## M1D2: Perform protein purification protocol

- 1. Prelab discussion
- 2. Purify MAX-6xHis protein
- 3. Electrophorese confirmation digest



"Don't pick it up," I say, and he says, "It's just a *plasmid*, what harm could it do?" Well just look at him now...who knows *what* protein he's expressing!

#### Overview of Mod 1 experiments:



# What is our protein of interest?

- MAX functions as a transcription factor
  - Forms homodimers and heterodimers
  - Dimerizes with Myc, which is an oncogenic transcription factor
  - Homodimers and heterodimers compete for binding at promoters to provide regulatory system of target genes



#### Closer look at pET28a\_MAX-6xHis



- 6xHis
- *lac* operator
- lacl
- T7 promoter
- T7 terminator
- RBS

#### Overview of protein expression system

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## T7 RNA polymerase transcribes MAX-6xHis



E. coli BL21

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pET28a\_MAX-6xHis

#### Lacl repressor blocks transcription at *lac* operator



pET28a\_MAX-6xHis

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## IPTG 'induces' MAX-6xHis expression



pET28a\_MAX-6xHis

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## How did we induce protein expression?



# How will you purify MAX-6xHis?

- First, need to lyse cells to release proteins
  - B-PER (Bacterial Protein Extraction Reagent):
  - Lysonase:

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• Proteinase inhibitor:



# 6xHis tag binds to Ni<sup>2+</sup> resin / column





- Ni<sup>2+</sup> chelated onto agarose resin via nitrilotriacetic acid (NTA) ligand
- His tag chelates to Ni<sup>2+</sup> causing protein to 'stick' to resin / column

# Non-specific binders washed from Ni<sup>2+</sup> resin / column using imidazole



- Low concentration of imidazole included in wash buffer
- Imidazole competes for binding to Ni<sup>2+</sup> resin
  - Low affinity binders / non-specific binders are outcompeted and released from the resin

## Imidazole used to elute protein from column



- Elution buffer contains higher concentration of imidazole compared to wash buffer
- Increased concentration allows imidazole to out-compete 6xHis for binding to Ni<sup>2+</sup> resin

### Demonstration of protein purification



## Wrap-up of confirmation digest



• How do you visualize migration through the gel?

• How do you visualize DNA bands in the gel?

# For today...

- Start protein purification protocol
- Complete gel electrophoresis during lysis incubation
- Be sure to clearly label all tubes containing protein purification aliquots!

# For M1D3...

- Draft a figure of your confirmation digest results for your Data summary
  - All figures should include a TITLE and CAPTION

# Notes on figure making:

- Image should not be the entire page
  - Only needs to be large enough to be clear
- Title should be conclusive
  - Don't include what you did, rather include what you found / discovered
- Caption should not include methods details
  - Define abbreviations, symbols, etc.



Figure X: Title is the take-home message of the experimental data.

Caption includes all of the details necessary to understand the data presented in the figure...not methods!!