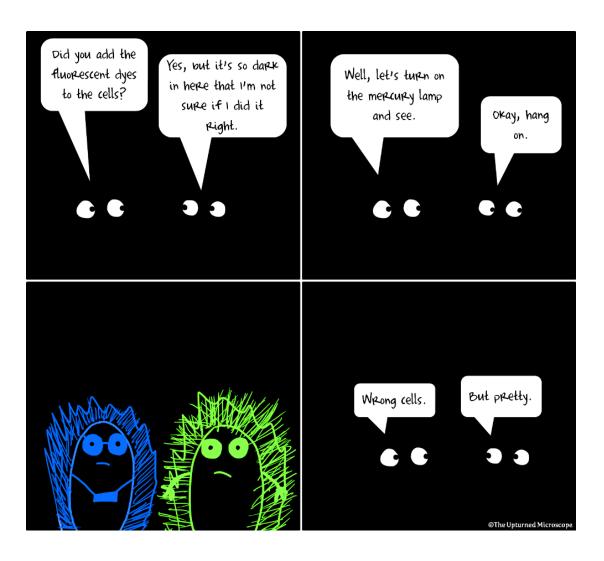
M2D5: Immunofluorescence staining

- 1. Prelab
- 2. Fix yeast
- 3. Antibody staining of Fet4_mutant

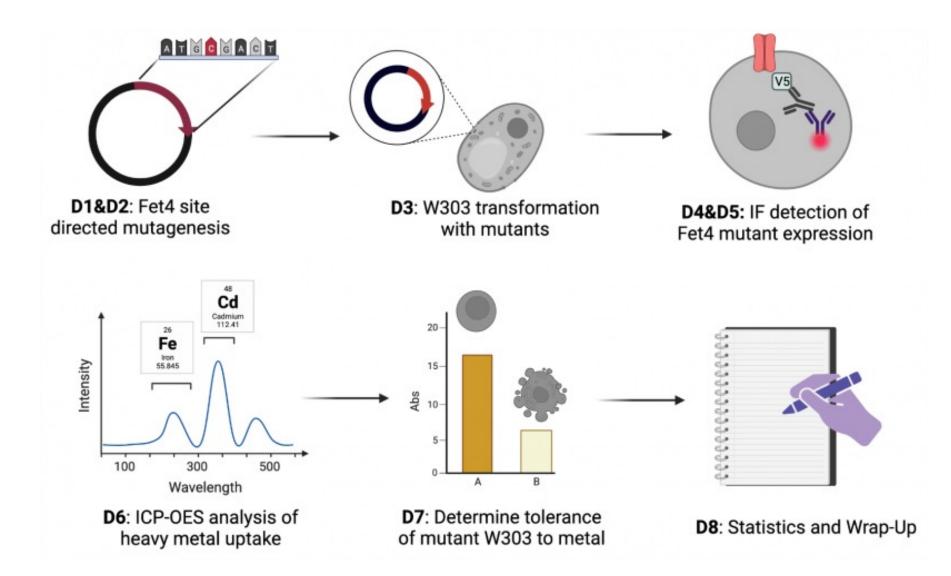


Overview of Mod 2 experiments

Last lab:

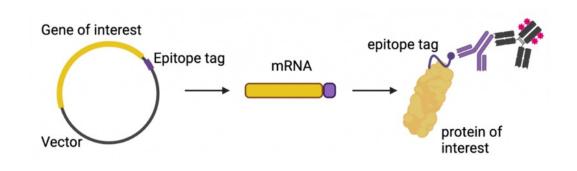
This lab:

Next lab:

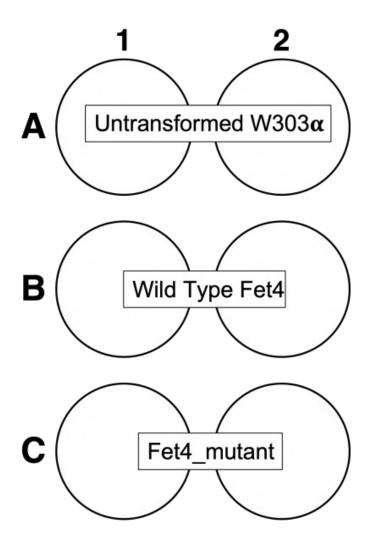


Using immunofluorescence: Expression of Fet4_mutant in yeast

• Yeast cells transformed to express Fet4_mutant protein



- Why untransformed cells?
- Why wild-type Fet4?



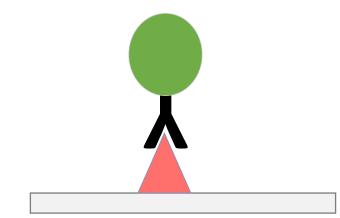
Using immunofluorescence: Identification of Fet4_mutant expression



V5

DAPI

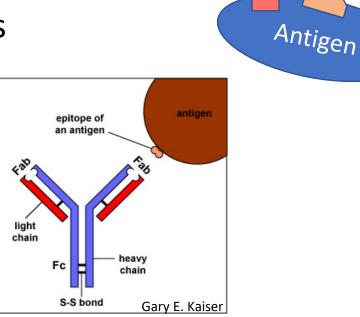
Merge



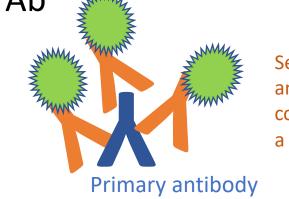
protein of interest	🔺 V5
primary antibody	k mouse anti-V5
secondary antibody	none
Fluorophore (conjugated to secondary antibody) exc./ em. wavelengths	488/525 nm

Considerations for using antibodies in the lab

- Antibodies bind to specific epitopes on antigens
 - Antigens may have multiple epitopes
- Primary antibody recognizes the antigen
 - Specific protein sequence
 - Specific conformation of protein
 - Specific state of protein (i.e. phosphorylation)



- Secondary Ab recognizes the species of the primary Ab
 - Often conjugated to tag for visualization
 - Enzyme or fluorophore
 - Amplifies signal through multiple bindings
 - Consider sample species when choosing antibodies!



Secondary antibody conjugated to a fluorophore

Epițopes

Polyclonal vs. monoclonal antibodies

Polyclonal

- How it's made: animal (often rabbit) immunized with antigen of interest then antibodies collected from blood sera and affinity purified
- Advantages:
 - Less expensive and faster to produce than monoclonal
 - Multiple antibodies in one polyclonal mixture can increase antigen recognition by binding multiple epitopes
 - Especially useful for proteins with low expression
- Disadvantages:
 - Variability from lot to lot

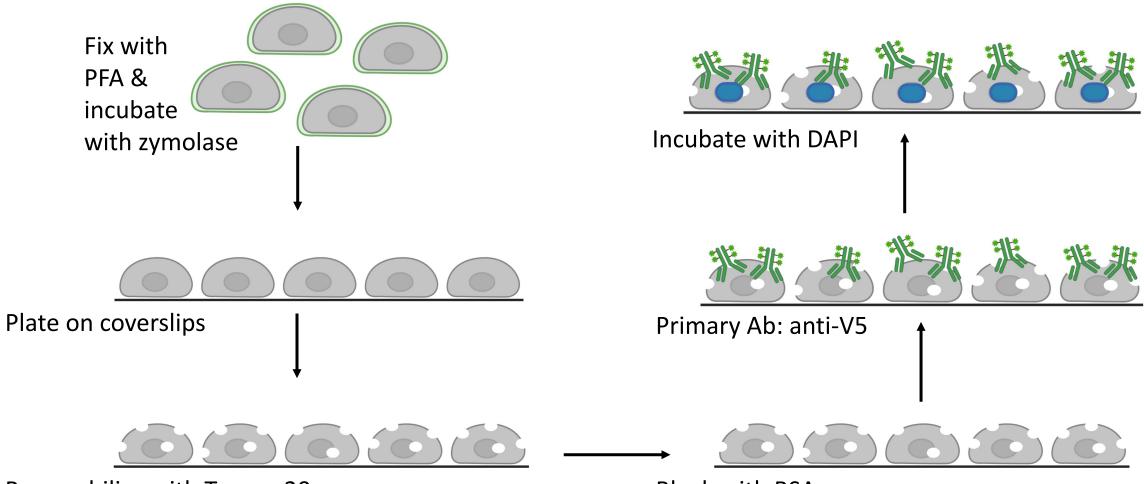


<u>Monoclonal</u>

- How it's made: animal (usually mouse) immunized with antigen of interest then B cells from spleen are harvested and fused with myeloma cells to create hybridoma cell line that will continually produce single antibody clone
- Advantages:
 - Very consistent
 - Binds single epitope (can also be disadvantage)
- Disadvantages:
 - More expensive and requires animal sacrifice



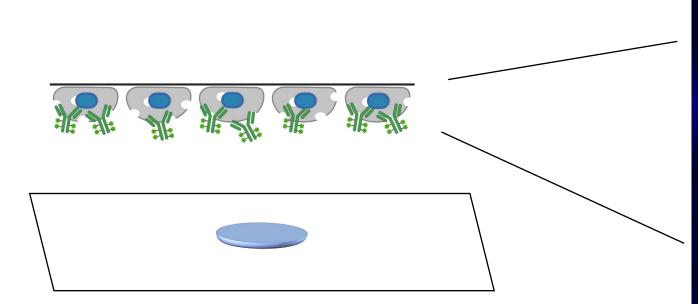
Using immunofluorescence (IF) in yeast: steps in protocol



Permeabilize with Tween-20

Block with BSA

Finish IF by mounting coverslips on slides



Mount coverslip on glass slide with mounting media

Blue= DAPI Green= antibody staining

For today:

- 1. Fix yeast samples
- 2. Perform IF
 - 1. Downtime: Look at new alignments on Dropbox that show the mutations
- 3. Mount coverslips on slides for imaging

For M2D6

Write methods for M2D2-M2D5
*** Individual Assignment ***