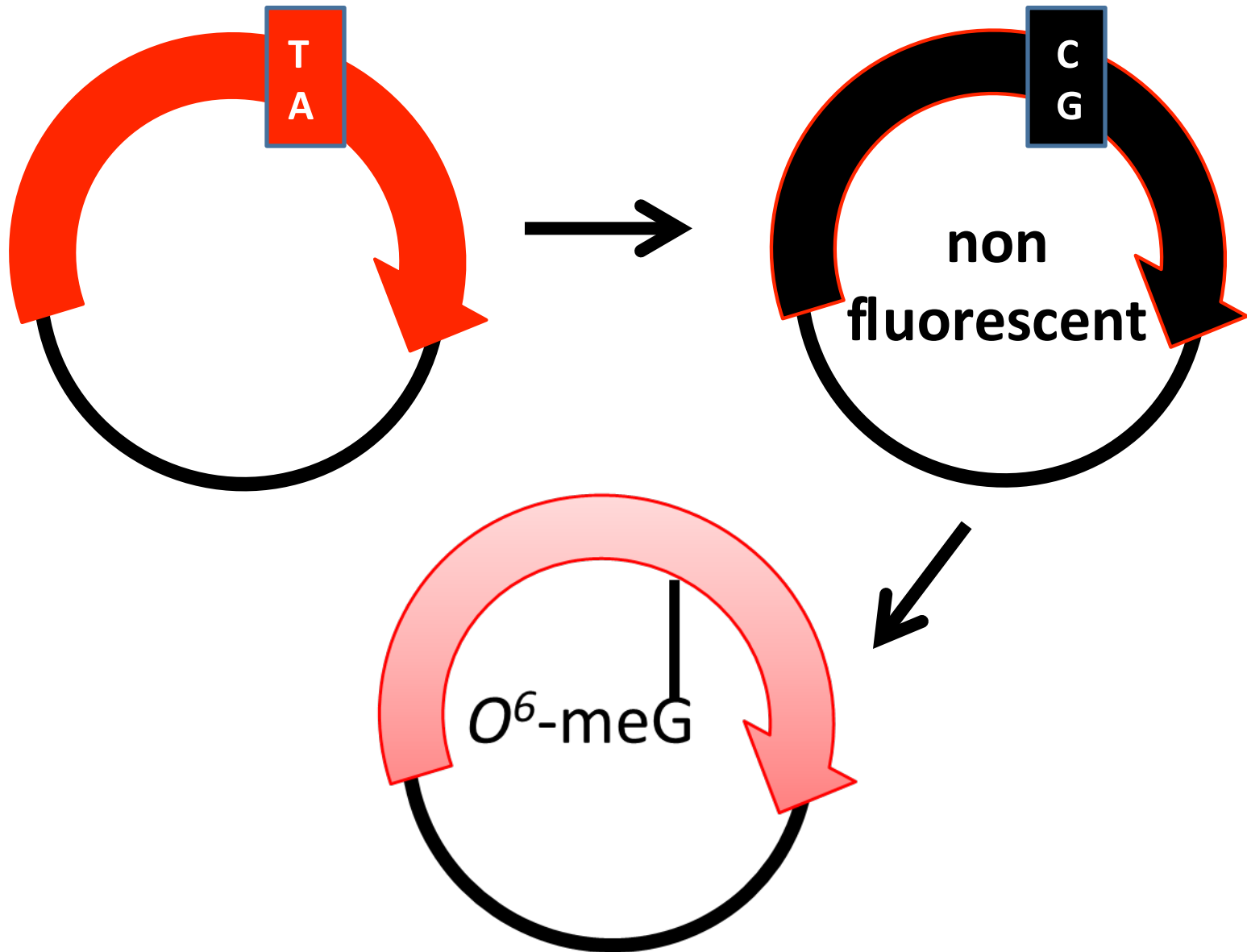
Error-free Bypass



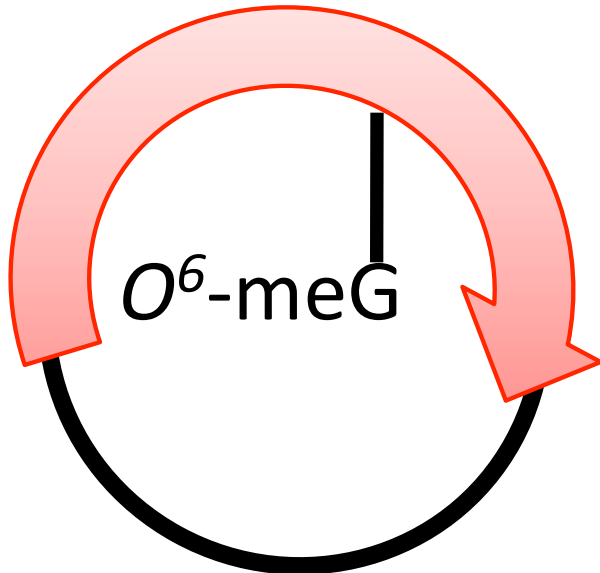
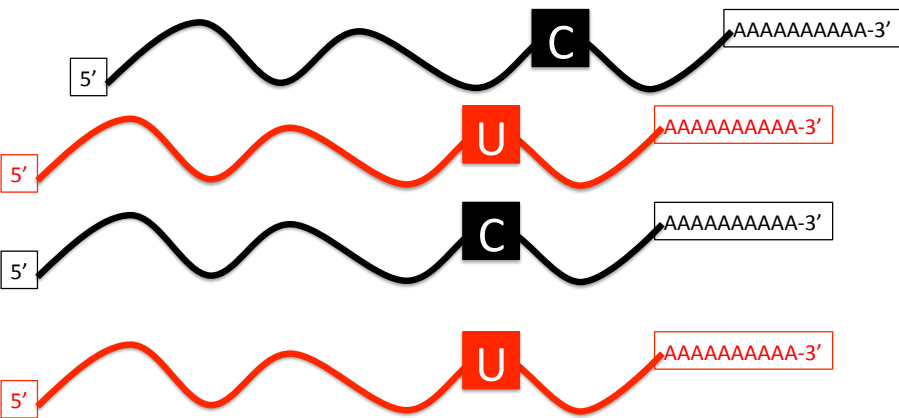
Transcriptional Mutagenesis



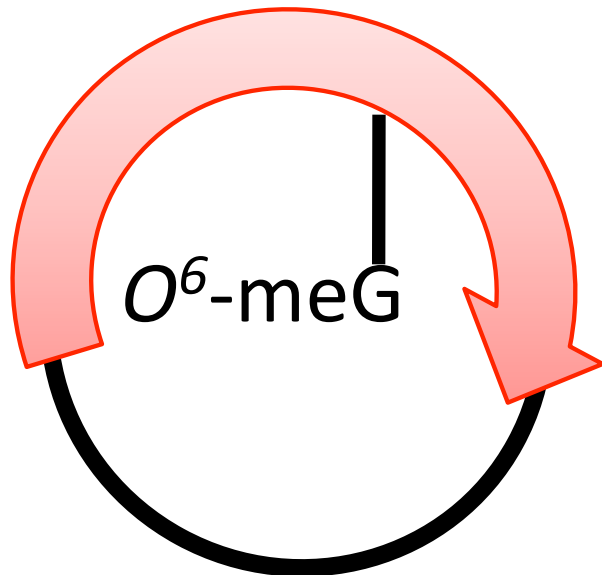
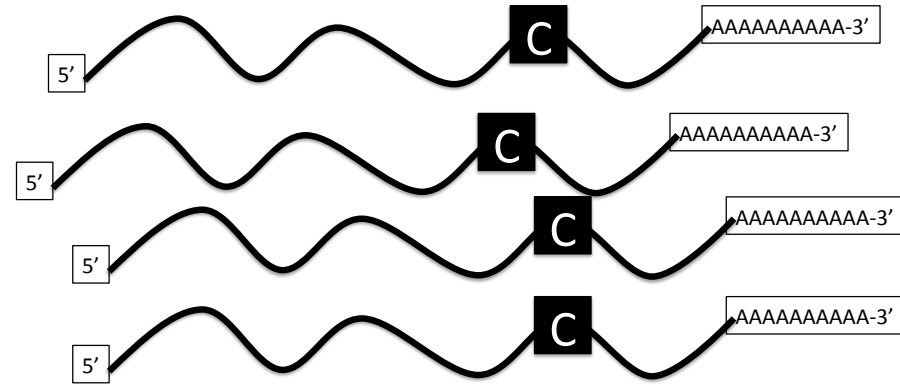
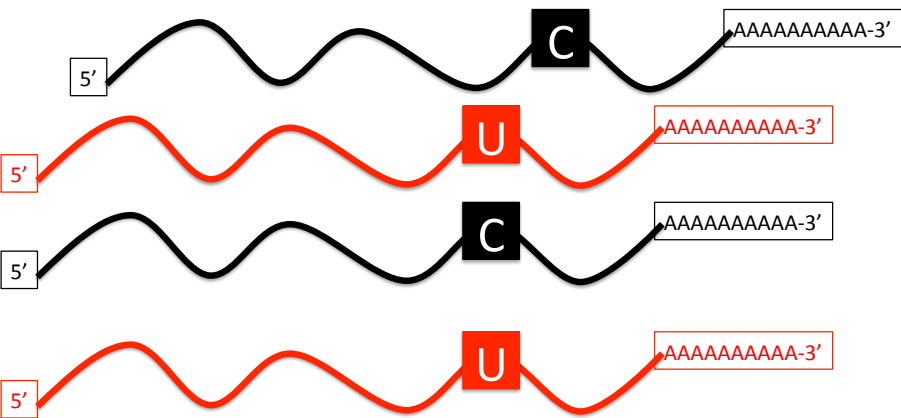
# Manipulation of the RFP Reporter Gene



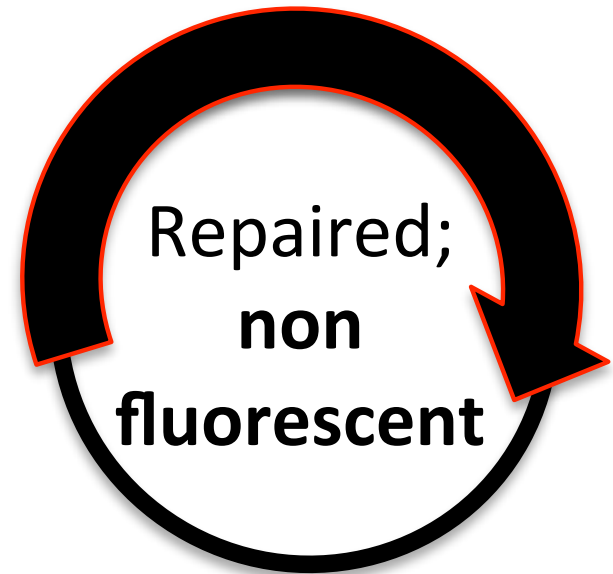
# RNA Pol II occasionally inserts U at $O^6$ -meG Transcriptional Mutagenesis



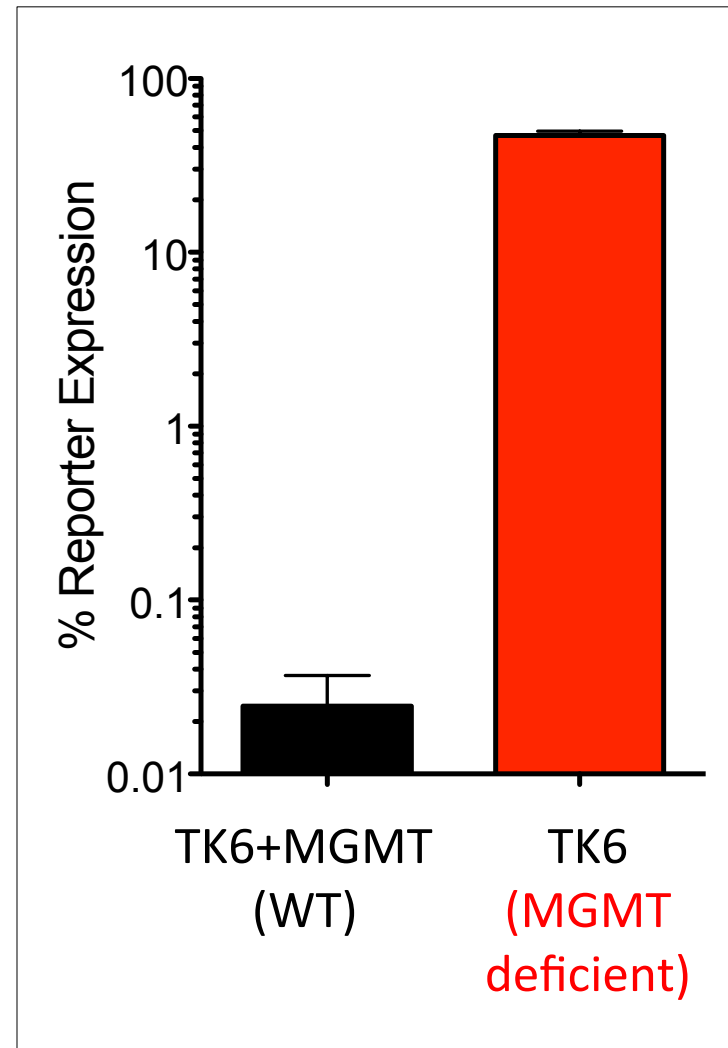
# RNA Pol II occasionally inserts U at $O^6$ -meG Transcriptional Mutagenesis



**MGMT**  
Direct Reversal

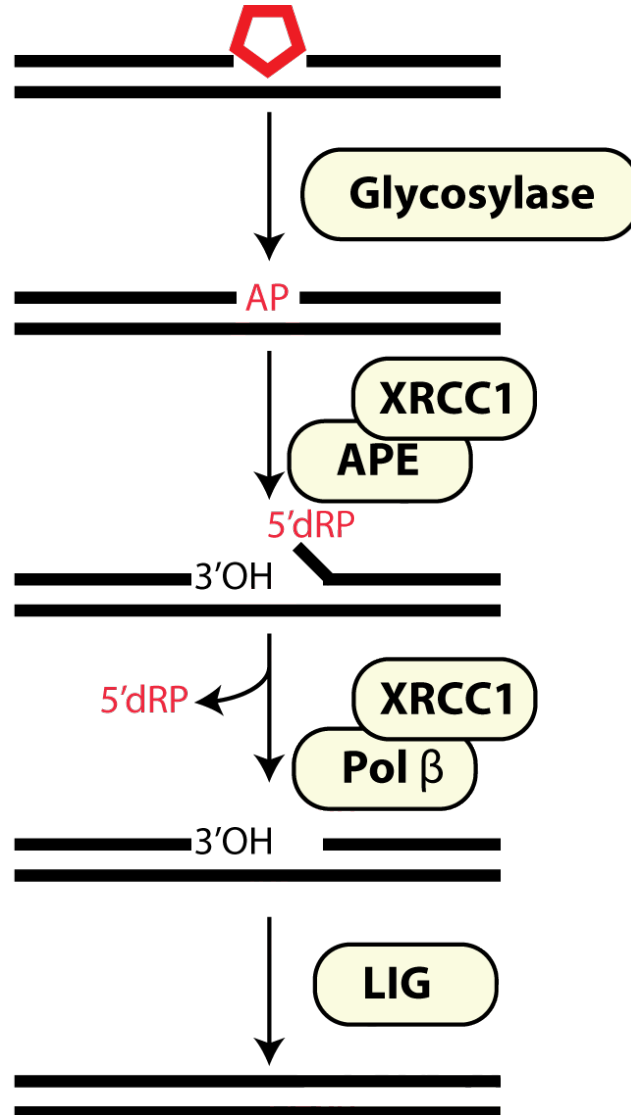


# FM-HCR for $O^6$ -MeG repair via Direct Reversal (DR) by MGMT

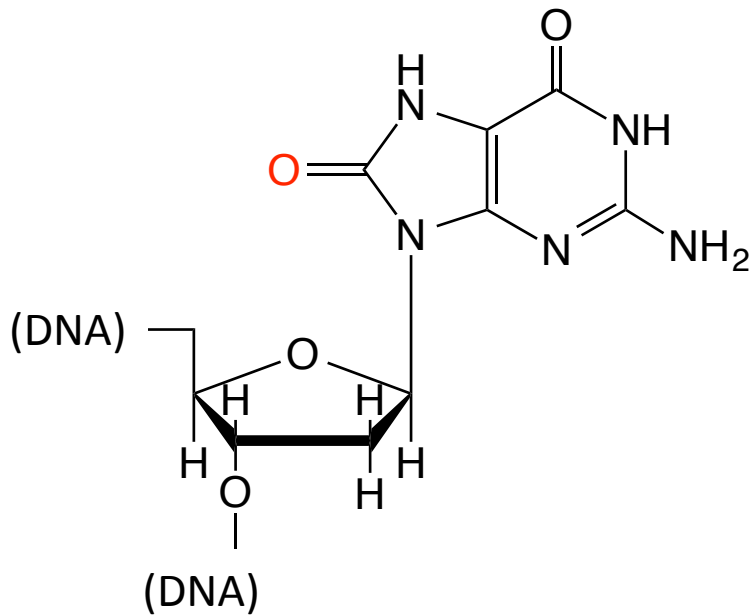


# Base Excision Repair (BER)

For oxidized, deaminated and alkylated base damage



# Base excision repair of 8-oxoG



8-oxo-guanine

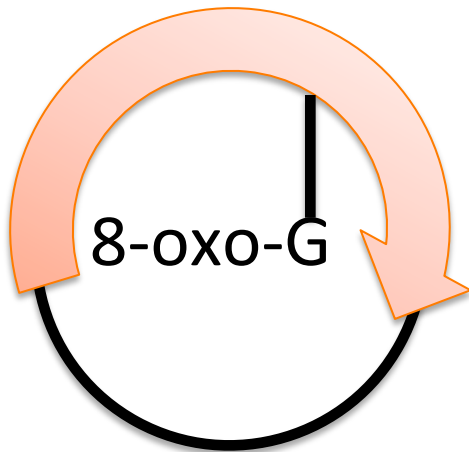
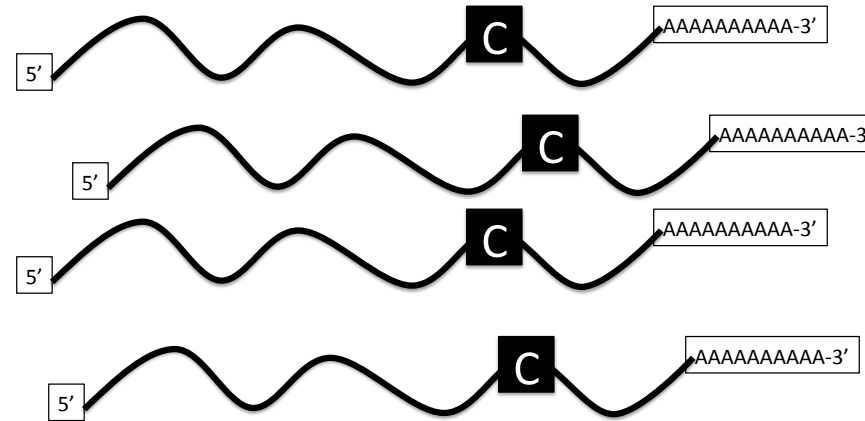
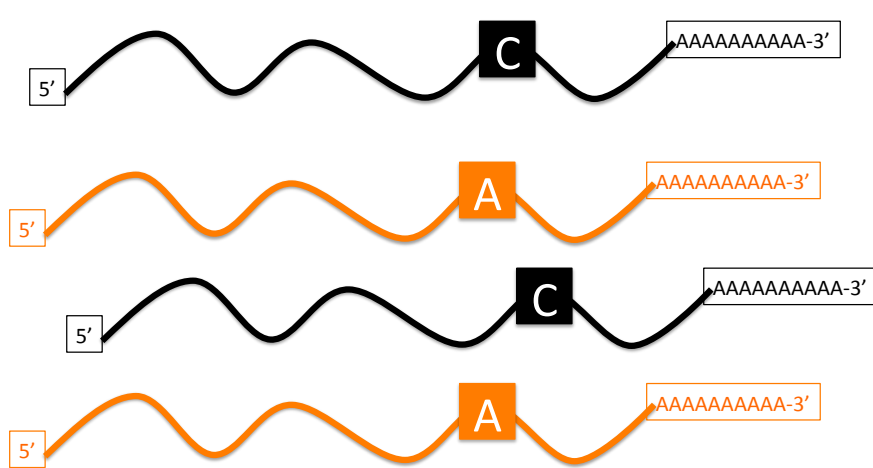


OGG1 DNA Glycosylase

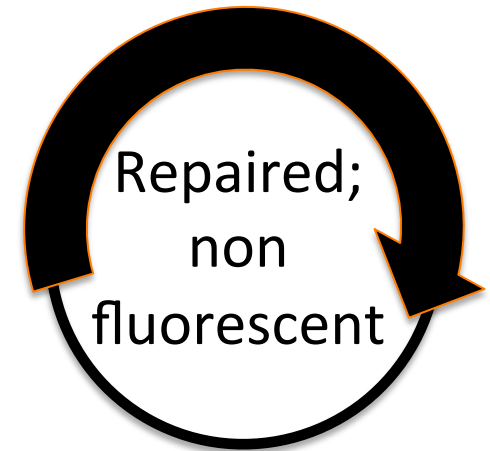


# RNA Pol II occasionally inserts A at 8-oxoG

## Transcriptional Mutagenesis

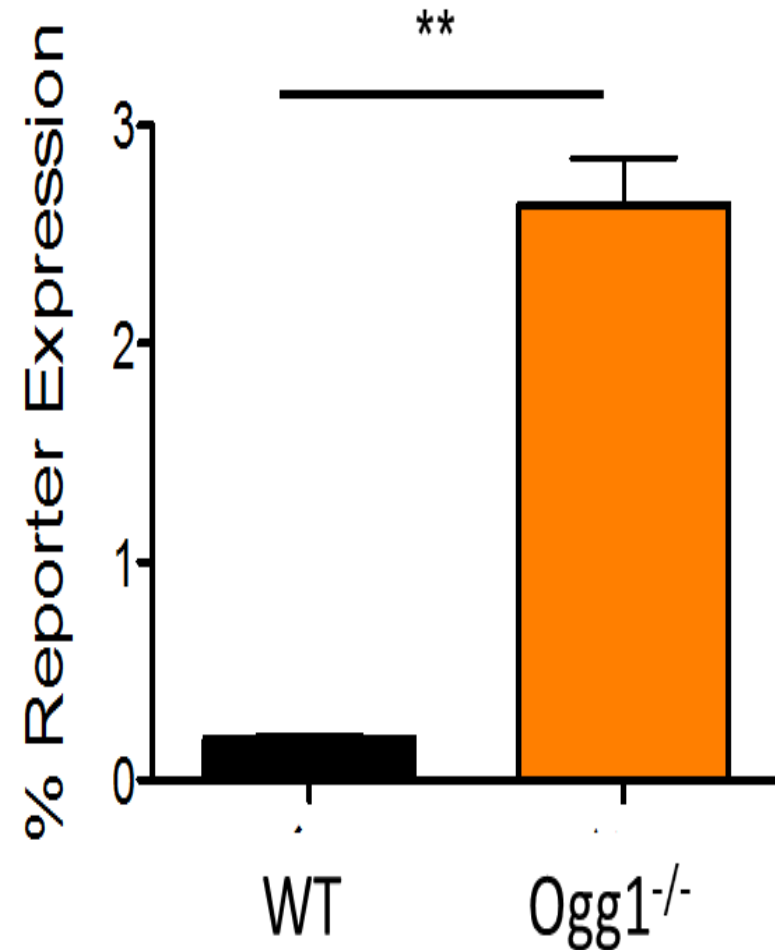


Base Excision Repair

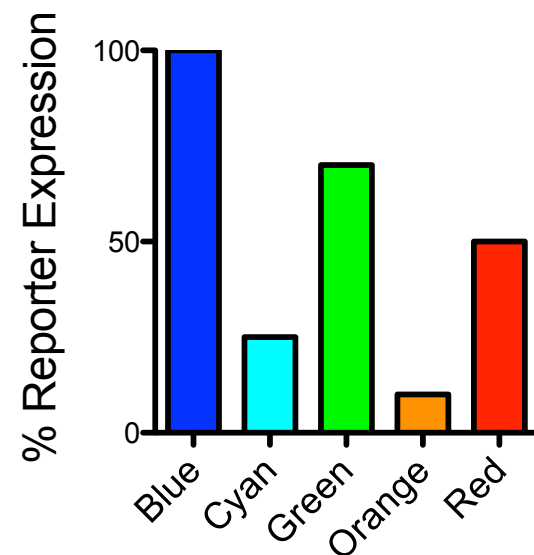
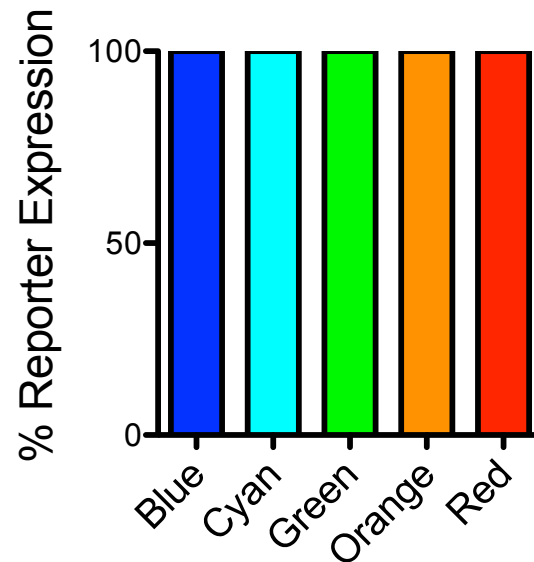
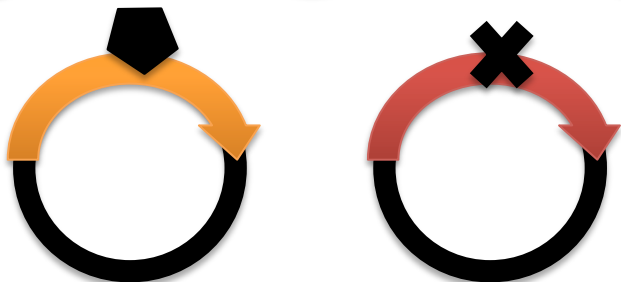
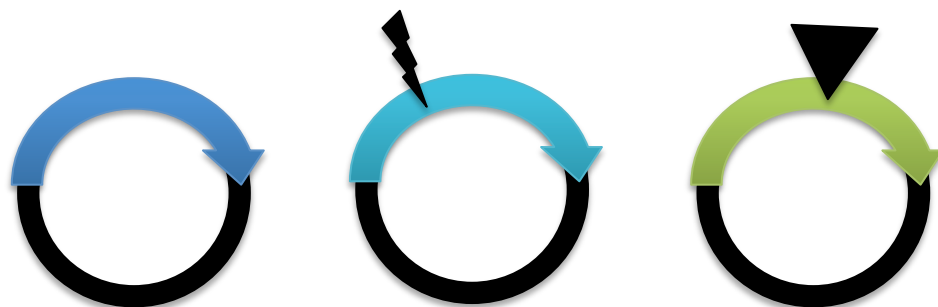
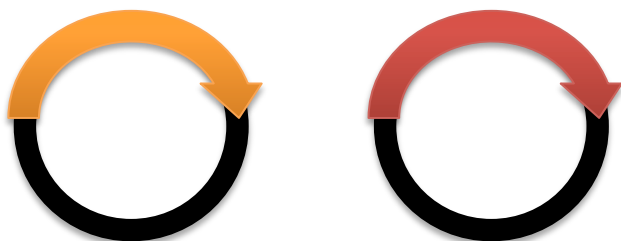
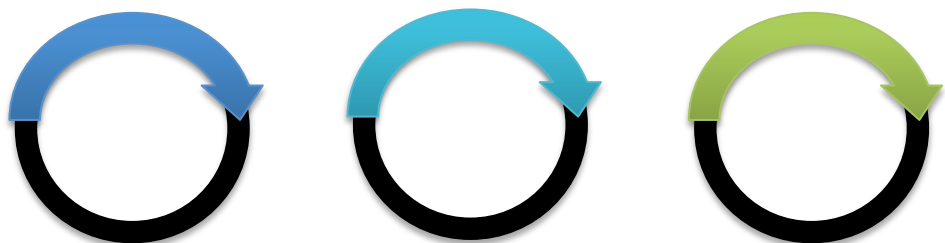


Repaired;  
non  
fluorescent

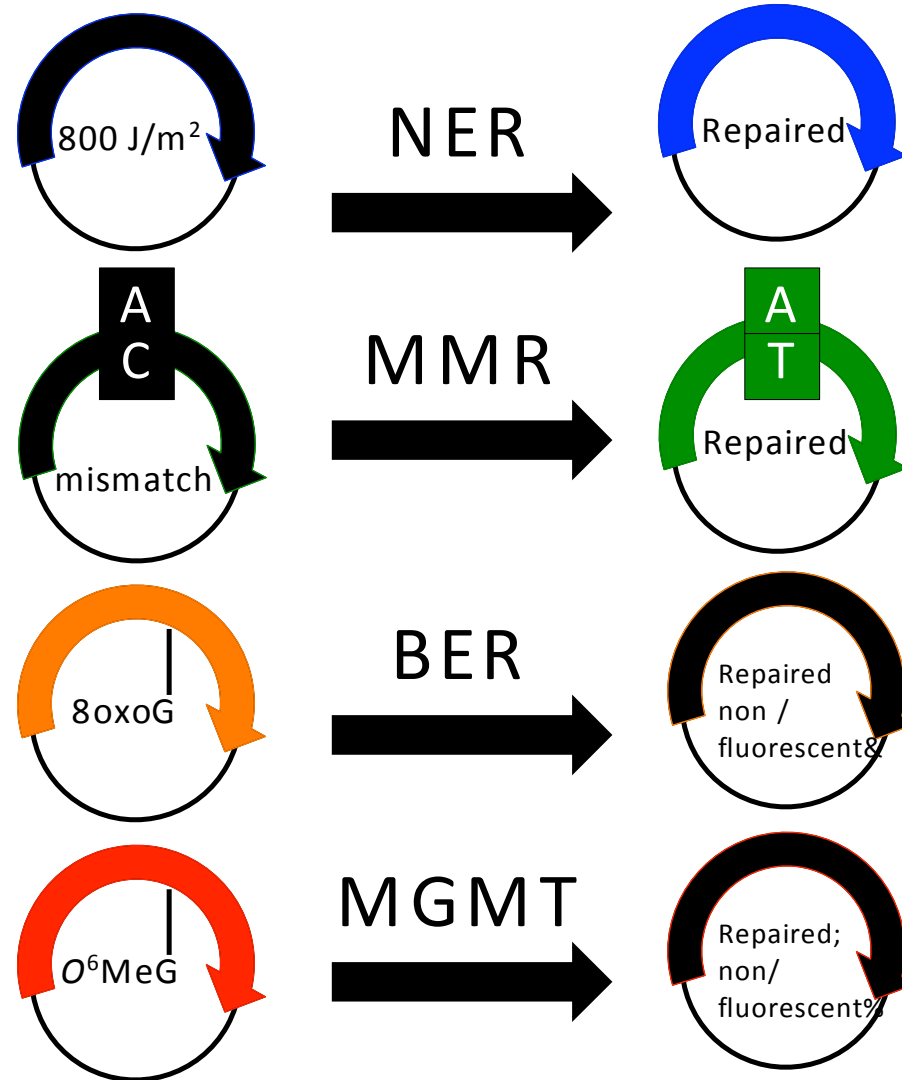
# FM-HCR for Ogg1-initiated BER



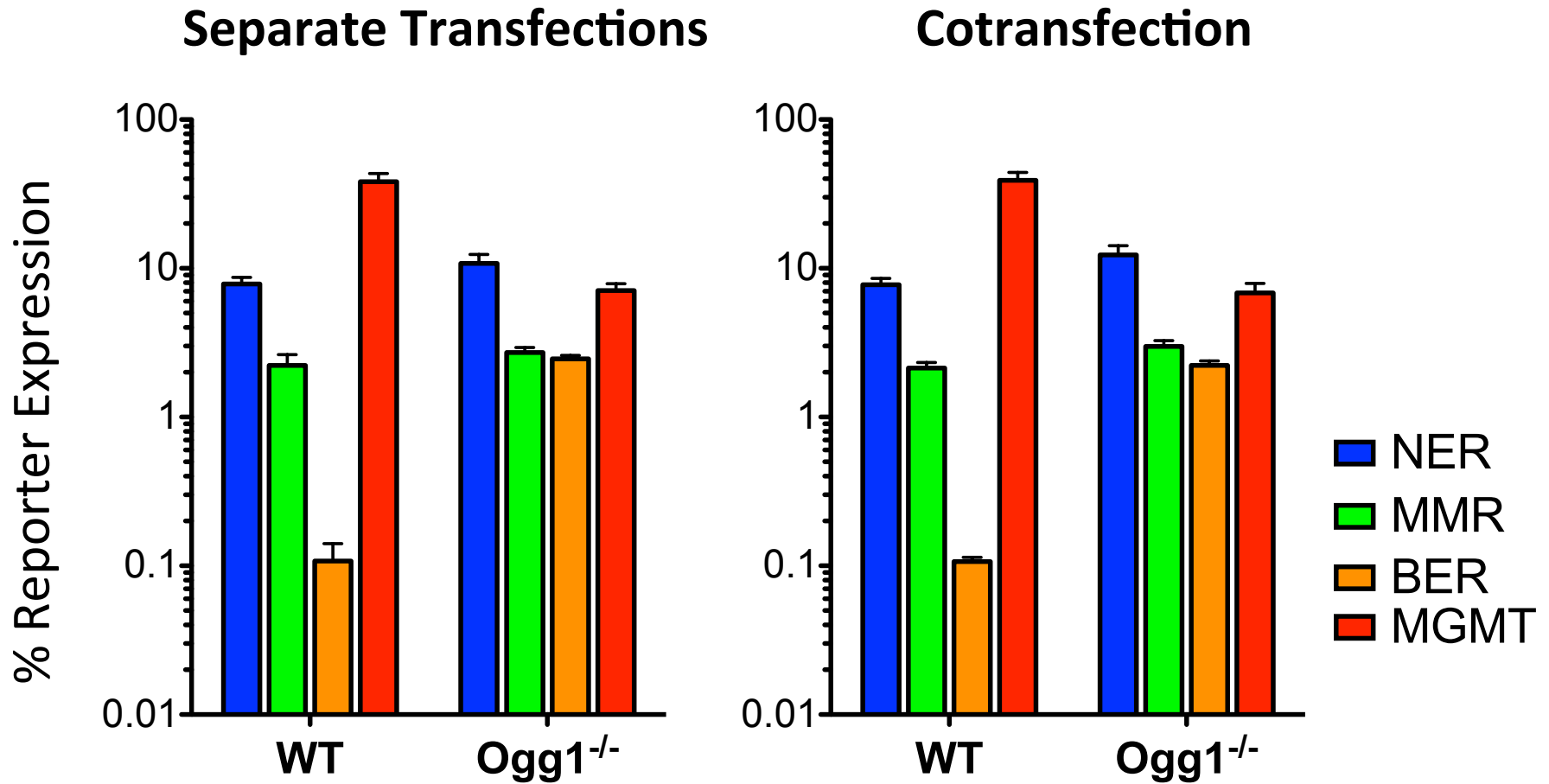
# 5 color HCR assay applications



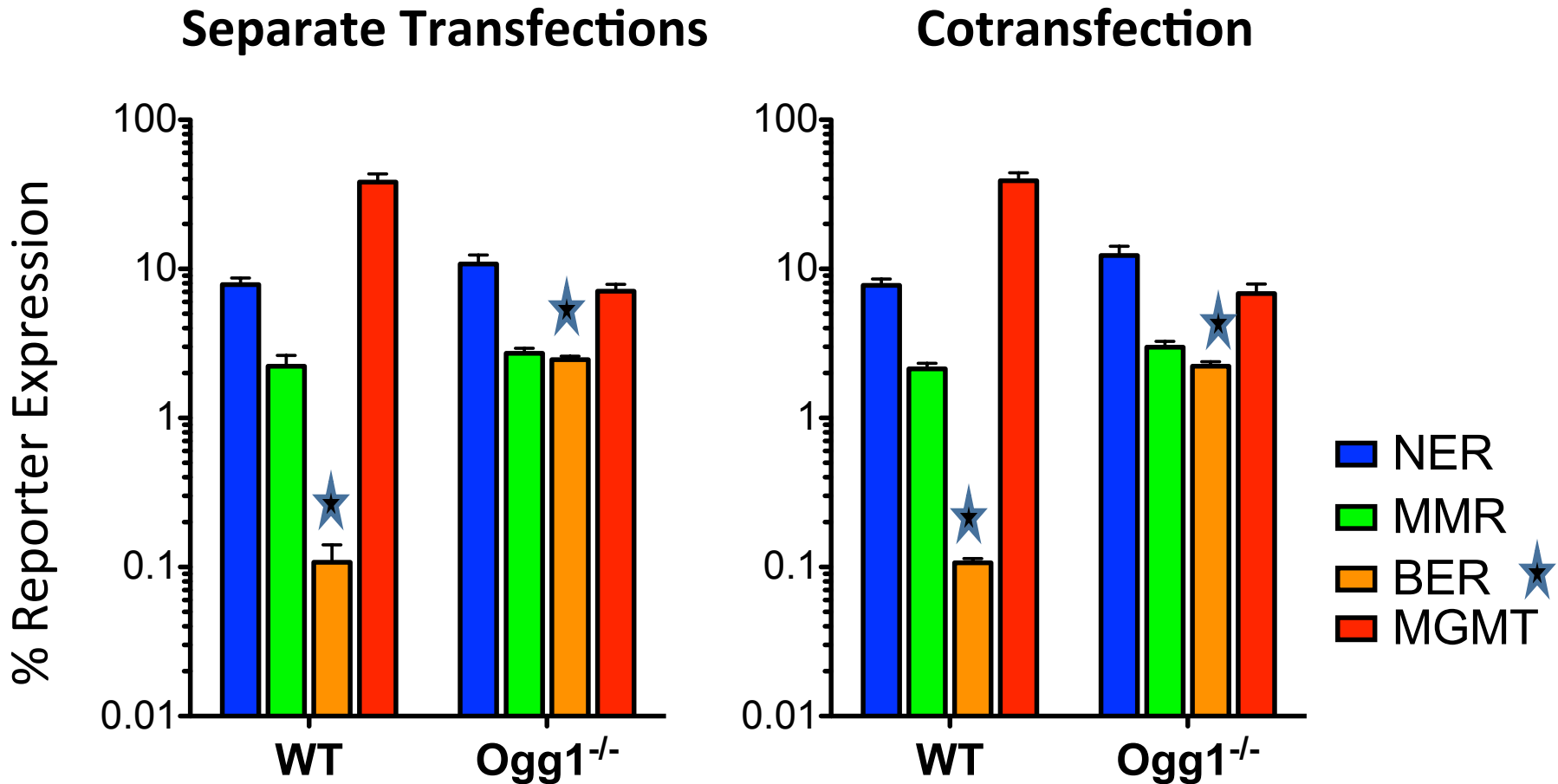
# Can We Measure DRC in 4 pathways in a single assay: Example 1



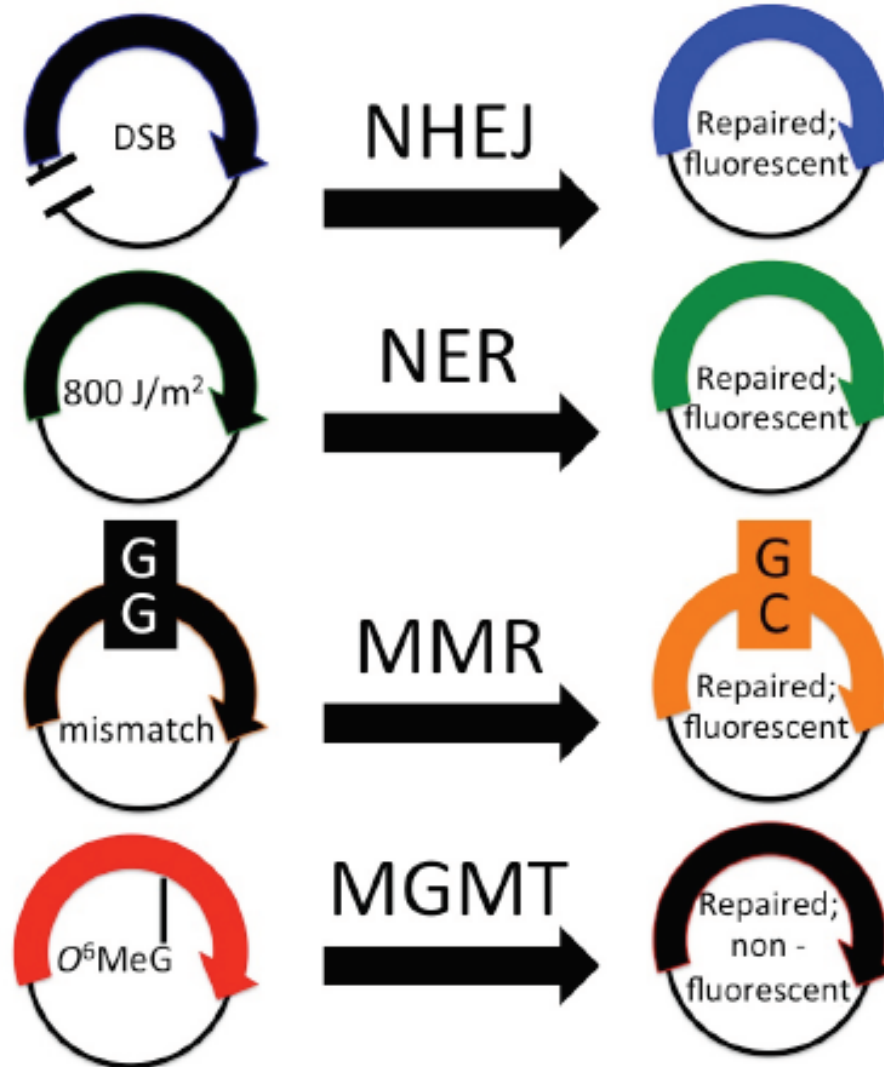
# Can We Measure DRC in 4 pathways in a single assay: Example 1



# Can We Measure DRC in 4 pathways in a single assay: Example 1

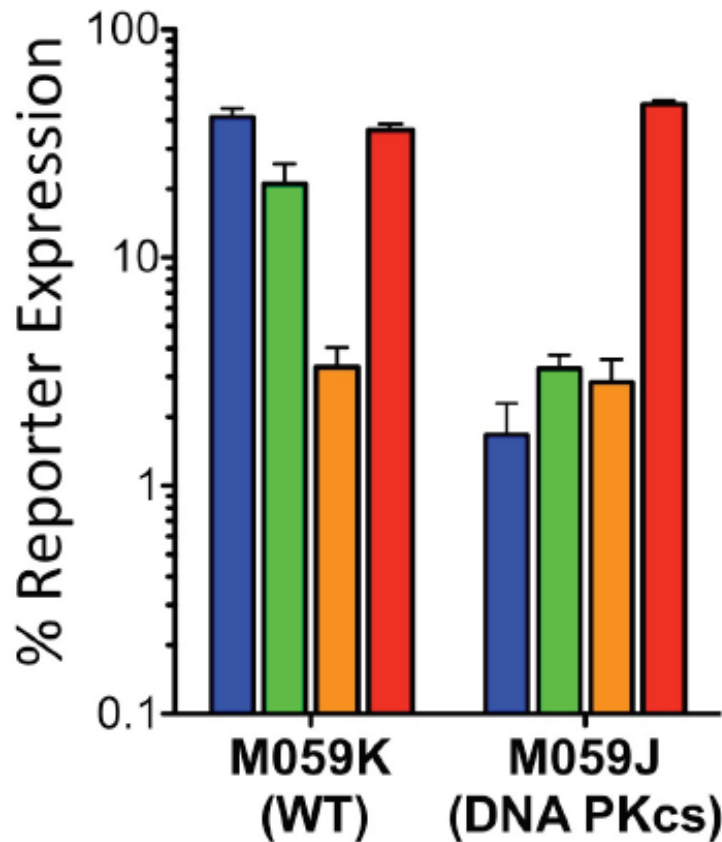


# Can We Measure DRC in 4 pathways in a single assay: Example 2

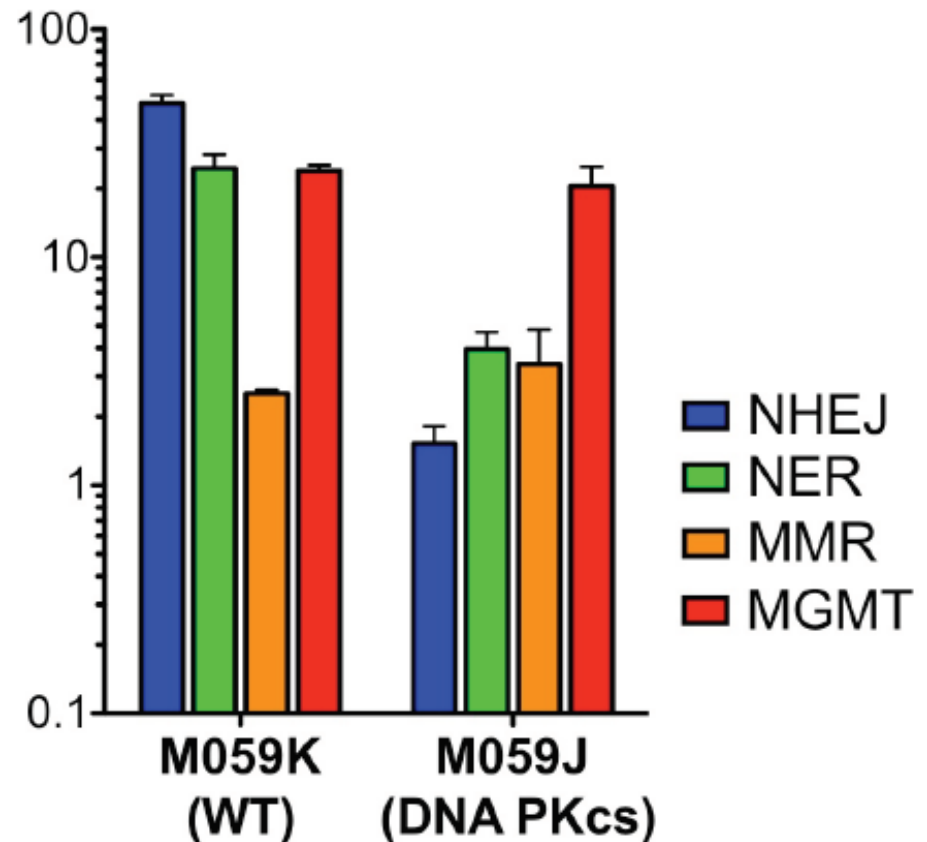


# Can We Measure DRC in 4 pathways in a single assay: Example 2

## b Separate



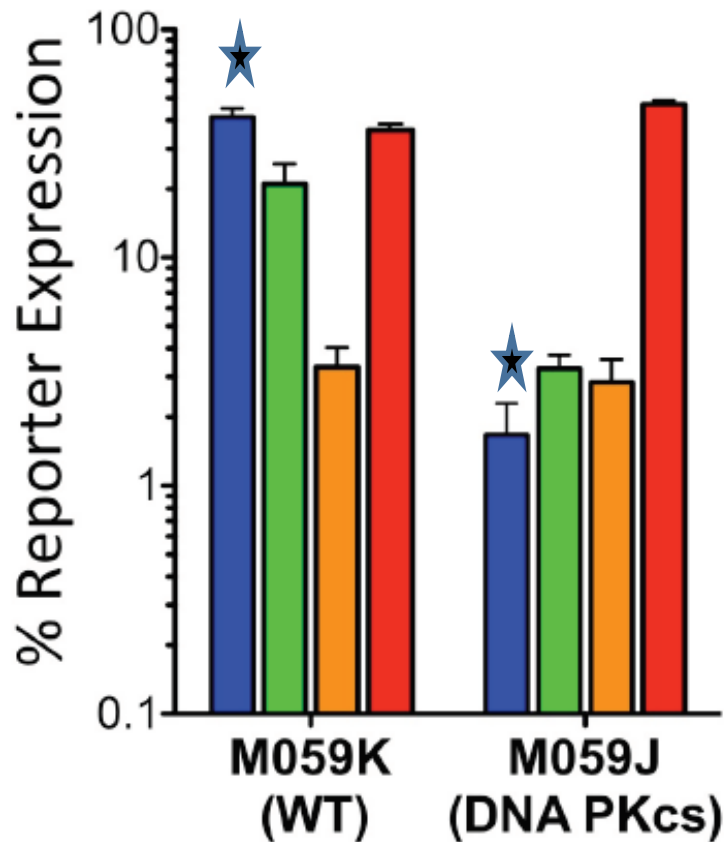
## c Together



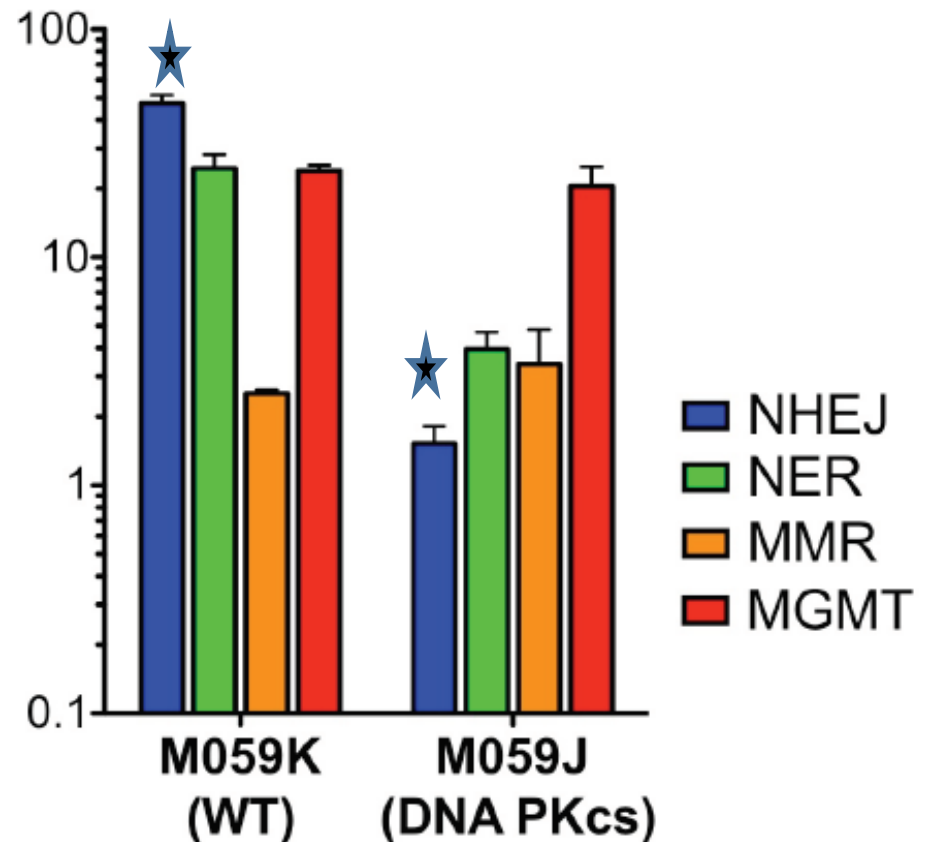


# Can We Measure DRC in 4 pathways in a single assay: Example 2

## b Separate

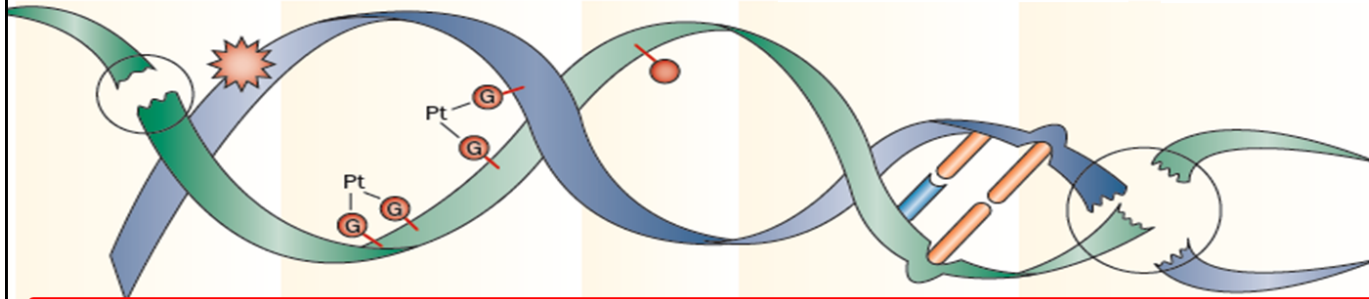


## c Together



# DNA Repair Capacity - Multiplexed

NATURE REVIEWS | CLINICAL ONCOLOGY VOLUME 9 | MARCH 2012



Single-strand break  
Single-base damage

Bulky lesions  
Crosslinks

O<sup>6</sup>MeG

Mismatch

Double-strand break

**BER**

**NER**

**DR**

**MMR**

**HR**

**NHEJ**

OGG1  
PARP1 PARP2

XPD, XPC  
DDB1/XPE  
RNA pol β  
CSA, CSB

AGT

MSH2/MSH6  
MLH1/PMS2

ATM  
MRN complex

KU70, KU80

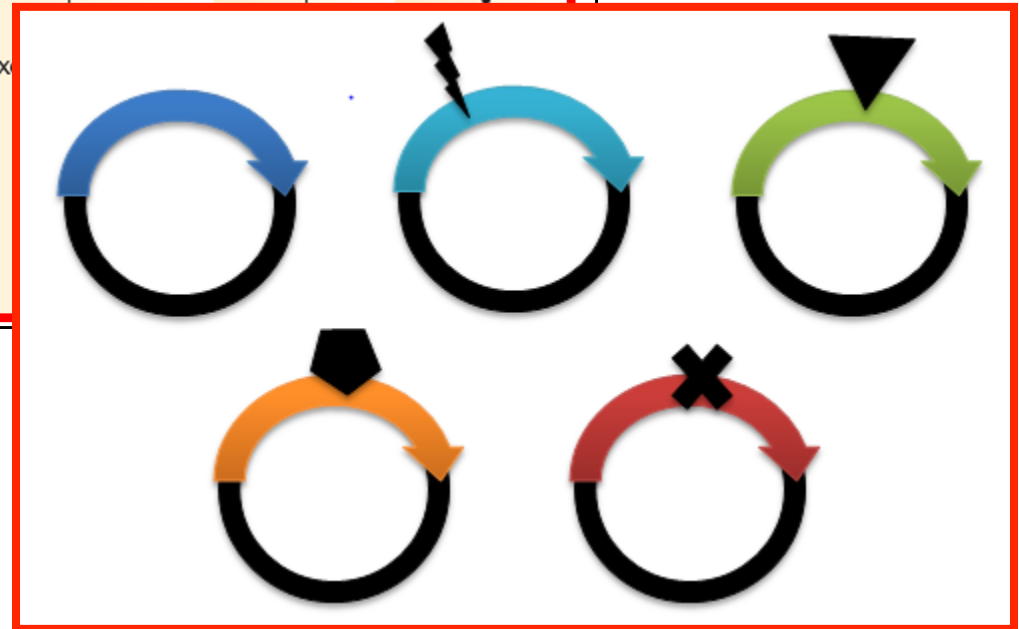
XRCC1

ERCC1/XPF

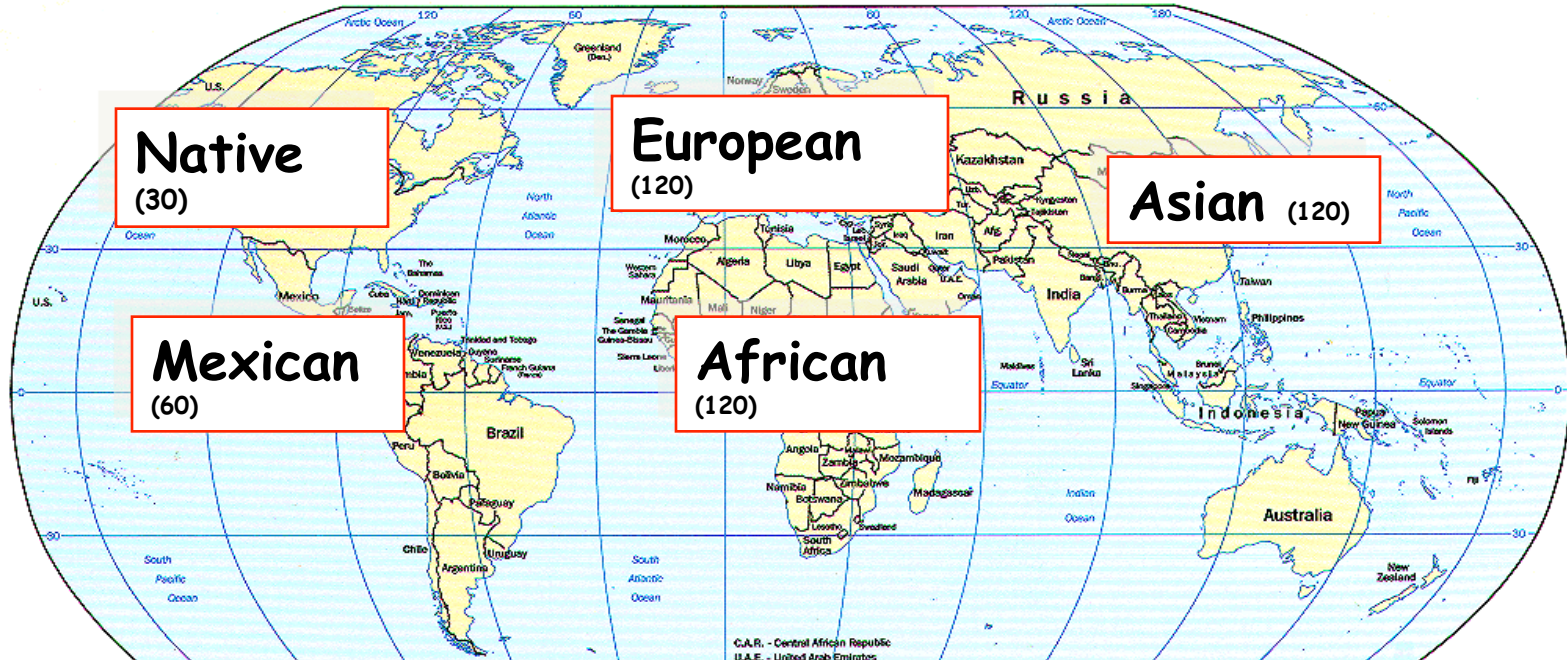
EX

Pol β  
PCNA  
FEN 1  
Ligase III

PCNA  
Pol δ  
Pol ε  
Ligase I



# Coriell Lymphoblastoid Cell line collection derived from ethnically diverse HEALTHY humans

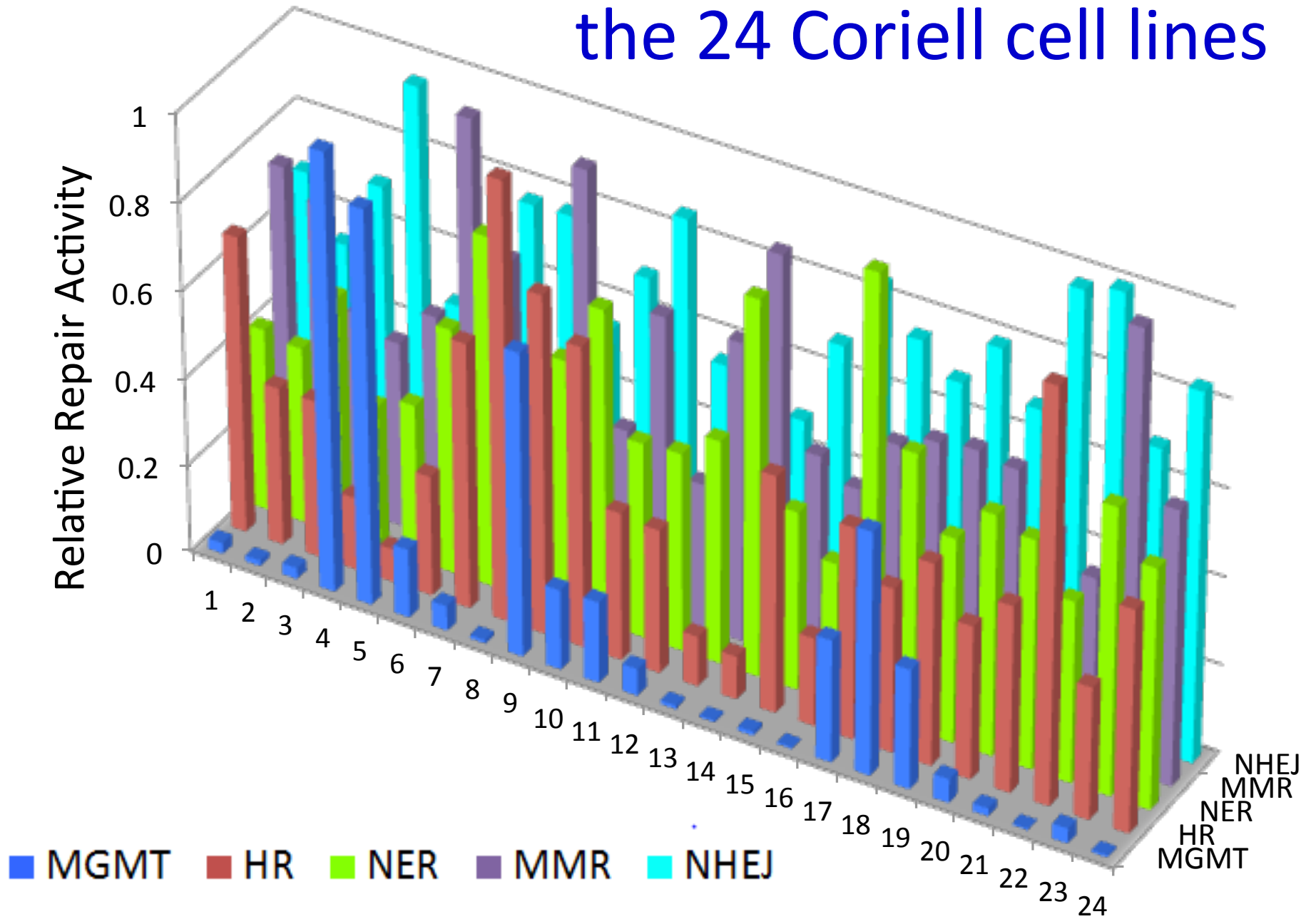


**450 healthy unrelated US residents with ancestry from around the globe**

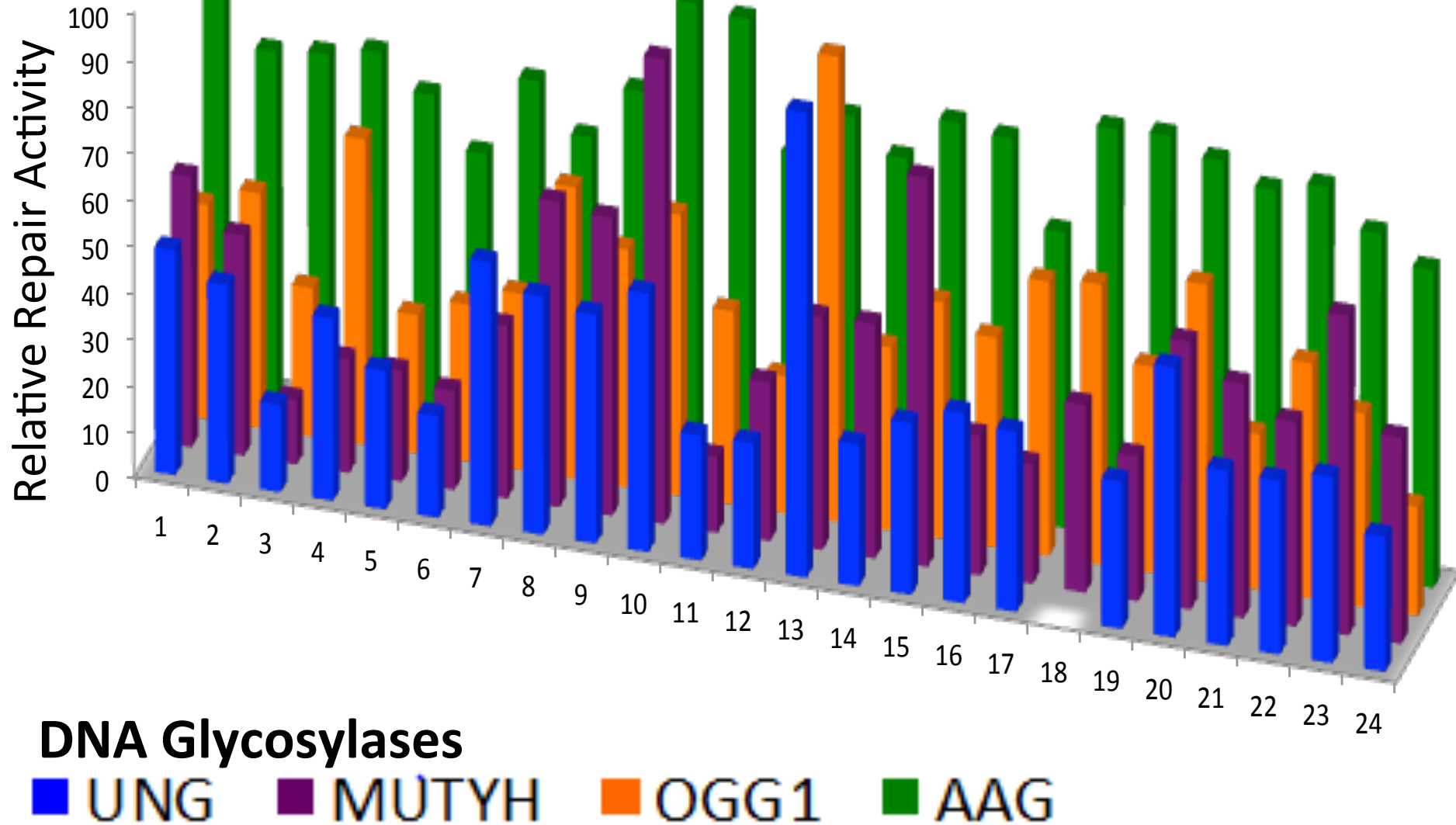
Nested subsets: 90, 44, **24**, 8

Ethical reasons: no medical, phenotypic, or ethnic information is provided

# DNA Repair Capacity in 5 pathways for the 24 Coriell cell lines



# DNA Repair Capacity for BER initiated by 4 different DNA Glycosylases in the 24 Coriell cell lines

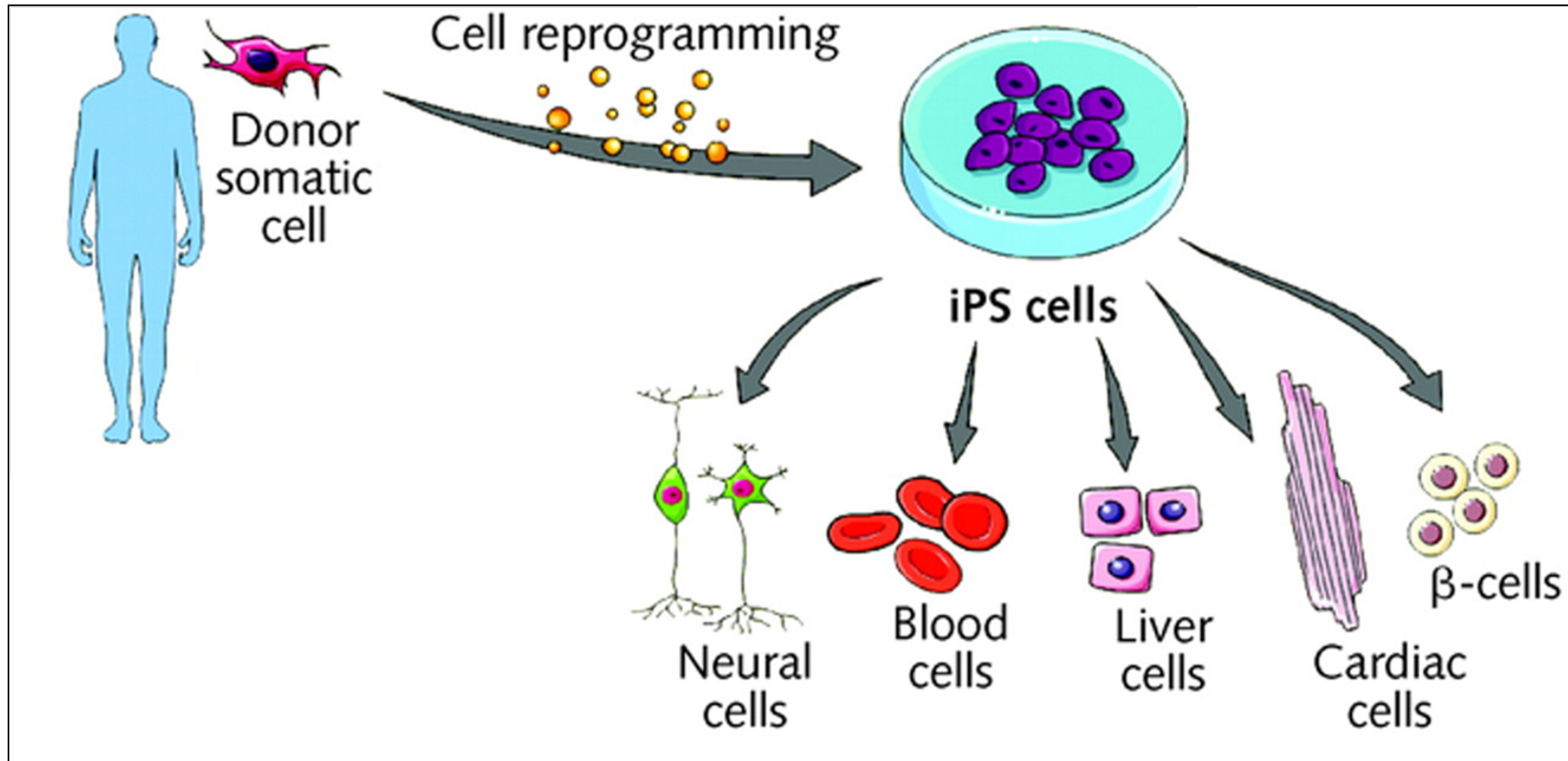


# Ongoing and Future plans post-Pioneer

- Mathematical Modeling testing predictive value of FM-HCR data.
- Exploring inter-individual variations DRC in human lymphocytes.
- DRC in glioblastoma tumor cells predicting chemo/radiotherapy outcome (with Jann Sarkaria, Mayo Clinic).
- RNAseq to overcome limitations using only 5 fluorescent reporters.

- Clinical, Basic & Population Research grants under review.
- Determine whether global (DRC) correlates to:
  - (i) disease susceptibility
  - (ii) efficacy of chemo/radiotherapy
  - (iii) chemo/radiotherapy side effects
  - (iv) degenerative diseases.

Ultimately want to measure DRC for every major DNA repair activity in many different cell types – ideally derived from each individual



# The Pioneer Team



Isaac (Alex)  
Chaim



Dr. Zachary Nagel



Patrizia  
Mazzucato



Carrie  
Margulies



Dr. Anwaar  
Ahmad



Siobhan  
McRee



Anthony  
Forget



**NIEHS**  
National Institute of  
Environmental Health Sciences